

Potential for adaptive evolution at species range margins: contrasting interactions between red coral populations and their environment in a changing ocean

Jean-Baptiste Ledoux^{1,2,3}, Didier Aurelle³, Nathaniel Bensoussan⁴, Christian Marschal³, Jean-Pierre Féral³ & Joaquim Garrabou^{3,2}

¹CIMAR/CIIMAR, Centro Interdisciplinar de Investigação Marinha e Ambiental, Universidade do Porto, Rua dos Bragas 177, 4050-123 Porto, Portugal

²Institut de Ciències del Mar CSIC, Passeig Marítim de la Barceloneta 37-49, Barcelona, Spain

³Aix Marseille Université, CNRS, IRD, Avignon Université, IMBE, UMR 7263, 13397 Marseille, France

⁴IPSO FACTO, SCOPARL, Pôle Océanologie et Limnologie, 37 rue Saint-Sebastien, F-13006 Marseille, France

Keywords

Common garden, *Corallium rubrum*, deep refugia hypothesis, marginal populations, phenotypic buffering, potential for local adaptation, reciprocal transplants.

Correspondence

Jean-Baptiste Ledoux, CIMAR/CIIMAR, Rua dos Bragas, n. 289 4050-123 Porto, Portugal.
Tel: (+351) 22 340 18 00;
Fax: (+351) 22 339 06 08;
E-mail: jbaptiste.ledoux@gmail.com

Present address

Jean-Baptiste Ledoux, CIMAR/CIIMAR, Centro Interdisciplinar de Investigação Marinha e Ambiental, Universidade do Porto, Rua dos Bragas 177, 4050-123 Porto, Portugal
Departamento de Biologia, Faculdade de Ciências, Universidade do Porto, Rua do Campo Alegre, 4169-007, Porto, Portugal

Funding Information

J-B. Ledoux is supported by a post-doctoral grant SFRH/BPD/74400/2010 from Fundação para a Ciência e Tecnologia (FCT). This study was funded by the "Provence-Alpes-Côte d'Azur" region (J-B. Ledoux Doctoral Fellowship) and by the IFB Foundation (Institut Français de la Biodiversité), the environmental department of TOTAL-FINA-ELF group, an "Action Concertée Initiative" (ACI) program "Jeunes chercheurs 2003" (number JC5360) from the French Research Ministry and the MEDCHANGE project funded by the Agence Nationale pour la Recherche (ANR).

Received: 28 May 2014; Revised: 21 October 2014; Accepted: 22 October 2014

Ecology and Evolution 2015; 5(6): 1178–1192

doi: 10.1002/ece3.1324

1178

Abstract

Studying population-by-environment interactions (PEIs) at species range margins offers the opportunity to characterize the responses of populations facing an extreme regime of selection, as expected due to global change. Nevertheless, the importance of these marginal populations as putative reservoirs of adaptive genetic variation has scarcely been considered in conservation biology. This is particularly true in marine ecosystems for which the deep refugia hypothesis proposes that disturbed shallow and marginal populations of a given species can be replenished by mesophotic ones. This hypothesis therefore assumes that identical PEIs exist between populations, neglecting the potential for adaptation at species range margins. Here, we combine reciprocal transplant and common garden experiments with population genetics analyses to decipher the PEIs in the red coral, *Corallium rubrum*. Our analyses reveal partially contrasting PEIs between shallow and mesophotic populations separated by approximately one hundred meters, suggesting that red coral populations may potentially be locally adapted to their environment. Based on the effective population size and connectivity analyses, we posit that genetic drift may be more important than gene flow in the adaptation of the red coral. We further investigate how adaptive divergence could impact population viability in the context of warming and demonstrate differential phenotypic buffering capacities against thermal stress. Our study questions the relevance of the deep refugia hypothesis and highlights the conservation value of marginal populations as a putative reservoir of adaptive genetic polymorphism.

Introduction

In the human-dominated Earth ecosystem, global change raises the question of the capacities of populations to cope with the increase of selective pressures (Hoffmann and Sgro 2011). Disentangling the impacts of adaptive processes in population-by-environment interactions (PEIs) is therefore one of the main challenges in evolutionary and conservation biology (Allendorf et al. 2010). In this context, local adaptation, adaptive phenotypic plasticity, and particularly, phenotypic buffering have received increasing attention from both theoretical (Bürger and Lynch 1995) and empirical (Antoniazza et al. 2010) perspectives (Reusch and Wood 2007; Reusch 2014). We define local adaptation as the patterns and processes driven by divergent selection leading to specific PEIs, such that locally adapted individuals exhibit higher relative fitness in their habitat than foreign individuals (Kawecki and Ebert 2004). Adaptive phenotypic plasticity is the capacity for an individual's genotype to produce distinct phenotypes, increasing its relative fitness in response to environmental variations (Via et al. 1995; Pigliucci 2005). The particular case of phenotypic buffering relies on the organism's ability to maintain its physiological functions despite extreme stress (Reusch 2014). Although the interactions between local adaptation and adaptive plasticity are still debated, these two processes are not mutually exclusive (Ghalambor et al. 2007; Baythavong and Stanton 2010). Local adaptation to environmental heterogeneity can rely on adaptive plasticity when heterogeneity is predictable in time over the organism's lifespan (e.g., seasonal heterogeneity; Scheiner 1993) or occurs within populations (i.e., at a scale smaller than gene flow; Baythavong 2011). In addition to the central role of selection, adaptive processes are therefore intrinsically linked to other evolutionary forces, such as gene flow and genetic drift (Alleaume-Benharira et al. 2006; Bridle et al. 2010).

In the study of adaptive processes, the importance of populations at species range margins has recently been emphasized (Hampe and Petit 2005; Kawecki 2008). Compared to core populations, marginal populations have generally been considered to be demographically and genetically impaired because of the combination of low-quality habitats, low densities, and geographic isolation (Lawton 1993). However, some studies have challenged this center-periphery hypothesis (Sagarin and Gaines 2002) and proposed that marginal populations can be locally adapted to the atypical ecological characteristics of their environment (e.g., Garner et al. 2004; Orizaola et al. 2010). These two hypotheses have different implications for conservation biologists. The former implies that marginal populations are vulnerable *per se*, as they are maintained via gene flow from core populations (Hoff-

mann and Blows 1994). On the contrary, the latter suggests that marginal populations offer the opportunity to characterize the responses of populations facing a new regime of selection and can thus serve as a reservoir of adaptive genetic variation (Hampe and Petit 2005; Kawecki 2008).

The controversial status of marginal populations can deeply impact the development of conservation policies. Focusing on coastal ecosystems, the selective pressures linked to global change (e.g., thermal stress) are stronger in the shallow (0–30 m depth) compared to the mesophotic (30–100 m depth) zone, resulting in an intense decline of shallow compared to mesophotic populations within species (Bongaerts et al. 2010). The deep refugia hypothesis (Glynn 1996) proposes that mesophotic populations could act as refugia and as a source of recruits for shallow populations. Accounting for the bathymetric range of various marine species, we rephrase this hypothesis as the question of whether marginal populations can be replenished by more central ones. Thus far, the deep refugia hypothesis has been tested by searching for neutral connectivity between different bathymetric zones (Bongaerts et al. 2010; Costantini et al. 2011; van Oppen et al. 2011; Brazeau et al. 2013), thus assuming identical PEIs between depths. This does not account for the local adaptations observed from marine taxa (Conover et al. 2006; Marshall et al. 2010; Sanford and Kelly 2010) and at the edge of species range (Kawecki 2008). Therefore, characterization of the PEIs at range margins in marine species is a necessary task to enhance our understanding of the consequences of global change on marine biodiversity and to propose relevant conservation measures.

The Mediterranean red coral, *Corallium rubrum*, is an engineer species in the coralligenous community, which is one of the richest but also most threatened Mediterranean communities (Ballesteros 2006). This species is a sessile, aposymbiotic, and long-lived cnidarian that exhibits slow population dynamics (Garrabou and Harmelin 2002) and late sexual maturity (at 10 years of age; Torrents et al. 2005). *Corallium rubrum* displays a fragmented distribution centered on the western Mediterranean basin and dwells in heterogeneous habitats, as illustrated by its bathymetric distribution, ranging from depths of 5 to 800 m (Costantini et al. 2010). Red coral populations are genetically structured at the scale of tens of meters (Ledoux et al. 2010a; Aurelle et al. 2011; Aurelle and Ledoux 2013), in accordance with the restricted effective dispersal of this species (Ledoux et al. 2010b).

The red coral is overharvested for use in jewelry (Bruckner 2009). Recently, the populations of the north-western Mediterranean dwelling at the upper edge of this species' bathymetric distribution (from 5 to 30–50 m, depending on the region) have been impacted by large-

scale mass mortality events (Garrabou *et al.* 2009) linked to positive thermal anomalies in the water column (Bensoussan *et al.* 2010). These mass mortalities events impacted a large number of species. Within each impacted species, populations were differentially affected by the mortalities (Garrabou *et al.* 2009). In the red coral, significant differences in the rates of tissue necrosis have been observed between populations and between individuals within populations (Garrabou *et al.* 2009). *In situ* temperature records (Bensoussan *et al.* 2010) and *in aquaria* experiments (Torrents *et al.* 2008) have demonstrated the central role of temperature in these events and revealed that impacted populations are close to the limit of their ecological tolerance.

