INTRODUCTION

The stock fluctuations of small pelagic species (e.g. sardine and anchovy) are often attributed to the relative success of recruitment. The variability of this success takes place in early life stages through the action of environmental factors. Indeed, physical processes (enrichment, concentration and reten-
tion) play a key role in this success, which combines to yield favourable reproductive habitats for small pelagic fishes and also other species (Agostini and Bakun, 2002). While *Engraulis encrasicolus* spawning of the Iberian Peninsula have been extensively studied and related to the main productive events in the different areas, the information is very scarce for the northern African waters. The variability of the anchovy recruitment in the Bay of Biscay has been mainly explained by upwelling intensity (Borja et al., 1998). The enrichment assured by Rivers Rhone and Ebro and the development of the deep fluorescence maximum influence northwestern Mediterranean anchovy (García and Palomera, 1996; Olivar et al., 2001; Sabatés et al., 2001; Lloret et al., 2004).

Despite the importance of the small pelagic resources in Tunisian waters, as in other North African Mediterranean areas information on early life stages is very scarce. In Tunisian waters, the first survey to study the abundance and distribution of anchovy eggs was carried out in 1972 by Ktari-Chakroun (1979), while the second one was carried out in 2000 (Zarrad, 2001). These surveys were limited in space and/or time and did not cover either the whole spawning area or the whole spawning period. To overcome this lack of information, a new research program, “ESSATEL”, was launched to study early life stages of small pelagic fishes in the gulf of Tunis in relation to environmental factors.

The Tunis Gulf is located in the central Mediterranean Sea. Near this gulf are the Sardinia Channel in the northeast, and the Strait of Sicily in the northwest, where the water exchanges between western and eastern Mediterranean basins occurs. In the Strait of Sicily, surface Modified Atlantic Water (MAW) with low salinity (37.03 to 37.15; Sammari et al., 1999) flows into the eastern Mediterranean, while the Levantine Intermediate Water (LIW) enters the western Mediterranean. The Tunis Gulf receives the outflow of the most important River in Tunisia: the Majreda River situated on the western coast of the Gulf.

According to Agostini and Bakun (2002), the prevailing westerly winds produce coastal upwelling off the south coasts of Sicily and Sardinia and downwelling occurs along the eastern coast of Tunisia. The Ekman transport field carries water from the upwelling (enrichment) zone towards the downwelling (concentration) zone along the east and north coast of Tunisia. The topography of the Tunisian coast creates protected areas (gulfs of Tunis and Hammamet), which also support concentration, accumulation of chlorophyll and retention areas for pelagic eggs and larvae, such as those of anchovy, by providing shelter from surface currents and wind mixing.

Among the small pelagic species, anchovy (*Engraulis encrasicolus*) represents 6% (6000 t) of small pelagic exploitable biomass (100000 t) in Tunisia (Ben Abdallah and Gaamour, 2004). This species is considered under-exploited in Tunisian waters, with a current landing of the order of 380 t per year (DGPA, 1999-2004). Influenced by its economic importance in European markets, a revision of the Tunisian fisheries policy was carried out in order to improve the exploitation of this resource. However, the sustainability of the resource must rely on appropriate management measures based on sound scientific advice arising from research of the whole life cycle (egg, larval development, juvenile and adult).

This paper presents the first information on the seasonal horizontal distribution and the levels of abundance of anchovy eggs and larvae in the Gulf of Tunis and their relationship with sea surface temperature, salinity, chlorophyll *a* and depth.

**MATERIAL AND METHODS**

Four multidisciplinary surveys were carried out on board the R/V Hannibal, one in each season: in summer (from 26 to 30 August 2002), in autumn (from 21 to 25 October 2002), in winter (from 13 to 17 February 2003) and in spring (from 11 to 15 April 2003). Samples were taken on a regular grid of 27 stations (4.8 miles in east-west and 6 miles in north-south direction) covering the whole Gulf of Tunis. Two additional stations were located near the mouths of the Rivers Majreda and Meliane (Fig. 1).

