

SCIENTIFIC REPORTS



OPEN

Photoinhibition of *Phaeocystis globosa* resulting from oxidative stress induced by a marine algicidal bacterium *Bacillus* sp. LP-10

Received: 09 March 2015
Accepted: 22 October 2015
Published: 25 November 2015

Chengwei Guan^{1,2,*}, Xiaoyun Guo^{1,3,*}, Yi Li^{1,4}, Huajun Zhang¹, Xueqian Lei¹, Guanjing Cai¹, Jiajia Guo¹, Zhiming Yu⁵ & Tianling Zheng¹

Harmful algal blooms caused by *Phaeocystis globosa* have resulted in staggering losses to coastal countries because of their world-wide distribution. Bacteria have been studied for years to control the blooms of harmful alga, however, the action mechanism of them against harmful algal cells is still not well defined. Here, a previously isolated algicidal bacterium *Bacillus* sp. LP-10 was used to elucidate the potential mechanism involved in the dysfunction of *P. globosa* algal cells at physiological and molecular levels. Our results showed *Bacillus* sp. LP-10 induced an obvious rise of reactive oxygen species (ROS), which was supposed to be major reason for algal cell death. Meanwhile, the results revealed a significant decrease of photosynthetic physiological indexes and apparent down-regulated of photosynthesis-related genes (*psbA* and *rbcS*) and protein (PSII reaction center protein D1), after treated by *Bacillus* sp. LP-10 filtrates, suggesting photoinhibition occurred in the algal cells. Furthermore, our results indicated that light played important roles in the algal cell death. Our work demonstrated that the major lethal reason of *P. globosa* cells treated by the algicidal bacterium was the photoinhibition resulted from oxidative stress induced by *Bacillus* sp. LP-10.

Microalgae play an important role in carbon sink during the transportation of the carbon to deep waters. However, many of them lead to harmful algal blooms (HABs), causing adverse effects on marine ecosystems and economics^{1,2}, even harm people by producing toxins³. Many methods were developed to manage the HABs^{4,5}. However, physical and chemical methods are not supposed to be optimal choices in controlling HABs, because of their obvious defects of high costs and unavailability for large-scale application, and more importantly, bringing about secondary pollution. Hence, biological methods are regarded as more promising approaches to regulate HABs^{6,7}.

Marine bacteria are important in regulating the growth dynamics of microalga^{8,9}. Among them, a group of bacteria, now called algicidal bacteria, are suggested to show inhibitory effects on algal growth, which are considered as excellent candidates to control HABs¹⁰. The action mode of algicidal bacteria on target algal cells were divided into two types, one is lysing the algal cells by contacting with the cells

¹State Key Laboratory of Marine Environmental Science and Key Laboratory of Ministry of Education for Coastal and Wetland Ecosystems, School of Life Sciences, Xiamen University, Xiamen 361102, China. ²Tobacco Science Institute of Jiangxi Province, Nanchang 330025, China. ³School of Chemistry and Chemical Engineering, Xiamen University, Xiamen 361102, China. ⁴College of Life Sciences, Henan Normal University, Xinxiang 453007, China. ⁵Key Laboratory of Marine Ecology and Environmental Science, Institute of Oceanology, Chinese Academy of Sciences, Qingdao 266071, China. *These authors contributed equally to this work. Correspondence and requests for materials should be addressed to T.Z. (email: wshwzh@xmu.edu.cn) or Z.Y. (email: zyu@ms.qdio.ac.cn)

surface directly, named direct attack mode, the other is lysing the algal cells by releasing active substances/allelochemicals, named indirect attack mode^{11,12}. Most algicidal bacteria reported so far belongs to the latter action mode^{13,14}.

Despite a lot of algicidal bacteria were isolated, the action mechanisms of them against harmful algal cells were still not well defined. Recently, some researches revealed that reactive oxygen species (ROS) played significant roles in the inhibitory effects of algicidal bacteria on algal growth^{15,16}. Some allelochemicals were documented to trigger the producing of ROS, leading to algal cells damage^{17,18}.