In this study, we focus on red coral populations dwelling at different depths (shallow = 20 m and mesophotic = 40 m) within the upper edge of this species' bathymetric range. We combine reciprocal transplants and *in situ* common garden experiments, comparisons between estimates of neutral genetic (F_{ST} ; Wright 1943) and phenotypic (P_{ST} ; Leinonen *et al.* 2006) differentiation between populations and population genetics analyses to: (1) test the potential for local adaptation at fine spatial scales; (2) reveal the relative impacts of evolutionary forces on the observed patterns; and (3) evaluate the mitigating effects of adaptive processes *in situ* in the context of global change.

Materials and Methods

Study system

We conducted the study at two localities: Riou Island (RI) (43°10'22.11"N; 5°23'21.93"E) and Palazzu Island (PZ) (42°22'48.61"N; 8°32'46.87"E) (Fig. 1A). These localities were chosen because their summer thermal regimes are among the most contrasting in the northwestern Mediterranean (Bensoussan *et al.* 2010). In each locality and based on a previous population genetics study (Ledoux *et al.* 2010a), we selected two genetically differentiated red coral populations separated by 100 m, but dwelling at two different depths corresponding to the shallow and mesophotic zones (20 and 40 m designated RI-20, RI-40 and PZ-20, PZ-40). The populations are exposed to different thermal conditions. The shallow habitats (RI-20 and PZ-20) are warmer and more variable than the mesophotic habitats (RI-40 and PZ-40) (Bensoussan *et al.* 2010; Fig. 1B; Table S1a Supplementary information). The shallow populations have been impacted by mass mortality events (Garrabou *et al.* 2009). They can occasionally be exposed to temperatures above 24°C, which has been proposed as the upper thermal threshold for the red coral (Torrents *et al.* 2008). In the framework of the deep refugia hypothesis, the

mesophotic populations can therefore be considered putative sources of recruits.

Experimental design

The following experiments were designed to account for the low abundance and low population dynamics of the red coral and the limitations induced by fieldwork conducted via SCUBA diving in the mesophotic habitat (40 m).

Reciprocal transplant experiments (RTEs)

We performed two independent RTEs (Fig. 1C) at the two localities from June to November 2006, encompassing the summer period. The aim of these experiments was to test the potential for local adaptation of red coral populations. Each RTE included four different treatments: one control (colonies transplanted at their native depth) and one transplant (colonies transplanted at the foreign depth) in each of the two habitats (shallow: 20 m and mesophotic: 40 m). The apical tips of 5 to 7 cm in length (from now on "colony") of 48 individuals per population were randomly sampled via SCUBA diving and fixed underwater on experimental plates (Appendix S1). Overall, the two RTEs involved 192 colonies from four populations, which were shared among 24 plates, with three plates per treatment and eight colonies per plate.

Common garden experiment (CGE)

A CGE (Fig. 1C) was conducted on a dimly lit overhang at a depth of five meters to simulate a positive thermal anomaly *in situ* based on a previous thermal survey (Bensoussan *et al.* 2010; Fig. 1B; Table S1b). The aim of this experiment was to test for differential phenotypic buffering in populations originating from different depths. Because of the logistic constraints linked to the monitoring of the phenotypic response (see below), this CGE was only conducted in Riou. We collected 24 colonies from RI-20 and 24 colonies RI-40 that were randomly distributed among six experimental plates (three per depth of origin). The colonies were individually identified to survey their levels of tissue necrosis and survival from June to September 2006. As a control, we used the control treatments from the RTE.

Trait measurements

Based on the biological characteristics of the red coral (i.e., low dynamics, longevity, absence of symbiosis) and the depths of the experiments, which limit the fieldwork, we selected three phenotypic traits to assess the impact of environmental conditions on the colonies. We analyzed

the mean maximum growth in diameter in the RTEs and the level of tissue necrosis and the survivorship of the colonies in the CGE. These traits are a good proxy for fitness because the reproductive output in gorgonians is function of their size (Hall and Hughes 1996; Linares *et al.* 2008). Moreover, growth and survival rates are among the most commonly used fitness measures (Leimu and Fischer 2008). Quantification of the mean maximum growth in diameter during the experimental period was made possible by labeling the colonies with calcein prior to transplantation (Appendix S1). At the end of the RTEs, each colony was divided in two parts: The basal portion

was air-dried for growth analysis, and the other portion was fixed in 95° ethanol for genotyping (see below). Using stereomicroscope images obtained from sections of the colonies (Appendix S2), we computed the mean maximum growth in diameter as the mean of the 10 greatest distances between the calcein labeling and the periphery of the section. This growth analysis was performed on 71 colonies: Ten per treatment, except for the transplants from 20 m to 40 m, 40 m to 20 m, and the 40 m to 40 m controls at Palazzu in which eight, seven, and six colonies were analyzed due to labeling failure. In the CGE, we surveyed the level of tissue necrosis and the

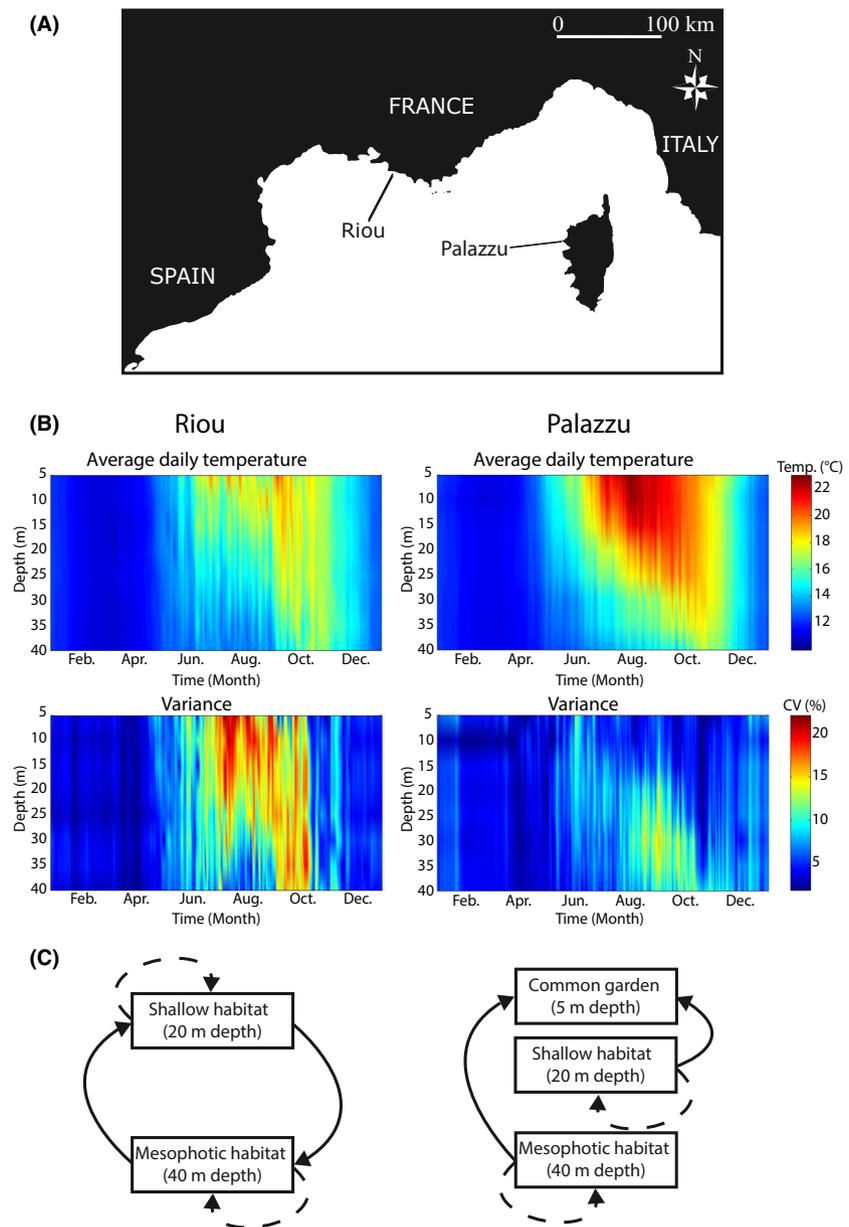


Figure 1. (A) Study sites; (B) Annual thermal regimes: average daily temperature (°C) profiles from 5 to 40 m depths and associated variances (CV in %) computed from hourly times series collected from 1999 to 2012 at Riou and from 2004 to 2011 at Palazzu; (C) Experimental protocols for the RTEs (left panel) and for the CGE (right panel). Dashed and solid arrows represent the control and transplant treatments respectively.

survivorship of the colonies ten times between June and September.

Microsatellite analysis

Of the 192 individuals utilized in the RTEs, 173 (43, 43, 42, and 45 for RI-20, RI-40, PZ-20, and PZ-40, respectively) were genotyped for 12 microsatellites loci. Due to extensive partial or total mortality of the colonies in the CGE, genotyping was not performed. PCR amplifications were conducted following Ledoux *et al.* (2010b). To avoid bias due to null alleles, we retained seven loci for statistical analyses (*Mic13*, *Mic20*, *Mic22*, *Mic24*, *Mic26*, *Mic27*, and *COR46bis*; Appendix S3).

