

4. Algae-microbiome interactions: integrative overview from biology to chemistry

4PO.1

A COMMON GARDEN EXPERIMENT TO TEST RECOVERY OF MICROBIAL BIODIVERSITY FROM CLONAL BLADES OF *PORPHYRA UMBILICALIS* (STRAIN P.U.M.1)

Charlotte TC Quigley¹ (charlotte.quigley@maine.edu), Inara R Mendonca¹ (inara.regina@gmail.com) and Hilary G Morrison² (hmorrison1981@gmail.com)

¹School of Marine Sciences, University of Maine, Orono 04469, United States and ²Josephine Bay Paul Center, Marine Biological Laboratory, Woods Hole 02543, United States

We used a clonal strain (P.um.1) of *Porphyra umbilicalis* in common garden experiments to evaluate recovery of the microbiome using different methods of tissue stabilization. Blades of identical size were grown in the laboratory in the same vessel and then separated into two culture chambers, each containing a set of three blades (bubbled vigorously at 12°C, 40 μmol photons m² s⁻¹, 12:12 (L:D), West-McBride enriched sterile seawater, changed weekly). After 3 weeks, adjacent pieces from the margin and subtending vegetative tissue were processed to produce 6 replicates of each preparative treatment: 1) flash freeze, store at -80 C, and grind finely with a mortar and pestle, 2) flash freeze, lyophilize, and powder (via GenoGrinder, 2 min, 1600 strokes/min, zirconium beads), 3) dry with silica gel, and powder, or 4) dry with silica gel, lyophilize, and powder. The first processing method was repeated across two batches of replicates (n=6 blades/batch). Additionally, treatment effects on the rhizoid microbiome were assessed. DNA was extracted with a Qiagen DNeasy Plant MiniKit, and the v6 region of the 16S rDNA was used to compare the effect of different processing methods on the bacteria that were recovered, while PNA probes reduced chloroplast and mitochondrial contamination. While small variations in biodiversity were found between samples, differences were not attributable to the chamber location or method of post-harvest tissue stabilization. Because of earlier antibiotic treatments, P.um.1 has much lower bacterial biodiversity compared to wild blades, but blades grow and reproduce normally. Proteobacteria and Bacteroidetes comprised most of the microbial biodiversity, but five phyla were represented. (Supported by NSF 1442231, 1442106, and 0741907).

4PO.2

TRANSCRIPTOME ANALYSIS ON THE GREEN SPOT DISEASE RESPONSIVE GENES IN CULTIVATED *PYROPIA* SPP.

Soo Hyun Im¹ (shi3938@kongju.ac.kr), Jong Won Han¹ (jwhan@kongju.ac.kr), Tatyana A Klochkova¹ (tanya@kongju.ac.kr) and Gwang Hoon Kim² (ghkim@kongju.ac.kr)

¹Biology, Kongju National University, Kongju 314-701, South Korea and ²Biology, Kongju National University, Kongju 314-701, South Korea

Green spot disease (GSD) is one of the most serious diseases in *Pyropia* sea farms of Korea, which cause multi-million dollar of loss every year. More than half of commercial *Pyropia* product collected in the market showed a trace of GSD infection. The causative agent of this disease has recently been identified as a virus. To understand innate immunity of *Pyropia* to this virus infection we performed transcriptome analysis using 454 pyrosequencing and microarray analysis. About 220million reads, 850Mbp of genomic information were analyzed using transcriptome data collected from 454 pyrosequencing with CLC Genomic Workbench. About 23,000 probes for microarray analysis has been designed using 454 pyrosequencing data. Results showed 1,237 genes responsive to virus infection. Among them RNA-binding proteins were most highly upregulated after the infection. RNA-binding proteins (RBPs) are known to inhibit RNA virus infection in eukaryotic cells. The expression of these genes was analysed during the progress of the infection using qPCR method. Results indicated that the causative agent of GSD is a RNA virus and *Pyropia* cells are using RBPS to inhibit the progress of the infection.

4PO.3

IDENTIFYING PROKARYOTIC CONSORTIA THAT LIVE IN CLOSE INTERACTIONS WITH ALGAE

Assia Saltykova¹ (assia.saltykova@ugent.be), Sergio Pulido-Tamayo² (spulido99@gmail.com), Marie Pažoutová³ (marie.pazoutova@gmail.com), Stefan A Rensing⁴ (stefan.rensing@biologie.uni-marburg.de),

Tomoaki Nishiyama⁵ (tomoakin@kenroku.kanazawa-u.ac.jp), Yves Van de Peer⁶ (yves.vandepeer@psb.vib-ugent.be), Kathleen Marchal² (kathleen.marchal@intec.ugent.be) and Stephane Rombauts⁶ (strom@psb.vib-ugent.be)