As is known, environmental stresses usually induce algal cell to produce ROS, which change the oxidative level and breaks the redox balance of the algal cells^{19,20}. Normally, antioxidant systems of cells, including superoxide dismutase (SOD), peroxidase (POD), catalase (CAT) and glutathione (GSH) etc., could protect themselves from oxidative damage by clearing the numerous ROS to keep cells at a stable redox level^{21,22}.

Photosynthetic systems are fundamental for the growth of plant and algal cells. Generally, performing oxygenic photosynthesis will cause damage to the photosynthetic apparatus²³. The reaction center of photosystem II (PSII) was one of the main damage target sites, of which the protein D1 is sensitive and vulnerable to stresses²⁴. Photosynthetic organisms execute self-repair program to replace the impaired protein D1 via *de novo* synthesis. Normally, the protein D1 content is in a dynamic balance. However, when the protein D1 damage rate exceeds the repair rate, photoinhibition occurs in photosynthetic organisms²⁴. Recent research revealed high light induced lower expression of protein D1 in diatom *Phaeodactylum tricorutum* than low light exposure²⁵. Recent researches showed that environmental factors, such as NaCl, heat and low temperature can also lead to photoinhibition by interrupting the *de novo* synthesis of protein D1^{23,26}.

Phaeocystis globosa Scherffel (Prymnesiophyceae) is a HABs causing species with world-wide distribution, which caused huge economic losses to coastal countries²⁷. *Bacillus* sp. LP-10, a marine bacterium obtained in our laboratory before, was proved to be effective in controlling the growth of *P. globosa*²⁸. However, few work were performed to illustrate the action mode of the bacterium on *P. globosa*. To better understand the action mechanism of the algicidal mechanism, a systematic investigation of physiological and molecular alterations in algal cells was carried out. And the physiological data coupled with transcript and protein analysis provided new insights into the algicidal mechanism of the marine bacterium on target algal cells.

Materials and Methods

Strains and culture conditions. Cultures of *P. globosa* PG03 were supplied by the State Key Laboratory of Marine Environmental Sciences, Xiamen University, China and *f/2* (-Si) medium was used to prepare algal cultures for the experiments²⁹. The light conditions were set to a 12h/12h light-dark cycle under cool-white fluorescent light with an intensity of 50 $\mu\text{mol photons m}^{-2}\text{s}^{-1}$ at $20 \pm 1^\circ\text{C}$.

The *Bacillus* sp. LP-10 strain was previously isolated in surface water samples from the East China Sea in our laboratory, and the GenBank accession number is KF411459. The marine bacterial strain was prepared in autoclaved Zobell 2216E medium and cultured at 28°C for 48 h with 150 rpm rotation speed³⁰. The bacterium cultures at stationary phase were collected and centrifuged at 8000 g for 10 min. The cell-free filtrates of *Bacillus* sp. LP-10 were prepared by filtering the supernatant through 0.22 μm size pore Millipore membrane filters. The cell-free filtrates of *Bacillus* sp. LP-10 was used in the whole study unless specially declaration. To ensure that the *Bacillus* sp. LP-10 is safe for the environment, the toxicity of the strain were tested. The test results indicated *Bacillus* sp. LP-10 were environmentally friendly to the aquatic environment (Supplementary Information).

ROS assays. The fluorescence probe 2', 7'-dichlorofluorescein diacetate (DCFH-DA) is used to measure the ROS level in algal cells treated with cell-free filtrates of *Bacillus* sp. LP-10³¹. The fluorescence intensity was measured at an excitation wavelength of 488 nm and an emission wavelength of 525 nm. For ROS detection, 20 mL algal cultures treated with strain LP-10 filtrates were centrifuged at 5000 g for 5 min, the pellets were washed twice with sterile *f/2* medium, then the algal cells were incubated with DCFH-DA (final concentration 10 μM) at 37°C for 30 min in the dark. The superfluous DCFH-DA probe was removed by centrifugation and the pellets were re-suspended in 500 μL sterile *f/2* medium after washing twice. The fluorescence intensity was measured using a microplate reader and the fluorescence photos of 5% LP-10 treatment was taken by an epifluorescence microscope.