Data analysis

Field experiments

In the RTEs, we tested the “local vs. foreign criterion” (Kawecki and Ebert 2004): Local adaptation is supported when local samples show higher fitness than foreign samples in each habitat. The mean maximum growth for each colony in each treatment was considered as the response variable. We considered the data to represent a fully two-way design of samples (replicates) with the “depth of origin” (two levels) and “treatment depth” (two levels) as fixed factors. These data were subjected to a two factorial univariate PERMANOVA test based on Euclidean distance (Anderson 2001; McArdle and Anderson 2001) and accounting for the interaction between both factors. For the CGE, the level of necrosis for each colony was the response variable, and the “depth of origin” was a fixed factor. We conducted a one-way univariate PERMANOVA test based on Euclidean distance (Anderson 2001; McArdle and Anderson 2001). Although the variables were univariate, we conducted PERMANOVA tests because the null distribution of the test statistic in PERMANOVA is produced through permutation ($n = 9999$) avoiding the normality assumptions required for parametric tests. Analyses were carried out using PRIMER v.6 (Clarke and Gorley 2006) in the PERMANOVA+ module (Anderson *et al.* 2008).

Population genetic analyses

Microsatellite characteristics, Hardy–Weinberg equilibrium, genetic diversity, and population structure

The computations conducted for microsatellite characteristics, Hardy–Weinberg equilibrium, and genetic diversity are presented in Appendix S3. Global and pairwise differentiations were computed using the θ estimator of F_{ST} (Weir and Cockerham 1984) in FREENA (Chapuis and

Estoup 2007). We tested for genotypic differentiation in all samples and in all pairs of samples using GENEPOP (Rousset 2008).

To represent the genetic distances between the samples, we produced a phenogram using POPULATIONS 1.2.30 (Langella, 2010) with the neighbor-joining algorithm (Saitou and Nei 1987) and the distance measure of Nei *et al.* (1983) (*Da*).

Impact of migration and drift in ecologically divergent populations

We estimated the level of connectivity for each population using STRUCTURE v.2.3.2 (Pritchard *et al.* 2000; Falush *et al.* 2003) by computing the mean percentage of assignment of individuals for each sample. We performed ten runs with a burn-in of 100,000, followed by 300,000 iterations with the number of genetic clusters (K) set to 4 (number of populations) using a locprior model (Hubisz *et al.* 2009), allowing for admixture and correlated allele frequencies between clusters with and without the recessive allele options (Falush *et al.* 2007). CLUMPP v.1.1 (Jakobsson and Rosenberg 2007) and DISTRUCT v.1.1 (Rosenberg 2004) were employed to average the assignment scores over the ten runs and for graphical display.

The level of genetic drift was estimated using the standard linkage disequilibrium method (Hill 1981) with Waples' (2006) correction to compute a contemporary value of the effective population size (N_e). The computations were performed with LDN_e under the random-mating model, excluding rare alleles with frequencies of less than 0.02 and using the jackknife option to estimate confidence intervals (Waples and Do 2008, 2010).

P_{ST}-F_{ST} comparisons and sensitivity analysis

Comparison of differentiation measures based on quantitative traits (Q_{ST} ; Spitze 1993) and neutral molecular markers (F_{ST}) allows the estimation of the relative impacts of neutral and selective processes on populations divergence. Divergent selection is suggested when Q_{ST} exceeds F_{ST} whereas when F_{ST} is greater than Q_{ST} , stabilizing selection drives the evolution of the considered trait. If the trait evolves neutrally, then Q_{ST} should be equal to F_{ST} (Leinonen *et al.* 2013). The computation of Q_{ST} requires knowledge on the additive genetic variance within and between populations for the considered trait (Brommer 2011). These components of genetic variance are estimated by performing multigenerational experiments in controlled environments. However, for many species, rearing individuals in the laboratory is not feasible for a variety of reasons such as late sexual maturity, as observed in the red coral (10 years; Torrents *et al.*

2005). Despite being questioned (see below), the approximation of Q_{ST} using P_{ST} , which measures phenotypic differentiation between populations, has been widely employed for studying the potential for local adaptation in the wild (Leinonen et al. 2008, 2013).

To complement the field experiments, we therefore computed P_{ST} values as $P_{ST} = c\sigma_{GB}^2 / (c\sigma_{GB}^2 + 2h^2\sigma_{GW}^2)$, where σ_{GB}^2 and σ_{GW}^2 represent the among-sample and the within-sample variance components for the considered phenotypic trait; h^2 represents the assumed additive genetic proportion of differences between individuals within populations (“narrow-sense heritability”); and c represents the proportion of the total variance presumed to occur because of additive genetic effects across populations (Brommer 2011). We first considered the null assumption that the genetic architecture of the trait remained equal across populations, with $c/h^2 = 1$ (Saether et al. 2007; Brommer 2011). For each RTE, we estimated P_{ST} between each pair of samples in each habitat using the mean maximum growth in diameter. For the CGE, P_{ST} was computed between the two samples based on the level of necrosis within the common garden. The estimators of $F_{ST}(\theta)$ and P_{ST} were compared based on their 95% confidence intervals (95% CI). The 95% CI of θ was computed using bootstraps over loci in FREENA. We estimated the 95% CI of the P_{ST} by resampling over the maximum growth of the individuals within each sample for the RTEs and over individual necrosis rates within each sample for the CGE.

P_{ST} - F_{ST} comparisons are questioned due to intrinsic characteristics of the two measures (Pujol et al. 2008; Whitlock 2008; Edelaar and Björklund 2011a). F_{ST} could be biased for highly polymorphic markers such as microsatellites (Hedrick 2005). Nevertheless, the relevance of alternative measures of population differentiation for multi-allelic markers such as G'_{ST} (Hedrick 2005) or D (Jost 2008) in P_{ST} - F_{ST} comparisons has been questioned (Leinonen et al. 2008; Edelaar and Björklund 2011a; Edelaar et al. 2011b). Accordingly and following Edelaar and Björklund (2011a), we complemented the comparisons using an allozyme-based F_{ST} between populations separated by 80 km ($F_{STall} = 0.1$; Abbiati et al. 1993). Because allozyme loci are less polymorphic than microsatellites and bearing in mind the occurrence of isolation by distance in the red coral (Ledoux et al. 2010a), we considered F_{STall} as an upper limit for the genetic differentiation between RI-20 versus RI-40 and PZ-20 versus PZ-40. On the other hand, the approximation conducted using P_{ST} instead of Q_{ST} can confound the environmental, nonadditive, and additive genetic bases of phenotypic differentiation (Pujol et al. 2008). The accuracy of this approximation is determined by the ratio between c and h^2 , which are unknown when phenotypic rather than

quantitative traits are used (Brommer 2011). Accordingly, Saether et al. (2007) and Brommer (2011) recommended testing the robustness of P_{ST} - F_{ST} comparisons with respect to the variation in c and h^2 in the conservative range of values where $c \leq h^2$ ($0 < c/h^2 \leq 1$). Considering that the P_{ST} - F_{ST} comparisons were conducted based on the results obtained for populations in the same habitat, the c value is likely to be low. We implemented this sensitivity analysis for the P_{ST} - F_{ST} comparisons that were significant under the null hypothesis (see Results). We evaluated the robustness of each comparison by estimating the c/h_{crit}^2 (i.e., the c/h^2 value beyond which P_{ST} is significantly higher than F_{ST} ; Brommer 2011) considering the upper limit of the 95% CI of θ and F_{STall} . Considering the intrinsic characteristics of the P_{ST} and the F_{ST} previously exposed, caution is necessary when interpreting P_{ST} - F_{ST} comparisons. In this context, Brommer (2011) showed that the lower the c/h_{crit}^2 value, the more robust the conclusion regarding the impact of selection on the considered trait and suggested that c/h_{crit}^2 should be lower than 0.2 to draw realistic inferences.

The computations were performed in R (<http://cran.r-project.org/>), and an example of the scripts used for the P_{ST} - F_{ST} comparisons and the sensitivity analyses is presented in Appendix S4.

Results

Reciprocal transplant experiments (RTEs)

All the colonies used in the RTEs survived without showing necrosis. At Riou, significant differences in mean maximum growth were observed considering the depth of transplantation ($P < 0.01$), but not the origin of the sample ($P = 0.09$). A significant interaction between these two factors ($P < 0.01$) was highlighted. In the two habitats, the local sample exhibited a significantly higher value than the foreign one (Table 1a; Fig. 2A) supporting the “local vs. foreign criterion”.

At Palazzu, the growth values were not significantly different according to the origin of the sample ($P = 0.49$), the transplantation depth ($P = 0.55$) or their interaction ($P = 0.62$). Therefore, we could not reject the null hypothesis of no “local vs. foreign” differences (Table 1a; Fig. 2A).

Common garden experiment (CGE)

The colonies in the control treatments did not show any necrosis. The colonies coming from RI-40 showed earlier necrosis than those from RI-20, with the first signs of necrosis being concordant with an increase in temperature beyond 24°C (Fig. 3A). At the end of the

Table 1. (a) Results of the permutational univariate analyses conducted for the RTEs; (b) results of the PERMANOVA analysis conducted for the CGE.