¹Department of Information Technology (UGent), Department of Plant Biotechnology and Genetics (UGent), Department of Plant Systems Biology (VIB), Ghent University, Flanders Institute for Biotechnology, Ghent 9000 Ghent, Belgium; ²Department of Information Technology (UGent), Department of Plant Biotechnology and Bioinformatics (UGent), Department of Microbial and Molecular Systems (K.U. Leuven), Ghent University, University of Leuven, Ghent, Leuven 9000 Ghent, 3000 Leuven, Belgium; ³Department of Botany (JU), Institute of Parasitology (BC AS CR), University of South Bohemia, Biology Centre AS CR, Centre for Polar Ecology, Ceske Budejovice 37005, Czech Republic; ⁴Faculty of Biology, University of Marburg, Marburg D-35043, Germany; ⁵Advanced Science Research Center, Kanazawa University, Kanazawa 920-0934, Japan and ⁶Department of Plant Biotechnology and Genetics (UGent), Department of Plant Systems Biology (VIB), Ghent University, Flanders Institute for Biotechnology, Ghent 9000, Belgium

In whole-genome shotgun sequencing projects of marine algae bacterial populations are frequently being picked up. Because the algae are cultured in a defined environment before sequencing, the presence of such bacterial residuals potentially points towards symbiotic or opportunistic interactions between the bacterial communities and the algae. We have applied modern metagenomic tools to analyse the bacteria found within genome sequencing data of *Chara vulgaris*, *Prasiola crispa* and *Ostreococcus tauri*. This approach allowed us to identify and compare bacterial species associated with the algae, and reconstruct nearly full colateral bacterial genomes from every analysed low-complexity metagenomic dataset.

4PO.4

SUBSURFACE ASSOCIATIONS OF ACARYOCHLORIS-RELATED PICOCYANOBACTERIA WITH OIL-UTILIZING BACTERIA IN THE ARABIAN GULF WATER BODY: PROMISING CONSORTIA IN OIL SEDIMENT BIOREMEDIATION

Dhia A Al-Bader¹ (dhiaalbader@yahoo.com), Mohamad Eliyas¹ (mohelijyas@gamil.com) and Samir Radwan² (samir.radwan@ku.edu.kw)

¹Biology, Kuwait University, Kuwait 16030 safat, Kuwait and ²Biology, Kuwait, Kuwait 16030 safat, Kuwait

Two picocyanobacterial strains related to *Acaryochloris* were isolated from the Arabian Gulf, 3 m below the water surface, one from the north shore and the other from the south shore of Kuwait. Both strains were morphologically, ultrastructurally, and albeit to a less extent, phylogenetically similar to *Acaryochloris*. However, both isolates lacked chlorophyll d and produced instead chlorophyll a, as the major photosynthetic pigment. Both picocyanobacterial isolates were associated with oil-utilizing bacteria in the magnitude of 10^5 cells g^{-1} . According to their 16S rRNA gene sequences, bacteria associated with the isolate from the north were affiliated to *Paenibacillus* sp., *Bacillus pumilus*, and *Marinobacter aquaeolei*, but those associated with the isolate from the south were affiliated to *Bacillus asahii* and *Alcanivorax jadensis*. These bacterial differences were probably due to environmental variations. In batch cultures, the bacterial consortia in the nonaxenic biomass as well as the pure bacterial isolates effectively consumed crude oil and pure aliphatic and aromatic hydrocarbons, including very high-molecular-weight compounds. Water and diethylether extracts from the phototrophic biomass enhanced growth of individual bacterial isolates and their hydrocarbon-consumption potential in batch cultures. It was concluded that these consortia could be promising in bioremediation of hydrocarbon pollutants, especially heavy sediments in the marine ecosystem.

4PO.5

USING NEXT GENERATION SEQUENCING TO UNDERSTAND MICROBIOMES AND SEASCAPE GENOMICS OF RED SEAWEEDS

Juliet Brodie¹ (j.brodie@nhm.ac.uk), Chris Yesson² (chris.yesson@ioz.ac.uk), Gary Barker³ (gary.barker@bristol.ac.uk), Marian Yallop³ (Marian.Yallop@bristol.ac.uk) and Chris Williamson⁴ (c.williamson@nhm.ac.uk)

¹Life Sciences, The Natural History Museum, London SW75BD, United Kingdom; ²Institute of Zoology, Zoological Society of London, London NW1 4RY, United Kingdom; ³School of Biological Sciences, Bristol BS8 1TQ, United Kingdom and ⁴Life Sciences Department, London SW7 5BD, United Kingdom

Seaweeds are host to a wide range of epibiotic and endobiotic prokaryotic and eukaryotic organisms and the relationship between these organisms can be symbiotic, opportunistic or pathological. At present,