Temperature and salinity were measured at the surface for each station by means of a WTW conductivity meter. Seawater samples for chlorophyll *a* analysis were obtained with a Niskin bottle at the surface. These samples were filtered through a Whatman GF/C glass fibre, which was frozen at −20°C. Chlorophyll *a* was determined using the conventional technique proposed by Lorenzen (1967). After 90% acetone extraction, chlorophyll *a* was measured colorimetrically using a Jenway model 6405UV visible spectrophotometer.
Plankton samples were taken by oblique tows with a 60 cm mouth diameter bongo gear, fitted with nets of 335 µm mesh size. The vessel speed was 3 knots. Maximum sampled depth was 100 m, wherever possible, or 5 m above the bottom in shallower waters. A HydroBios flowmeter was fitted in the net mouth to determine the volume of filtered water. Following each tow, the plankton samples were preserved in 4% formaldehyde solution buffered with borax to maintain pH around 8.2. Subsequently, anchovy eggs and larvae were sorted. Larval standard length (SL) was measured to the nearest 0.1 mm using a stereomicroscope fitted with a micrometer. Length frequency distributions were presented as mean abundance of positive stations.

Numbers of eggs and larvae at each station were standardised to the number beneath a 10 m² area of sea surface using the computed volume of water filtered and depth sampled (Smith and Richardson, 1977). Maps of environmental parameters and anchovy egg and larva abundance distributions were made by means of Surfer Golden Software Inc., applying the kriging interpolation method. Correlation coefficient analysis was performed.

Fig. 1. – Study area and sampling stations.

Fig. 2. – Seasonal spatial distribution of sea surface temperature (°C) in the Gulf of Tunis.
between egg and larva abundance and environmental parameters using Statistica Software (StatSoft Inc.) at significance levels of $p<0.05$.

**RESULTS**

**Hydrography**

In summer, the sea surface temperature (SST) ranged between 25.1 and 26.2°C, with an average of 25.5°C. A patch of warmer waters was detected near Cape Sidi Bou Saïd (Fig. 2 and Table 1). In autumn, surface waters were colder, with an average of 21.8°C, and their horizontal distribution showed more heterogeneity, with a difference between maximum and minimum values of 2.6°C. The patch of relatively colder water (20°C) was found near Cape Sidi Bou Saïd and the patch of warmer water (22.6°C) was observed to the southwest of Zembra Island. In winter, surface waters were much colder, with an average of 13.4°C, which represents 8.4°C less than in autumn. Major horizontal differences were detected, with a difference of 4.2°C between colder waters (10.5°C), observed in the south-east of River Majreda mouth, and warmer waters, up to 14.7°C, located in the north of the study area. In spring there was a mean temperature increase of 2.7°C, reaching an average of 16.1°C. In this season warmer waters were located north of the River Majreda mouth and the colder waters were around Cape Bon. There was some positive gradient from the east to the west.

Summer and autumn sea surface salinity (SSS) distributions showed great similarity, with average SSS values of 37.3 and 37.0, respectively (Fig. 3).

### Table 1. – Range, mean and standard deviation (SD) of temperature, salinity and chlorophyll a by season in the Gulf of Tunis (from summer 2002 to spring 2003).

<table>
<thead>
<tr>
<th>Season</th>
<th>Temperature (°C)</th>
<th>Salinity SSS</th>
<th>Chlorophyll-a (mg/m³)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>range</td>
<td>mean ±SD</td>
<td>range</td>
</tr>
<tr>
<td>Summer</td>
<td>25.1-26.2</td>
<td>25.5 ±0.22</td>
<td>37.1-37.8</td>
</tr>
<tr>
<td>Autumn</td>
<td>20.0-22.6</td>
<td>21.8 ±0.45</td>
<td>37.0-37.5</td>
</tr>
<tr>
<td>Winter</td>
<td>10.5-14.7</td>
<td>13.4 ±0.93</td>
<td>22.3-36.4</td>
</tr>
<tr>
<td>Spring</td>
<td>15.0-18.0</td>
<td>16.1 ±0.86</td>
<td>29.5-36.8</td>
</tr>
</tbody>
</table>

**Fig. 3.** – Seasonal spatial distribution of sea surface salinity in the Gulf of Tunis.
For both seasons, weak horizontal variations, with maximum differences of 0.7 in summer and 0.5 in autumn, were observed. In both seasons, saltier waters were located in the inner part of the Gulf as a result of the high evaporation of the coastal waters. The relatively low salinity detected in the northernmost area seems to be related to the Atlantic waters. In winter, the lowest salinity value (22.3) was detected to the south of the River Majreda mouth. A positive gradient was found with a west-east fairly similar, since the lowest salinity value (29.5) was also observed to the south of the River Majreda mouth. However, in the eastern part there was low heterogeneity and no clear gradient. The average SSS was 36.2.