(a)						
Source of variation	Riou			Palazzu		
	Degrees of freedom	Mean square	Pseudo-F	Degrees of freedom	Mean square	Pseudo-F
Origin	1	1.2×10^{-3}	2.9 NS	1	1.9×10^{-4}	0.5 NS
Depth	1	9.1×10^{-3}	22.5**	1	1.5×10^{-4}	0.5 NS
Origin \times Depth	1	6.1×10^{-3}	15**	1	1.01×10^{-4}	0.6 NS
Residual	36	4.1×10^{-4}		27	4.1×10^{-4}	
Total	39			30		

(b)			
Source of variation	Degrees of freedom	Mean square	Pseudo-F
Origin	1	37,501	25.10**
Residual	44	1493.3	
Total	45	6.1×10^{-3}	

NS: nonsignificant; ** $P < 0.01$.

experiment, the RI-40 colonies were significantly more affected than those from RI-20 ($P < 0.01$) (Table 1b). Moreover, 83.3% (among which 66.7% did not show any necrosis) and 16.6% of the colonies from RI-20 and RI-40, respectively, survived in the common garden.

Population genetic analyses

No significant linkage disequilibrium was detected among the seven loci when all samples were considered (all $P > 0.05$ after FDR correction). Significant linkage disequilibrium was observed in PZ-40 for one pair of loci (*Mic22-Mic26*) after FDR correction. The mean (\pm standard deviation) observed heterozygosity over the loci (H_o Nei 1973) varied from 0.61 ± 0.23 for PZ-40 to 0.66 ± 0.22 for PZ-20, whereas the expected heterozygosity (H_e Nei 1973) varied from 0.67 ± 0.21 for PZ-20 to 0.76 ± 0.17 for RI-40. Significant heterozygote deficiencies were observed in all samples except PZ-20, with the obtained f values ranging from 0.02 for PZ-20 to 0.16 for RI-20 and RI-40. The allelic richness ($Ar_{(64)}$) ranged from 9.4 ± 7.6 for PZ-20 to 12.4 ± 9.2 for RI-40, showing a mean value of 10.7 ± 1.3 over the four samples (Table 2).

The global θ was 0.1 (95% CI: 0.07–0.13). The pairwise values and 95% CI for θ ranged from 0.02 (95% CI: 0.01–0.03) for RI-20 vs. RI-40 to 0.14 (95% CI: 0.09–0.19) for RI-20 vs. PZ-20. The exact tests for genotypic differentiation were significant at the global level and for the six pairwise comparisons (Table S2). PZ-20 and PZ-40 were the most divergent samples as illustrated by the phenogram (Fig. 4A).

The mean percentages of the assignment of the individuals to their sampling localities computed with STRUCTURE were 88.3, 60.3, 97.4, and 97.6% for RI-20, RI-40, PZ-20, and PZ-40, respectively (Fig. 4B). The values obtained without the null alleles option were equivalent (not shown). The harmonic mean estimates of N_e ranged from 70.8 for PZ-40 to 2084.6 for PZ-20, with a negative value being obtained RI-20 and RI-40, as expected when the population is sufficiently large that no notable linkage disequilibrium is induced through genetic drift (Waples and Do 2010). The confidence intervals showed an upper limit of infinity, expect for PZ-40. The confidence interval for PZ-40 overlapped with that for PZ-20 but was significantly lower than those for RI-20 and RI-40 (Table 2).

P_{ST} - F_{ST} comparisons and sensitivity analysis

Regarding the RTE conducted at Riou and considering the shallow habitat (20 m) and $c/h^2 = 1$, the P_{ST} was 0.42 (95% CI: 0.32–0.53). This P_{ST} was significantly higher than θ and F_{STall} with c/h_{crit}^2 values that were equal to 0.07 and 0.24, respectively (Fig. 2B). In the mesophotic habitat (40 m), the P_{ST} for $c/h^2 = 1$ was 0.06 (95% CI: 0.01–0.24), which was not significantly different from θ and F_{STall} . Regarding the RTE in Palazzu and considering the shallow habitat and $c/h^2 = 1$, the P_{ST} was 0.29 (95% CI: 0.01–0.68), which was not significantly different from θ and F_{STall} . Regarding the RTE in Palazzu, the P_{ST} values computed in the shallow and mesophotic habitat were 0. For the CGE, the P_{ST} under the null hypothesis was 0.44 (95% CI: 0.42–0.49), which was significantly higher than θ and F_{STall} . The c/h_{crit}^2 was 0.04 for θ and shifted to 0.15 when considering F_{STall} (Fig. 3B).

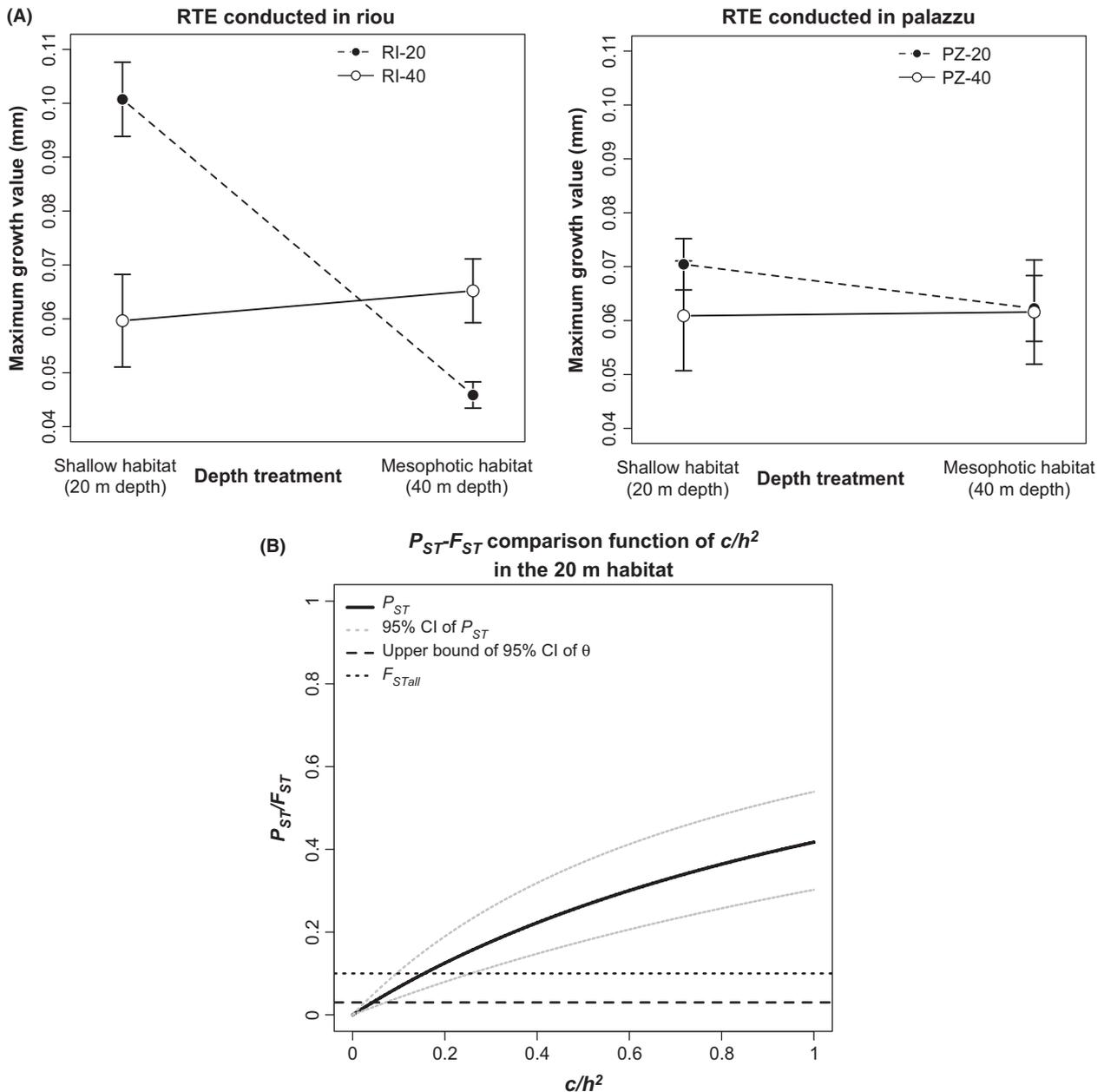


Figure 2. (A) Results of the RTEs based on the mean maximum growth in diameter (bars: standard error; SE = SD/√n with SD = standard deviation and n = sample number); (B) Sensitivity analysis of the P_{ST} - F_{ST} comparison conducted for the shallow habitat of the RTE in Riou. The occurrence of divergent selection on the considered trait (growth) is suggested when P_{ST} is significantly higher than F_{ST} . The robustness of the P_{ST} - F_{ST} comparison is tested by considering $c/h^2 \leq 1$ and by estimating the c/h^2_{crit} (i.e., the c/h^2 value beyond which P_{ST} is significantly higher than F_{ST} ; Brommer 2011) accounting for the upper limit of the 95% CI of θ and F_{STall} .