N-acetylcysteine (NAC), a common ROS scavenger, was used to clear the ROS in algal cells. 20 mL algal cultures were processed using four treatments respectively: (1) 5% (v/v) strain LP-10 filtrates (LP treatment); (2) 5% filtrates and NAC (final concentration 1 mM) (NAC + LP treatment); (3) NAC (final concentration 1 mM) (NAC control); (4) blank 2216E broth (5%) (CK). The autofluorescence of the algal cells was measured in 96-well microplates at an excitation wavelength of 440 nm and an emission wavelength of 680 nm. Similar operations were performed with glutathione (GSH) and ascorbic acid (AsA), two other ROS scavengers. The ROS contents were determined at 0, 4, 8, 12, 24 h after addition of ROS scavengers.

The MDA content and antioxidant activities assays. Algal cultures of *P. globosa* were treated by cell-free filtrates of *Bacillus* sp. LP-10 in 3, 4, 5% concentration, followed by sampling at different

Gene name	Forward primer (5'-3')	Reverse primer (5'-3')
18S	TCCGATAACGAACGAGAC	TGACGCAAACCTTCCACTT
<i>psbA</i>	AGTTGCTGGTTCTCTACTTTACG	TTCCCACTCACGACCGATG
<i>rbcS</i>	AAGTCTTACTGGAAATGTGGG	AGCAGGACGCTGAACGATG

Table 1. The sequences of the primer pairs for RT-PCR analysis.

time course with an interval of 12 h. Algal cells were collected at 5000 g for 5 min, and the pellets were washed twice with sterile seawater. The cells were then re-suspended in 1 mL PBS solution (0.1 M, pH 7.4), followed by sonication at 100 W for 60 times. The cell debris was removed by centrifugation at 12000 g for 10 min, and the supernatants were transferred to new Eppendorf tubes for malondialdehyde (MDA) and antioxidants analysis. The MDA content and activities of the antioxidants, including superoxide dismutase (SOD), catalase (CAT), peroxidase (POD) and GSH, were measured using MDA kits and corresponding antioxidant assay kits (Nanjing Jiancheng Bioengineering Institute) following the manufacturer's instructions. The contents of MDA and antioxidant were expressed on a protein basis.

Chlorophyll contents and variable chlorophyll fluorescence analysis. Chlorophyll contents were measured based on previously described methods with slight modifications³². 20 mL axenic exponential phase cell cultures of *P. globosa* were collected at 5000 g for 10 min and the pellets were re-suspended in 5 mL 90% ethanol overnight to extract the chlorophyll. After extraction, the pellets were removed using centrifugation, the optical density of the ethanol extract was measured at wavelengths of 645 and 664 nm using a VARY-50 spectrophotometer. The contents of chlorophyll *a* (Chl *a*), chlorophyll *b* (Chl *b*), and the ratio of these two pigments were calculated as follows:

$$\text{Chl } a \text{ (mg}\cdot\text{L}^{-1}\text{)} = 12.7A_{664} - 2.69A_{645}$$

$$\text{Chl } b \text{ (mg}\cdot\text{L}^{-1}\text{)} = 22.9A_{645} - 4.68A_{664}$$

$$\text{Ratio of Chl } a \text{ and Chl } b = \text{Chl } a / \text{Chl } b$$

where A_{645} and A_{664} were the optical densities of ethanol extracts of algal cells at wavelengths of 645 and 664 nm.

The variable chlorophyll fluorescence was used as a proxy for the photosynthetic efficiency and capacity of the algal cell. And the variable chlorophyll fluorescence was measured using a PAM-CONTROL fluorometer (Walz, Effeltrich, Germany). The maximum quantum yield (Fv/Fm) and the maximum relative electron transport rate (rETR) were determined based on previously described methods after the algal cells were dark-adapted for 15 min³³.