Low concentrations of chlorophyll $a$ at surface were recorded in summer (0.41 mg/m$^3$) and autumn (0.42 mg/m$^3$). For both seasons, relatively high concentrations were found south of the Bay of Tunis, particularly near Soliman lagoon, and the concentrations decreases gradually from south to north (Fig. 4).

In winter and spring, chlorophyll $a$ concentrations increased significantly to average values of 1.11 and 1.26 mg/m$^3$, respectively. In winter, high concentrations (over 1.7 mg/m$^3$) were observed near the River Majreda mouth, Cape Bon and the Soliman lagoon. In spring, even higher chlorophyll-$a$ concentrations (over 2.7 mg/m$^3$) were found once more near the River Majreda mouth and off Cape Bon, but there were high concentrations in the centre of the Gulf and also to the north of Zembra Island.

### Egg and larval distributions

Anchovy eggs were found in all seasons (Fig. 5 and Table 2). However, abundance observed in winter was very low. The highest anchovy egg abundance corresponded to spring, followed by summer (average abundances of 223 and 162 eggs/10 m$^2$, respectively). The area southwest of Zembra Island showed a peak of anchovy egg abundance during spring and summer (1255 and 1038 eggs/10 m$^2$, respectively). In summer, no eggs were found on the eastern coasts. However, a relatively high abundance (134 eggs/10 m$^2$) was found off the River Majreda mouth and off Cape Bon.

### Table 2. – Range, mean and standard deviation (SD) of anchovy egg and larva abundance by season in the Gulf of Tunis (from summer 2002 to spring 2003).

<table>
<thead>
<tr>
<th>Season</th>
<th>Eggs/10 m$^2$ range</th>
<th>mean</th>
<th>±SD</th>
<th>Larvae/10 m$^2$ range</th>
<th>mean</th>
<th>±SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Summer</td>
<td>0-1038</td>
<td>121</td>
<td>179</td>
<td>0-445</td>
<td>88</td>
<td>129</td>
</tr>
<tr>
<td>Autumn</td>
<td>0-314</td>
<td>47</td>
<td>80</td>
<td>0-62</td>
<td>13</td>
<td>21</td>
</tr>
<tr>
<td>Winter</td>
<td>0-84</td>
<td>7</td>
<td>18</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Spring</td>
<td>0-1306</td>
<td>223</td>
<td>394</td>
<td>0-16</td>
<td>1</td>
<td>3</td>
</tr>
</tbody>
</table>

Fig. 4. – Seasonal spatial distribution of chlorophyll-$a$ (mg/m$^3$) in the Gulf of Tunis.
Meliane mouth (less than 20 m in depth). The anchovy egg distribution during autumn showed a wider distribution. Major egg densities were mainly located in the north of the study area, centred in the Gulf of Tunis, showing a gradual decrease in egg abundance towards the coasts.

Fig. 5. – Seasonal and spatial distributions of anchovy eggs in the Gulf of Tunis.

Fig. 6. – Seasonal and spatial distributions of anchovy larvae in the Gulf of Tunis.

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Larvae were more abundant in summer (88 larvae/10 m$^2$), whereas they were absent in winter and scarce in spring (1 larva/10 m$^2$) (Fig. 6). In summer, they were concentrated (more than 350 larvae/10 m$^2$) in the centre of the Gulf. However, an isolated small patch (18 larvae/10 m$^2$) was identified near the River Meliane mouth. As in summer, autumn showed the highest larval abundance (60 larvae/10 m$^2$) in the centre of the Gulf. A decreasing abundance trend from this point was observed towards the western and eastern coasts. However, no larvae were observed at the southernmost end of the Gulf or in the north off Cape Bon. In spring, larvae were only found to the south and west of Zembra Island with a maximum of 16 larvae/10 m$^2$.