Discussion

Patterns and processes linked to ecology and evolution in marginal habitats are central in our understanding of the responses of natural populations facing global change (Hampe and Petit 2005; Kawecki 2008; Sexton et al. 2009). While neutral processes at species range

margin are receiving increasing attention (Eckert et al. 2008), empirical studies examining adaptation in these populations remain scarce (Bridle and Vines 2006 but see Kawecki 2008) despite their relevance for the conservation of biodiversity. The present study is a first step toward a better characterization of the processes underlying the interactions between marginal red coral

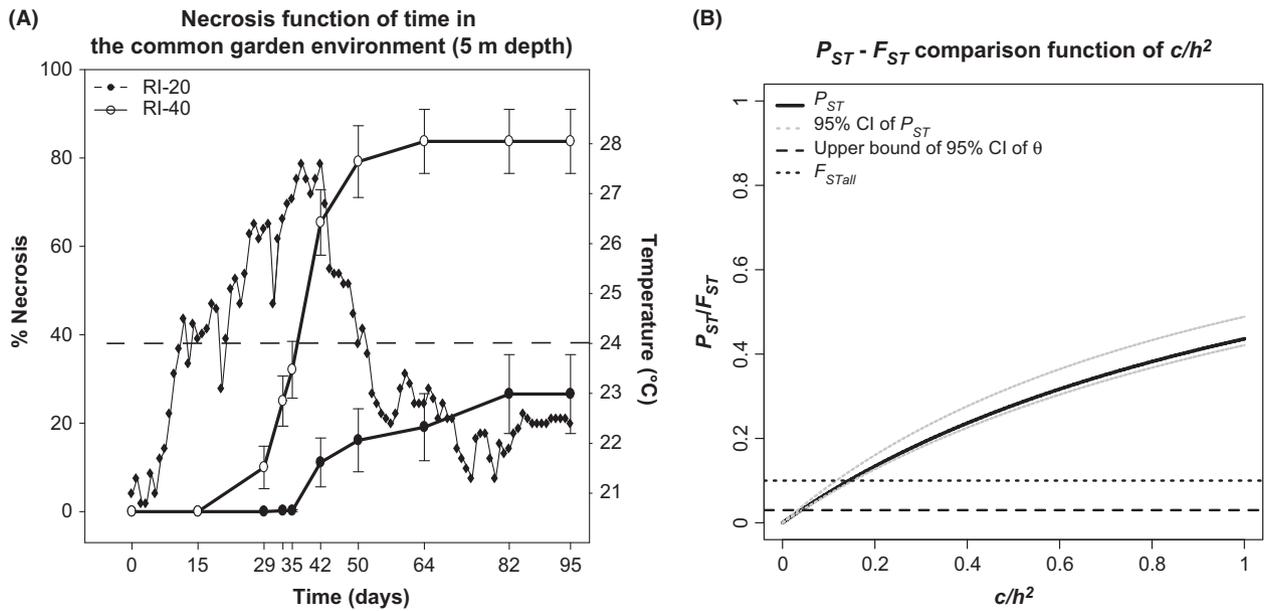


Figure 3. (A) Results of the CGE based on the percentage of necrosis (bars: standard error; SE = SD/ \sqrt{n} with SD = standard deviation). Filled (●) and open (○) circles indicate necrosis for RI-20 ($n = 24$) and RI-40 ($n = 24$), respectively: black diamonds (◆) and dashed lines correspond to the temperature measured in the common garden environment and to the thermal threshold of 24°C; (B) Sensitivity analysis of the P_{ST} - F_{ST} comparison. The occurrence of divergent selection on the considered trait (necrosis) is suggested when P_{ST} is significantly higher than F_{ST} . The robustness of the P_{ST} - F_{ST} comparison is tested by considering $c/h^2 \leq 1$ and by estimating the c/h^2_{crit} (i.e., the c/h^2 value beyond which P_{ST} is significantly higher than F_{ST} ; Brommer 2011) accounting for the upper limit of the 95% CI of θ and F_{STall} .

Table 2. Genetic characterization of the samples used for the RTEs.

Sample name (number of individuals)	Genetic diversity				Effective population size N_e (95% CI)
	Observed heterozygosity H_o (SD)	Gene diversity H_e (SD)	Estimator of F_{IS} f	Allelic richness $Ar_{(64)}$ (SD)	
RI-20 (43)	0.63 (0.30)	0.75 (0.16)	0.16***	11 (7.5)	-595.1.7 (226.4-∞)
RI-40 (43)	0.63 (0.28)	0.76 (0.17)	0.16***	12.4 (9.2)	-533.6 (307.5-∞)
PZ-20 (42)	0.66 (0.22)	0.67 (0.21)	0.02 (NS)	9.4 (7.6)	2084.6 (125.6-∞)
PZ-40 (45)	0.61 (0.23)	0.71 (0.2)	0.15***	10 (6.6)	70.8 (45.4-136.2)

NS: nonsignificant deviation from panmixia; ***: significant deviation from panmixia at 0.01; SD: standard deviation; 95% CI: 95% confidence interval.

populations and their environment in the context of climate change.

Contrasting population-by-environment interactions: the potential for local adaptation in marginal red coral populations

Using the mean maximum growth in diameter as a proxy for fitness, we validate the “local vs. foreign criterion” for the RTE conducted at Riou between one shallow and one mesophotic population separated by approximately one hundred meters. This criterion was rejected at Palazzu.

Focusing on Riou, two main hypotheses may explain the observed pattern of PEIs. One or both populations may be locally adapted to the environment due to the impact of divergent selection. Alternatively, by chance, other evolutionary forces may induce a pattern of differentiation concordant with the “local vs. foreign criterion” (Kawecki and Ebert 2004). The P_{ST} - F_{ST} comparisons computed for each habitat in the two RTEs allow us to refine this result. Indeed, while no difference was observed at Palazzu, the P_{ST} value computed under the null hypothesis ($c/h^2 = 1$) was significantly higher than the F_{ST} in the shallow habitat in Riou. The sensitivity analyses showed that this difference between the two measures of differentiation might be

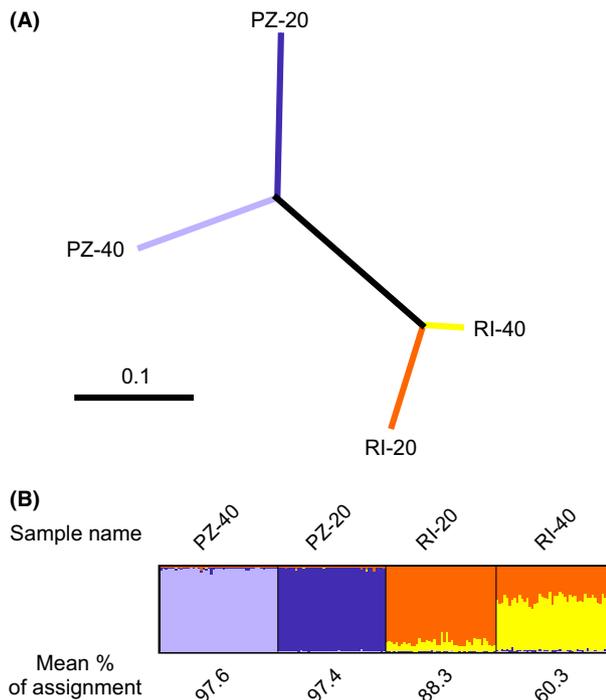


Figure 4. (A) Neighbor-joining phenogram using the distance of Nei et al. (1983) (D_a). The bootstrap values ($n = 1000$) were equal to 1000 (not shown); (B) Results of the clustering analysis conducted in STRUCTURE for $K = 4$ considering the individuals used in the RTEs. Each individual is represented by a vertical line partitioned into four colored segments that represent the fraction of individual membership in each of the four clusters. Mean percentage of assignment of individuals for each of the four samples is shown above the plot.

considered as relatively robust. Indeed, the c/h_{crit}^2 for the shallow habitat when considering θ or F_{STall} was lower than or slightly superior to 0.2, which is a threshold that has been suggested for realistic approximations of Q_{ST} by P_{ST} (Brommer 2011). The combination of the RTEs and the $P_{ST}-F_{ST}$ comparisons suggests therefore that some marginal shallow populations of *Corallium rubrum*, such as the RI-20 population, may potentially be locally adapted to their environment. This result fits with the re-evaluation of the importance of local adaptation in the marine realm (Conover et al. 2006; Marshall et al. 2010; Sanford and Kelly 2010). The present study is one of the few to focus on subtidal and benthic species (but see Prada et al. 2008; Bongaerts et al. 2011; Howells et al. 2012; Mariani et al. 2012) and on populations separated by approximately one hundred meters. Nevertheless, considering the limitations linked to the RTE and the $P_{ST}-F_{ST}$ comparisons, complementary studies involving replication at the population level (see Kawecki and Ebert 2004) or genomic approach (Palumbi et al. 2014) are needed to confirm the impact of divergent selection on the observed PEI.

Recent studies demonstrated the occurrence of complex spatial structure of PEIs in various organisms over their whole distribution area or at their range margins (e.g., Willi et al. 2007; Rogell et al. 2010; Kelly et al. 2011; Hice et al. 2012; Vergeer and Kunin 2013). The contrasting patterns of PEIs reported here are concordant with these studies. We can put forth environmentally or genetically based hypotheses to explain this result. The possibility that a lack of divergent selection induced the absence of local adaptation at Palazzu seems unlikely. Indeed, the two habitats show different abiotic properties, as illustrated by the two thermal regimes at PZ-20 and PZ-40, which were more divergent than between RI-20 and RI-40 (Bensoussan et al. 2010). It should therefore be considered that other selective pressures are at play, or, that the populations exhibit different genetic characteristics. Several theoretical studies have demonstrated the importance of the interaction between drift and gene flow in local adaptation (Alleaume-Benharira et al. 2006; Bridle et al. 2010). A meta-analysis of local adaptation also linked the size of populations with their propensity to be locally adapted (Leimu and Fischer 2008). Interestingly, our results support a stronger impact of drift at Palazzu compared to Riou, particularly in PZ-20. PZ-20 and PZ-40 showed the lowest expected heterozygosity and allelic richness and were characterized by the two lowest effective population sizes, and the confidence interval for PZ-40 was significantly smaller than those for RI-20 and RI-40. Moreover, the Palazzu populations were more isolated than those from Riou as illustrated by the higher mean assignment of individuals and genetic divergence. It is also noteworthy that the PZ-20, PZ-40 and RI-20, RI-40 populations belong to two genetic clusters, with the cluster encompassing PZ-20 and PZ-40 being more impacted by drift (Ledoux et al. 2010a). In accordance with model predictions demonstrating that the influence of the effective population size on adaptation increases in poorly connected populations (Alleaume-Benharira et al. 2006; Bridle et al. 2010), we hypothesize that the lack of local adaptation in Palazzu may be related to the stronger influence of the genetic drift on these populations. Accordingly, we posit that adaptive processes of the red coral may be more influenced by genetic drift than by gene flow. If confirmed, this result may have important consequences for the conservation of the species (see below).