Light condition assays. 20 mL algal cultures inoculated with 5% *Bacillus* sp. LP-10 filtrates, were respectively placed under different light conditions: the normal light intensity (normal light, 50 $\mu\text{mol photons m}^{-2}\text{s}^{-1}$), low light intensity (low light, 20 $\mu\text{mol photons m}^{-2}\text{s}^{-1}$) and the dark condition (dark). The autofluorescence of the algal cells was measured in 96-well microplates every 24 h at an excitation wavelength of 440 nm and an emission wavelength of 680 nm. The algal cultures without LP-10 filtrates under different light conditions were set up as control.

Gene transcription analysis. For total RNA extraction, 40 mL algal cultures were treated for 24 h and harvested at 5000 g for 10 min, then the pellets were quickly frozen in liquid nitrogen and homogenized followed by addition of 1 mL RNAiso reagent (TaKaRa Biochemicals, China). The total RNA was extracted following the manufacturer's instructions. Reverse transcription of RNA was performed using the PrimeScriptTM RT-PCR Kit (TaKaRa Biochemicals, China). Two photosynthesis genes (*psbA* and *rbcS*) were selected for real-time qPCR analysis. The real-time PCR was carried out using SYBR[®] Premix Ex TaqTM II (Tli RNaseH Plus) (TaKaRa Biochemicals, China). The 18S rRNA gene was used to standardize the results by eliminating variation in the quantity and quality. All of the primers used to amplify these genes were designed using software Primer premier 5.0 and the primer pairs are listed in Table 1. The RT-PCR program was: denaturation at 95 °C for 30 s, 40 cycles of 95 °C for 5 s and 60 °C for 30 s, followed by a melt curve program from 60 °C to 95 °C by 0.5 °C increases each 5 s. The relative gene expression abundance was quantified using the $2^{-\Delta\Delta C_t}$ method³⁴.

Protein extraction and immunodetection of protein D1. 20 mL algal cultures were centrifuged at 5000 g for 10 min, the pellets were washed twice with PBS solution and then re-suspended in 1 mL PBS solution. The suspension was then sonicated at 100 W for 60 times to lyse the algal cells. The debris

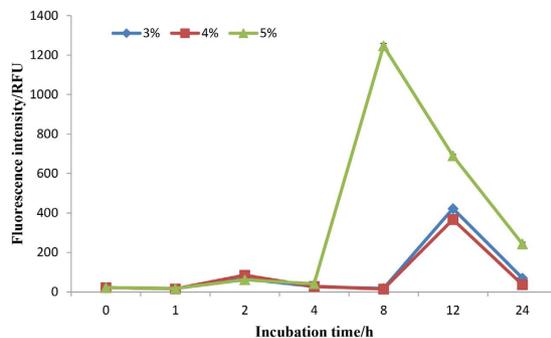


Figure 1. The ROS fluorescence intensity of algal cells under *Bacillus* sp. LP-10 treatment of different concentrations (3, 4, 5%).

was removed at 12000 g for 10 min and the supernatant were transferred to new Eppendorf tubes. The protein concentrations of the supernatant were determined using Bradford assays as mentioned above.

2 μ g total protein were separated by SDS-PAGE in a 12% polyacrylamide gel, the resolved protein was then transferred to PVDF membranes and probed with the antibody (Agriser) raised in rabbits against PSII reaction center protein D1. After incubation with secondary anti-rabbit IgG antibody coupled with horseradish peroxidase for 3 h, the membranes were reacted with ECL solution. Developed films were scanned with an Epson Expression 1680 Scanner²⁵.