Length (SL) sizes of larvae ranged between 2 and 14 mm in summer and 2 and 15 mm in autumn (Fig. 7). Length frequency distributions of larvae showed that in summer the 4 mm size class was dominant (43%) and more than 91% of larvae were smaller than 6 mm. In autumn, two size classes (4 mm and 5 mm) dominated, each one representing 27%. More than 44% of the larvae were larger than 6 mm.

**Eggs, larvae and environmental conditions**

In summer, autumn and spring abundances of anchovy eggs and larvae were correlated positively and significantly, the coefficients being 0.41 (p=0.047), 0.65 (p=0.000) and 0.42 (p=0.026), respectively.

There were positive and significant correlations of the depth with the horizontal distribution of anchovy eggs in all seasons, and with larvae in summer and autumn (Table 3). Egg abundances showed a negative and significant correlation with salinity in autumn ($r=-0.44$; $p=0.016$) and a positive correlation with temperature in winter ($r=0.42$; $p=0.022$). A negative and significant correlation was observed between larval abundance and salinity in autumn ($r=-0.46$; $p=0.013$).

The SST is the main factor controlling the anchovy spawning intensity. The highest egg abundances were found when the SST increased (spring and summer). In contrast, when the SST decreased the egg abundances were low (autumn and winter). For the larvae, the highest abundance was observed when the SST reached its maximum (summer).

In spring, the area of maximum abundance of anchovy eggs coincided with the maximum primary production (chlorophyll $a$).

**DISCUSSION**

The SST, the SSS and the chlorophyll $a$ distributions in the study area allow us to distinguish two periods: summer-autumn and winter-spring. The
first period was characterised by warmer (25.5 and 21.8°C), saltier waters (37.3 and 37.2) and lower chlorophyll a concentrations (0.41 and 0.43 mg/m³). The second period was characterised by colder (13.4 and 16.1°C), lower-salinity waters (35.1 and 36.2) and higher chlorophyll a levels (1.11 and 1.27 mg/m³). Moreover, the first period was characterised by horizontal homogeneity and the second by a considerable heterogeneity. The differences observed between these two periods seem to be a result of the Majreda outflow, the air temperature and wind intensity variations.

Despite this temporal variation in the environmental parameters of the Gulf of Tunis, anchovy eggs were caught in all the seasons. However, the main spawning period was spring-summer, when SST ranged between 16.1 and 25.5°C between the end and the beginning of the two periods indicated above. Our results agree in general with those exposed by Demir (1965) and Arbault and Boutin (1977), who stated that the most favourable temperature range for anchovy spawning was 15 to 25°C and 15.8 to 24°C, respectively. Moreover, anchovy spawning can occur with the minimum temperature of 13°C (Dekhnik, 1954; Demir, 1959; Palomera, 1992), as occurred in the present winter survey (13.4°C). Moreover, in the northern Adriatic sea, Zavodnik (1970) caught anchovy eggs in February with water temperatures of 11.6°C. Our results also agree with those found by Motos et al. (1996) and Palomera (1992), in the sense that the anchovy spawning peak takes place in the transition period between winter (cold and vertically homogenous waters) and summer (warm and stratified waters), the abundance of anchovy eggs and larvae being positively correlated to the warming rate of surface waters.

The spawning period of anchovy shows some differences among Mediterranean regions, but in all cases Engraulis encrasicolus showed the maximum spawning intensity in spring and/or in summer, as can be seen in Table 4. The wide spawning period observed in the present study could be explained by the temporal difference in the spawning of different cohorts. Cendrero et al. (1981) and Santiago and Eltink (1988) demonstrated that the young specimens (1 year) spawn in June and the older (2 years and more) spawn early, in April and May. Moreover, García and Palomera (1996) concluded that the spawning period of anchovy is longer in southern latitudes of the Mediterranean (seven months) than in northern ones (five months).

The spawning grounds of Engraulis encrasicolus in nearby areas are usually related to places where enrichment processes occur: fronts, upwellings, convergence areas and river runoff (Motos et al., 1996; Palomera, 1992; Bakun and Agostini, 2001; Agostini and Bakun, 2002; García-Lafuente et al., 2002; Cuttita et al., 2004). In our case the period with maximum abundance of anchovy eggs (spring) coincided with the maximum primary production (highest measured concentrations of chlorophyll a).