Phenotypic buffering for thermal stress, likely driven by a heterogeneous environment

The *in situ* CGE conducted at Riou supported the occurrence of differential responses to environmental disturbance. The percentage of survival in the stressful

environment of the common garden was significantly higher for the shallow colonies than for the mesophotic colonies. The temperature of the common garden was likely a predominant agent in the observed responses. Indeed, the first signs of necrosis in RI-40 coincided with a temperature increase beyond 24°C, which was suggested as a thermal threshold for the red coral (Torrents *et al.* 2008). Such differential phenotypic buffering between populations facing thermal stress has been previously reported in corals (e.g., Howells *et al.* 2012) and Mediterranean gorgonians (e.g., Torrents *et al.* 2008). Focusing on two red coral populations separated by 5 km, Torrents *et al.* (2008) demonstrated *in aquaria* that the population dwelling in the warmer habitat (10 m) was significantly less affected by a thermal stress than the population dwelling at mesophotic depth (40 m). Our study substantially refines these results because the differential response was observed *in situ* and between individuals originating from shallow and mesophotic populations separated by approximately one hundred meters. This is concordant with the independent functioning of populations suggested by the significant genetic structure reported at this spatial scale (Ledoux *et al.* 2010a). This is also supported by field surveys conducted after mass mortality events that found contrasting levels of necrosis between populations separated by the same range of distances (Garrabou *et al.* 2009). Differential phenotypic buffering between populations may be due to distinct processes. In the present case, whether the observed differential phenotypic buffering relies on genetic adaptation or environmental effects remains an open question that requires further studies. Nevertheless, a bunch of evidences allowed us to suggest that a genetic adaptation of the shallow colonies to their local environment may explain their ability to buffer the thermal stress induced by the common garden habitat. First, the difference between the two indices of differentiation, the F_{ST} and the P_{ST} , was significant and robust ($c/h_{crit}^2 = 0.04$), as expected when divergent selection leading to genetic adaptation is involved in the observed pattern (Saether *et al.* 2007; Brommer 2011; Leinonen *et al.* 2013). Then, the degree and predictability of environmental heterogeneity seems to be an important driver in the evolution of adaptive plasticity (Scheiner 1993; Via *et al.* 1995). Interestingly, the thermal regime in the shallow environment is characterized by strong seasonality with more contrasting winter and summer periods than in the mesophotic environment (Bensoussan *et al.* 2010). Temperature fluctuations associated with nonlethal stress observed in summer in the shallow habitat (Bensoussan *et al.* 2010) could have beneficial impacts on the red coral colonies, increasing their thermotolerance. This is supported by a recent study focused on the expression of heat shock proteins in the red coral that

showed differential expression of HSP70 as a function of the thermal history of individuals (Haguenauer *et al.* 2013). Beneficial impacts of nonlethal stress on thermotolerance have also been reported for many terrestrial (e.g., Deutsch *et al.* 2008) and marine species (e.g., Barshis *et al.* 2010; Oliver and Palumbi 2011; Carilli *et al.* 2012). Accordingly, we posit that the phenotypic buffering capacities of the shallow colonies might constitute a genetically based adaptation to the seasonal thermal fluctuations observed in the shallow environment. Nevertheless, complementary studies are needed to formally confirm this hypothesis and to test the importance of environmental effects such as maternal effects or epigenetic inheritance on the differential phenotypic buffering.

Red coral conservation and the deep refugia hypothesis

Complementary studies involving more populations and phenotypic traits are needed to generalize our conclusions. Considering the limitations of the P_{ST} - F_{ST} comparisons, genomic approaches should also be considered to confirm the role played by divergent selection in the observed patterns. Nevertheless, we demonstrated the occurrence of contrasting PEIs with potential for local adaptation at fine spatial scales in a survey involving only four populations. This implies the existence of putatively important adaptive diversity in marginal red coral populations.

From a conservation perspective, genetic drift is an important evolutionary force due to its impact on genetic diversity and inbreeding and, ultimately, on the evolutionary potential of a population (Frankham *et al.* 2002; Allendorf and Luikart 2007). In this context, our results are particularly relevant. Indeed, genetic drift seems to play a central role in the PEIs in the red coral because the drifting populations of Palazzu are not locally adapted. Conservation priority should thus focus on this evolutionary force by restraining the density erosion of marginal populations directly caused by anthropogenic activities (e.g., harvesting) in order to preserve their evolutionary potential. Additionally, we showed that under realistic environmental stresses, the performance of mesophotic colonies was significantly lower than that of shallow ones. Combining this result with the limited connectivity observed between populations (Ledoux *et al.* 2010a), the ability of mesophotic populations to replenish shallow populations appears restricted. Using mesophotic populations as a source for restoration efforts aimed at shallow habitats is also expected to be inefficient. Furthermore, we demonstrated the capacity of the shallow population at Riou to cope with the expected increase in sea temperature over short to intermediate timescale (Somot

et al. 2008). Shallow populations adapted to marginal environmental conditions could therefore act as reservoir of adaptive genetic variation (Willi et al. 2007; Gienapp et al. 2008). These results unambiguously question the validity of the deep refugia hypothesis for the red coral and other sessile organisms with similar life history traits. Accordingly, we call for the development of studies focused on the adaptive potential of shallow and marginal marine populations to test whether the shallow reservoir hypothesis, rather than the deep refugia hypothesis, should be considered in marine conservation efforts.

Acknowledgments

We are grateful to Bernard de Ligondes, Roland Graille, and Frédéric Zuberer (Centre d'Océanologie de Marseille) for support during sampling and design of the experimental plates and to Jean-Marie Dominici (Réserve Naturelle de Scandola, Parc Régional de Corse) for his help during the experimentation conducted in Palazzu. We thank Caroline Rocher for technical assistance during genotyping analyses and Guillem Salazar for his support during R programming. We show gratitude to Drs. Ophélie Ronce and Frédérique Viard, Jose M. Montoya and Jérôme Goudet for useful comments during the preparation of the manuscript and to Dr. Clément Gilbert for precious advices during the redaction process. We are also grateful to Jean-Georges Harmelin for sharing data regarding genetic differentiation based on allozyme loci. This study was funded by the "Provence-Alpes-Côte d'Azur" region (J.-B. Ledoux Doctoral Fellowship) and by the IFB Foundation (Institut Français de la Biodiversité), the environmental department of TOTAL-FINA-ELF group, an "Action Concertée Initiative" (ACI) program "Jeunes chercheurs 2003" (number JC5360) from the French Research Ministry and the MEDCHANGE project funded by the Agence Nationale pour la Recherche (ANR) J.-B. L. is supported by a post-doctoral grant SFRH/BPD/74400/2010 from Fundação para a Ciência e Tecnologia (FCT; Portugal).

Conflict of Interest

None declared.