Results

Effects of strain LP-10 on intracellular ROS content of algal cells. The intracellular ROS content of algal cells incubated with 3, 4 and 5% *Bacillus* sp. LP-10 filtrates showed an obvious variation during incubation time (Fig. 1). The fluorescence intensity of algal cells at all treatment concentrations was kept at a low level (< 100 RFU) in the early, then increased rapidly to a high level in all three treatment concentrations in 8 h (~1200 RFU) or 12 h (~400 RFU), respectively. Subsequently, the ROS levels of all treatments decreased to a low level, but still higher than the initial levels. The results shown in the fluorescence photos of algal cells treated by 5% filtrates were consistent with the fluorescence intensity determined by microplate reader (Fig. S1).

Effects of ROS scavengers addition on algal cell death. The impacts of addition of ROS scavengers NAC, GSH and AsA on algal cell death are shown in Fig. 2A. The addition of ROS scavengers NAC significantly influenced the algicidal activities of *Bacillus* sp. LP-10 on *P. globosa* cells. The addition of the LP-10 filtrates significantly lower the fluorescence intensity of algal cells, which was obviously less than that of the control (CK). When LP-10 treatment was added with NAC (NAC + LP), the fluorescence intensity of algal cells was much higher than that of the LP-10 treatment without NAC. And the fluorescence intensity of the NAC + LP treatment was approximate to the CK control at 24 h. However, the fluorescence intensity of the NAC + LP treatment was lower than the control at 48 h and 72 h, but still higher than the LP-10 treatment. The trends of addition of scavengers GSH and AsA showed similar pattern with the NAC.

In the ROS scavenger assays, the ROS contents of algal cells in each groups were determined and the results were presented in Fig. 2B. The results indicated that the ROS levels of the treatment groups supplemented with ROS scavengers was apparent lower than the LP-10 treatment. NAC, GSH and AsA could effectively remove the superfluous ROS in the algal cells.

Effects of strain LP-10 on the MDA content and antioxidant activities of algal cells. The MDA content increased significantly after the algal cells were treated with filtrates (Fig. S2), and the MDA content of the 4 and 5% treatment groups at 72 h were 10.78 and 29.53 times higher than that of the control. The MDA content of the 3% treatment group was also slightly higher than the control at 72 h, which implied that the addition of *Bacillus* sp. LP-10 filtrates promoted the lipid peroxidation of algal cells obviously.

SOD activities of the different treatment groups varied based on the concentrations of the LP-10 filtrates (Fig. 3A). All three treatments significantly enhanced SOD activities and the highest SOD activities of the 3, 4 and 5% treatment groups were 1.77-, 4.41- and 7.96-fold of the control. CAT activities shared a similar pattern with the SOD activities but with a rapid increase at the early phase (Fig. 3B). All three treatment groups showed a significant increase of CAT activities after 72 h incubation with LP-10 filtrates. POD activities of algal cells changed slower than CAT activities (Fig. 3C), and the 4 and 5% treatment groups were significant higher than the control. GSH is a type of non-enzymatic antioxidant and plays an important role in protecting cells from oxidative stress. Similarly, the addition of LP-10 filtrates also induced a rapid increase of GSH content in a short time (Fig. 3D), and the GSH content of the three treatment groups was 2.32, 2.47 and 2.93 times higher than that of the control after 72 h treatment.