In many places anchovy spawning is associated with river outflows (Richardson, 1981; Lima and Castello, 1995), as occurs in the northwestern Mediterranean with the Rivers Ebro and Rhone (Aldebert and Tournier, 1971; Palomera, 1992; Palomera and Sabatés, 1990) and in the Bay of Biscay with the Loire and the Garonne (Arbault and Boutin, 1977). Demir (1965) indicated a wider tolerance for this species, between 9 and 37. Taking into account these references, it seems that the Gulf of Tunis could be a favourable spawning habitat for anchovy thanks to the enrichment produced by the outflow of the Rivers Majreda and Meliane. However, in our case during the main spawning period (spring) eggs and larvae were absent or scarce in the areas with lower salinities (29.5), and there was a negative correlation between egg distri-

<table>
<thead>
<tr>
<th>Zone</th>
<th>T optimal (°C)</th>
<th>Spawning season</th>
<th>Spawning peak</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gulf of Biscay</td>
<td>14-18</td>
<td>March-August</td>
<td>May-June</td>
<td>Motos et al. (1996)</td>
</tr>
<tr>
<td>Mira estuary (Portugal)</td>
<td>15.5-19.5</td>
<td>Mars-November</td>
<td>April</td>
<td>Ré (1996)</td>
</tr>
<tr>
<td>NW Mediterranean</td>
<td>15-20</td>
<td>April-October</td>
<td>May-July</td>
<td>Palomera, 1992</td>
</tr>
<tr>
<td>NW Mediterranean</td>
<td>15-22</td>
<td>April-October</td>
<td>June-August</td>
<td>García and Palomera (1996)</td>
</tr>
<tr>
<td>Alboran Sea</td>
<td>19-23</td>
<td>Mars-November</td>
<td>August</td>
<td>Rodríguez and Rubin (1986); Rodríguez (1990)</td>
</tr>
<tr>
<td>Ligurian and N Tyrharian Seas</td>
<td>-</td>
<td>Mai-September</td>
<td>July</td>
<td>Albertelli et al. (1988)</td>
</tr>
<tr>
<td>Adriatic Sea</td>
<td>17-22</td>
<td>April-October</td>
<td>May-August</td>
<td>Regner (1996)</td>
</tr>
<tr>
<td>Gulf of Tunis</td>
<td>16-25</td>
<td>February-October</td>
<td>April-August</td>
<td>Present study</td>
</tr>
</tbody>
</table>
The limit of the shelf, where a deep chlorophyll abundances of anchovy eggs are always found at the river is great over the continental shelf, the high although the productivity induced by the outflow of detected at the surface. A similar situation were there were some other enrichment processes not offshore. It is likely that in the north of the gulf occur in spring, the main spawning areas were far Majreda outflow-induced enrichment processes that of the Gulf, beyond the 75 m isobath. Despite the Gulf of Tunis seem to be related to the central part are highly prone to predation (Alheit, 1987).

The maximum larval abundances were found in the deeper (more than 50 m) areas of the north part of the Gulf. The main distribution zone for Engraulis encrasicolus eggs in the northwestern Mediterranean has been reported at depths higher than 100 m (Palomera and Sabatés, 1990; Olivar et al., 2001).

The spawning areas and larval abundance in the Gulf of Tunis seem to be related to the central part of the Gulf, beyond the 75 m isobath. Despite the Majreda outflow-induced enrichment processes that occur in spring, the main spawning areas were far offshore. It is likely that in the north of the gulf there were some other enrichment processes not detected at the surface. A similar situation were found in the area of the Ebro river delta, where although the productivity induced by the outflow of the river is great over the continental shelf, the high abundances of anchovy eggs are always found at the limit of the shelf, where a deep chlorophyll a maximum is always observed between 50 and 70 m depth in the summer months (Palomera, 1992, Olivar et al., 2001).

This work is a first step that needs to be followed by others in order to obtain a better understanding of recruitment fluctuations of Tunisian anchovy populations. Indeed, it is necessary to analyse the relationships between abundances of early life stages of anchovy and more environmental parameters, to extend the study area in order to cover the whole spawning area of the stock, and to carry out multi-year studies.

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