References

- Abbiati, M., G. Santangelo, and S. Novelli. 1993. Genetic variation within and between two Tyrrhenian populations of the Mediterranean alcyonarian *Corallium rubrum*. *Mar. Ecol. Prog. Ser.* 95:245–250.
- Alleaume-Benharira, M., I. R. Pen, and O. Ronce. 2006. Geographical patterns of adaptation within a species' range: interactions between drift and gene flow. *J. Evol. Biol.* 19:203–215.
- Allendorf, F. W., and G. Luikart. 2007. Conservation and the genetics of populations. Blackwell Publishing, Oxford, UK.
- Anderson, M. J. 2001. A new method for non-parametric multivariate analysis of variance. *Austral Ecol.* 26:32–46.
- Anderson, M. J., R. N. Gorley, and K. R. Clarke. 2008. PERMANOVA for PRIMER: guide to software and statistical methods. Pp. 214. PRIMER-E Ltd, Plymouth, UK.
- Antoniazza, S., R. Burri, L. Fumagalli, J. Goudet, and A. Roulin. 2010. Local adaptation maintains clinal variation in melanin-based coloration of European Barn Owls *Tyto alba*. *Evolution* 64:1944–1954.
- Aurelle, D., and J. B. Ledoux. 2013. Interplay between isolation by distance and genetic clusters in the red coral *Corallium rubrum*: insights from simulated and empirical data. *Conserv. Genet.* 14:705–716.
- Aurelle, D., J. B. Ledoux, C. Rocher, P. Borsa, A. Chenuil, and J. P. Féral. 2011. Phylogeography of the red coral *Corallium rubrum*: inferences on the evolutionary history of a temperate gorgonian. *Genetica* 139:855–869.
- Ballesteros, E. 2006. Mediterranean coralligenous assemblages: a synthesis of present knowledge. *Oceanogr. Marine Biol.* 44:123–195.
- Barshis, D., J. Stillman, R. Gates, R. J. Toonen, L. W. Smith, and C. Birkeland. 2010. Protein expression and genetic structure of the coral *Porites lobata* in an environmentally extreme Samoan back reef: does host genotype limit phenotypic plasticity? *Mol. Ecol.* 19:1705–1720.
- Baythavong, B. S. 2011. Linking the spatial scale of environmental variation and the evolution of phenotypic plasticity: selection favors adaptive plasticity in fine-grained environments. *Am. Nat.* 178:75–87.
- Baythavong, B. S., and M. L. Stanton. 2010. Characterizing selection on phenotypic plasticity in response to natural environmental heterogeneity. *Evolution* 64:2904–2920.
- Bensoussan, N., J. C. Romano, J. G. Harmelin, and J. Garrabou. 2010. High resolution characterization of northwest Mediterranean coastal waters thermal regimes: to better understand responses of benthic communities to climate change. *Estuar. Coast. Shelf Sci.* 87:431–441.
- Bongaerts, P., T. Ridgway, E. M. Sampayo, and O. Hoegh-Guldberg. 2010. Assessing the 'deep reef refugia' hypothesis: focus on Caribbean reefs. *Coral Reefs* 29:309–327.
- Bongaerts, P., C. Riginos, K. B. Hay, M. J. van Oppen, O. Hoegh-Guldberg, and S. Dove. 2011. Adaptive divergence in a scleractinian coral: physiological adaptation of *Seriatopora hystrix* to shallow and deep reef habitats. *BMC Evol. Biol.* 11:303.
- Brazeau, D. A., M. P. Lesser, and M. Slattery. 2013. Genetic structure in the coral, *Montastraea cavernosa*: assessing genetic differentiation among and within mesophotic reefs. *PLoS One* 8:e0065845.

- Bridle, J. R., and T. H. Vines. 2006. Limits to evolution at range margins: when and why does adaptation fail? *Trends Ecol. Evol.* 22:140–147.
- Bridle, J. R., J. Polechová, M. Kawata, and R. K. Butlin. 2010. Why is adaptation prevented at ecological margins? New insights from individual-based simulations. *Ecol. Lett.* 13:485–494.
- Brommer, J. E. 2011. Whither Pst? The approximation of Qst by Pst in evolutionary and conservation biology. *J. Evol. Biol.* 24:1160–1168.
- Bruckner, A. W. 2009. Rate and extent of decline in *Corallium* (pink and red coral) populations: existing data meet the requirements for a CITES Appendix II listing. *Mar. Ecol. Prog. Ser.* 397:319–332.
- Bürger, R., and M. Lynch. 1995. Evolution and extinction in a changing environment - A quantitative-genetic analysis. *Evolution* 49:151–163.
- Carilli, J., S. D. Donner, and A. C. Hartmann. 2012. Historical temperature variability affects coral response to heat stress. *PLoS One* 7:e0034418.
- Chapuis, M. P., and A. Estoup. 2007. Microsatellite null alleles and estimation of population differentiation. *Mol. Biol. Evol.* 24:621–631.
- Clarke, K. R., and R. N. Gorley. 2006. PRIMERv6: user manual/tutorial. Pp. 192. PRIMER-E, Plymouth.
- Conover, D. O., L. M. Clarke, S. B. Munch, and G. N. Wagner. 2006. Spatial and temporal scales of adaptive divergence in marine fishes and the implications for conservation. *J. Fish Biol.* 69:21–47.
- Costantini, F., M. Taviani, A. Remia, E. Pintus, P. J. Schembri, and M. Abbiati. 2010. Deep-water *Corallium rubrum* L, 1758 from the Mediterranean Sea: preliminary genetic characterization. *Mar. Ecol.* 31:261–269.
- Costantini, F., S. Rossi, E. Pintus, C. Cerrano, J. M. Gili, and M. Abbiati. 2011. Low connectivity and declining genetic variability along a depth gradient in *Corallium rubrum* populations. *Coral Reefs* 30:991–1003.
- Deutsch, C., J. Tewksbury, R. Huey, K. S. Sheldon, C. K. Ghalambor, and D. C. Haak. 2008. Impacts of climate warming on terrestrial ectotherms across latitude. *Proc. Natl Acad. Sci. USA* 105:6668–6672.
- Eckert, C. G., K. E. Samis, and S. C. Loughheed. 2008. Genetic variation across species' geographical ranges: the central-marginal hypothesis and beyond. *Mol. Ecol.* 17:1170–1188.
- Edelaar, P., and M. Björklund. 2011a. If FST does not measure neutral genetic differentiation, then comparing it with QST is misleading. Or is it? *Mol. Ecol.* 20:1805–1812.
- Edelaar, P., P. Burraco, and I. Gomez-Mestre. 2011b. Comparisons between Q_{ST} and F_{ST} - how wrong have we been? *Mol. Ecol.* 20:4830–4839.
- Falush, D., M. Stephens, and J. K. Pritchard. 2003. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics* 164:1567–1587.
- Falush, D., M. Stephens, and J. K. Pritchard. 2007. Inference of population structure using multilocus genotype data: dominant markers and null alleles. *Mol. Ecol. Notes* 7:574–578.
- Frankham, R., J. D. Ballou, and D. A. Briscoe. 2002. Introduction to conservation genetics. Cambridge University Press, Cambridge, UK.
- Garner, T. W. J., P. B. Pearman, and S. Angelone. 2004. Genetic diversity across a vertebrate species' range: a test of the central-peripheral hypothesis. *Mol. Ecol.* 13:1047–1053.
- Garrabou, J., and J. G. Harmelin. 2002. A 20-year study on life-history traits of a harvested long-lived temperate coral in the NW Mediterranean: insights into conservation and management needs. *J. Anim. Ecol.* 71:966–978.
- Garrabou, J., R. Coma, M. Bally, N. Bensoussan, P. Chevaldonné, M. Cigliano, et al. 2009. Mass mortality in northwestern Mediterranean rocky benthic communities: effects of the 2003 heat wave. *Glob. Change Biol.* 15:1090–1103.
- Ghalambor, C. K., J. K. McKay, S. P. Carroll, and D. N. Reznick. 2007. Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Funct. Ecol.* 21:394–407.
- Gienapp, P., C. Teplitsky, J. S. Alho, J. A. Mills, and J. Merila. 2008. Climate change and evolution: disentangling environmental and genetic responses. *Mol. Ecol.* 17:167–178.
- Glynn, P. W. 1996. Coral reef bleaching: facts, hypotheses and implications. *Glob. Change Biol.* 2:495–509.
- Haguenaer, S., F. Zuberer, J. B. Ledoux, and D. Aurelle. 2013. Adaptive abilities of the Mediterranean red coral *Corallium rubrum* in an heterogeneous and changing environment: from population to functional genetics. *J. Exp. Mar. Biol. Ecol.* 449:349–357.
- Hall, V., and T. P. Hughes. 1996. Reproductive strategies of modular organisms: comparative studies of reef-building corals. *Ecology* 77:950–963.
- Hampe, A., and R. J. Petit. 2005. Conserving biodiversity under climate change: the rear edge matters. *Ecol. Lett.* 8:461–467.
- Hedrick, P. W. 2005. A standardized genetic differentiation measure. *Evolution* 59:1633–1638.
- Hice, L. A., T. A. Duffy, S. B. Munch, and D. O. Conover. 2012. Spatial scale and divergent patterns of variation in adapted traits in the ocean. *Ecol. Lett.* 15:568–575.
- Hill, W. G. 1981. Estimation of effective population size from data on linkage disequilibrium. *Genet. Res.* 38:209–216.
- Hoffmann, A. A., and M. W. Blows. 1994. Species borders - ecological and evolutionary perspectives. *Trends Ecol. Evol.* 9:223–227.
- Hoffmann, A. A., and C. M. Sgro. 2011. Climate change and evolutionary adaptation. *Nature* 470:479–485.
- Howells, E. J., V. H. Beltran, N. W. Larsen, L. K. Bay, B. L. Willis, and M. J. H. van Oppen. 2012. Coral thermal tolerance shaped by local adaptation of photosymbionts. *Nat. Clim. Chang.* 2:116–120.

- Hubisz, M. J., D. Falush, M. Stephens, and J. K. Pritchard. 2009. Inferring weak population structure with the assistance of sample group information. *Mol. Ecol. Resour.* 9:1322–1332.
- Jakobsson, M., and N. A. Rosenberg. 2007. CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. *Bioinformatics* 23:1801–1806.
- Jost, L. 2008. G_{st} and its relatives do not measure differentiation. *Mol. Ecol.* 17:4015–4026.
- Kawecki, T. J. 2008. Adaptation to marginal habitats. *Annu. Rev. Ecol. Evol. Syst.* 39:321–342.
- Kawecki, T. J., and D. Ebert. 2004. Conceptual issues in local adaptation. *Ecol. Lett.* 7:1225–1241.
- Kelly, M. W., E. Sanford, and R. K. Grosberg. 2011. Limited potential for adaptation to climate change in a broadly distributed marine crustacean. *Proc. R. Soc. B* 279:349–356.
- Langella, O. (2010) CNRS, UPR9034, France Populations.1.2.3.1 Available at <http://bioinformatics.org/~tryphon/populations/>
- Lawton, J. H. 1993. Range, population abundance, and conservation. *Trends Ecol. Evol.* 8:409–413.
- Ledoux, J. B., K. Mokthar-Jamai, C. Roby, J. P. Feral, J. Garrabou, and D. Aurelle. 2010a. Genetic survey of shallow populations of the Mediterranean red coral [*Corallium rubrum* Linnaeus, 1758]: new insights into evolutionary processes shaping nuclear diversity and implications for conservation. *Mol. Ecol.* 19:675–690.
- Ledoux, J. B., J. Garrabou, O. Bianchimani, P. Drap, J. P. Feral, and D. Aurelle. 2010b. Fine-scale genetic structure and inferences on population biology in the threatened Mediterranean red coral, *Corallium rubrum*. *Mol. Ecol.* 19:4204–4216.
- Leimu, R., and M. Fischer. 2008. A meta-analysis of local adaptation in plants. *PLoS One* 3:e0004010.
- Leinonen, T., J. M. Cano, H. Makinen, and J. Merila. 2006. Contrasting patterns of body shape and neutral genetic divergence in marine and lake populations of threespine sticklebacks. *J. Evol. Biol.* 19:1803–1812.
- Leinonen, T., R. O'hara, J. Cano, and J. Merila. 2008. Comparative studies of quantitative trait and neutral marker divergence: a meta-analysis. *J. Evol. Biol.* 21:1–17.
- Leinonen, T., R. J. S. McCairns, R. B. O'Hara, and J. Merila. 2013. Q_{ST} - F_{ST} comparisons: evolutionary and ecological insights from genomic heterogeneity. *Nat. Rev. Genetics* 14:179–190.
- Linares, C., R. Coma, and M. Zabala. 2008. Effects of a mass mortality event on gorgonian reproduction. *Coral Reefs* 27:27–34.
- Mariani, S., K. T. C. A. Peijnenburg, and D. Weetman. 2012. Independence of neutral and adaptive divergence in a low dispersal marine mollusk. *Mar. Ecol. Prog. Ser.* 446:173–187.
- Marshall, D. J., K. Monro, M. Bode, M. J. Keough, and S. Swearer. 2010. Phenotype-environment mismatches reduce connectivity in the sea. *Ecol. Lett.* 13:128–140.
- McArdle, B. H., and M. J. Anderson. 2001. Fitting multivariate models to community data: a comment on distance-based redundancy analysis. *Ecology* 82:290–297.
- Nei, M. 1973. Analysis of gene diversity in subdivided populations. *Proc. Natl Acad. Sci. USA* 70:3321–3323.
- Nei, M., F. Tajima, and Y. Tateno. 1983. Accuracy of estimated phylogenetic trees from molecular data II: gene frequency data. *J. Mol. Evol.* 19:153–170.
- Oliver, T., and S. R. Palumbi. 2011. Do fluctuating temperature environments elevate coral thermal tolerance? *Coral Reefs* 30:429–440.
- van Oppen, M. J., P. Bongaerts, J. N. Underwood, L. M. Peplow, and T. F. Cooper. 2011. The role of deep reefs in shallow reef recovery: an assessment of vertical connectivity in a brooding coral from west and east Australia. *Mol. Ecol.* 20:1647–1660.
- Orizaola, G., M. Quintela, and A. Laurila. 2010. Climatic adaptation in an isolated and genetically impoverished amphibian population. *Ecography* 33:730–737.
- Pigliucci, M. 2005. Evolution of phenotypic plasticity: where are we going now? *Trends Ecol. Evol.* 20:481–486.
- Prada, C., N. V. Schizas, and P. M. Yoshioka. 2008. Phenotypic plasticity or speciation? A case from a clonal marine organism. *BMC Evol. Biol.* 13:8–47.
- Pritchard, J. K., M. Stephens, and P. Donnelly. 2000. Inference of population structure using multilocus genotype data. *Genetics* 155:945–959.
- Pujol, B., A. J. Wilson, R. I. C. Ross, and J. R. Pannell. 2008. Are Q_{ST} - F_{ST} comparisons for natural populations meaningful? *Mol. Ecol.* 17:4782–4785.
- Reusch, T. B. H. 2014. Climate change in the oceans: evolutionary versus phenotypically plastic responses of marine animals and plants. *Evol. Appl.* 7:104–122.
- Reusch, T. B. H., and T. E. Wood. 2007. Molecular ecology of global change. *Mol. Ecol.* 16:3973–3992.
- Rogell, B., M. Eklund, H. Thörngren, A. Laurila, and J. Höglund. 2010. The effects of selection, drift and genetic variation on life-history trait divergence among insular populations of natterjack toad, *Bufo calamita*. *Mol. Ecol.* 19:2229–2240.
- Rosenberg, N. A. 2004. DISTRUCT: a program for the graphical display of population structure. *Mol. Ecol. Notes* 4:137–138.
- Rousset, F. 2008. GENEPOP'007: a complete re-implementation of the GENEPOP software for Windows and Linux. *Mol. Ecol. Resour.* 8:103–106.
- Saether, S. A., P. Fiske, J. A. Kalas, A. Kuresoo, L. Luigujoe, S. B. Piernney, et al. 2007. Inferring local adaptation from Q_{ST} - F_{ST} comparisons: neutral genetic and quantitative trait variation in European populations of great snipe. *J. Evol. Biol.* 20:1563–1576.

- Sagarin, R. D., and S. D. Gaines. 2002. The 'abundant centre' distribution: to what extent is it a biogeographical rule? *Ecol. Lett.* 5:137–147.
- Saitou, N., and M. Nei. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4:406–425.
- Sanford, E., and M. W. Kelly. 2010. Local adaptation in marine invertebrates. *Annu. Rev. Mar. Sci.* 3:509–535.
- Scheiner, S. M. 1993. Genetics and evolution of phenotypic plasticity. *Annu. Rev. Ecol. Syst.* 24:35–68.
- Sexton, J. P., P. J. McIntyre, A. L. Angert, and K. J. Rice. 2009. Evolution and ecology of species range limits. *Annu. Rev. Ecol. Syst.* 40:415–436.
- Somot, S., F. Sevault, M. Déqué, and M. Crépon. 2008. 21st century climate change scenario for the Mediterranean using a coupled atmosphere-ocean regional climate model. *Global Planet. Change* 63:112–126.
- Spitze, K. 1993. Population structure in *Daphnia obtusa*: quantitative genetic and allozymic variation. *Genetics* 135:367–374.
- Torrents, O., J. Garrabou, C. Marschal, and J. G. Harmelin. 2005. Age and size at first reproduction in the commercially exploited red coral *Corallium rubrum* (L) in the Marseilles area (France, NW Mediterranean). *Biol. Conserv.* 121:391–397.
- Torrents, O., E. Tambutté, N. Caminiti, and J. Garrabou. 2008. Upper thermal thresholds of shallow vs deep populations of the precious Mediterranean red coral *Corallium rubrum* L: assessing the potential effects of warming in the NW Mediterranean. *J. Exp. Mar. Biol. Ecol.* 357:7–19.
- Vergeer, P., and W. E. Kunin. 2013. Adaptation at range margins: common garden trials and the performance of *Arabidopsis lyrata* across its North-western European range. *New Phytol.* 197:989–1001.
- Via, S., R. Gomulkiewicz, G. D. Jong, S. M. Scheiner, C. D. Schlichting, and P. H. van Tienderen. 1995. Adaptive phenotypic plasticity: consensus and controversy. *Trends Ecol. Evol.* 10:212–217.
- Waples, R. S. 2006. A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. *Conserv. Genet.* 7:167–184.
- Waples, R. S., and C. Do. 2008. Ldne: a program for estimating effective population size from data on linkage disequilibrium. *Mol. Ecol. Resour.* 8:753–756.
- Waples, R. S., and C. Do. 2010. Linkage disequilibrium estimates of contemporary N_e using highly variable genetic markers: a largely untapped resource for applied conservation and evolution. *Evol. Appl.* 3:246–262.
- Weir, B. S., and C. C. Cockerham. 1984. Estimating F-statistics for the analysis of population structure. *Evolution* 38:1358–1370.
- Whitlock, M. 2008. Evolutionary inference from Q_{ST} . *Mol. Ecol.* 17:1885–1896.
- Willi, Y., J. van Buskirk, B. Schmid, and M. Fischer. 2007. Genetic isolation of fragmented populations is exacerbated by drift and selection. *J. Evol. Biol.* 20:534–542.
- Wright, S. 1943. Isolation by distance. *Genetics* 28:114–138.

Supporting Information

Additional Supporting Information may be found in the online version of this article:

Appendix S1. Experimental plates and calcein labeling.

Appendix S2. Analysis of the growth in diameter.

Appendix S3. Null allele frequencies, microsatellite characteristics, Hardy-Weinberg equilibrium and genetic diversity.

Appendix S4. R-scripts for the P_{ST} - F_{ST} comparisons and corresponding sensitivity analyses.

Table S1. (a) Mean temperature conditions from July 1st to September 30th: mean, coefficient of variation (CV) and percentage of time with temperature higher or equal to 20 and 24 °C at 5, 20 and 40 m depth at Riou and 20 and 40 m depth at Palazzu. (b) Mean and maximum temperature values and percentage of time with temperature higher or equal to 20 and 24°C at 5 m depth for the 2006 summer at Riou.

Table S2. Pairwise θ values and corresponding 95% CI interval are shown below the diagonal.