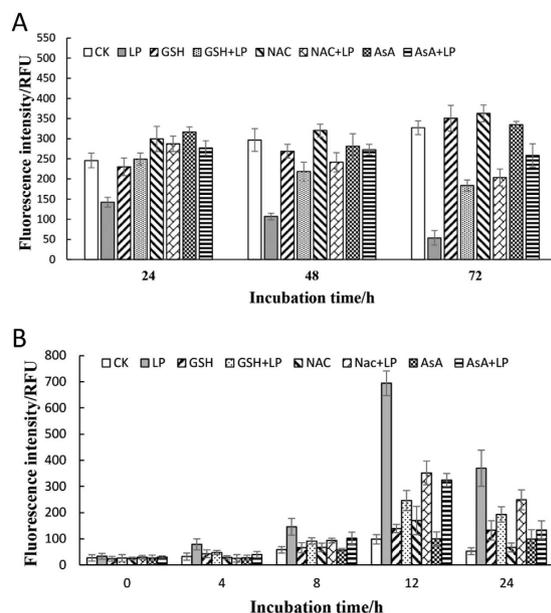


Figure 2. The algal autofluorescence intensity (A) and ROS fluorescence intensity (B) of algal cells under 5% *Bacillus* sp. LP-10 filtrates treatment with addition of ROS scavengers. CK represents algal cultures without any addition; LP represents algal cultures treated by 5% filtrates of *Bacillus* sp. LP-10; NAC represents algal cultures with addition of NAC; NAC + LP represents algal cultures with addition of NAC treated by 5% filtrates of *Bacillus* sp. LP-10. GSH and AsA are similar with those.

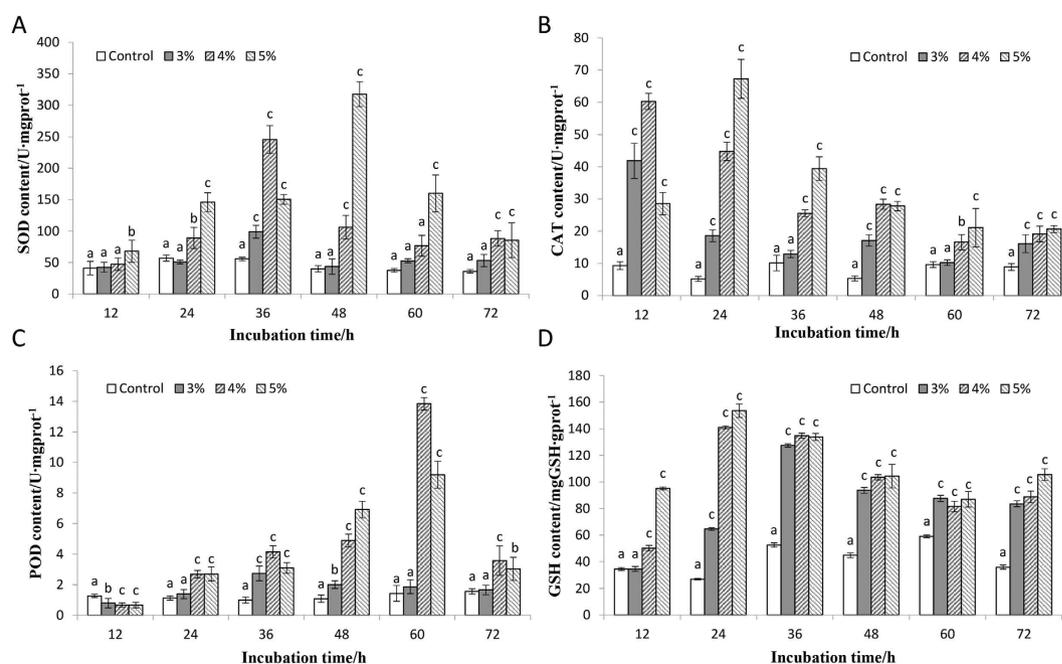


Figure 3. The SOD, CAT, POD and GSH activities of algal cells incubated with LP-10 filtrates of different concentrations (3, 4, and 5%). All data represent the means \pm S.D. (a) represents no significant difference, (b,c) represent statistically significant differences of $p < 0.05$ and $p < 0.01$ compared to the control respectively.

Effects of strain LP-10 on chlorophyll content and variable chlorophyll fluorescence. The contents of Chl *a* and *b* significantly decreased after algal cells were incubated with strain *Bacillus* sp. LP-10 filtrates (Fig. S3A,B). With the incubation time prolonged, decline of pigment contents showed the manner of concentration-dependent. The ratio of Chl *a* and Chl *b* were significantly impacted by the LP-10 filtrates (Fig. S3C). In the early phase, the ratio of the two pigments kept stable, but a rapid

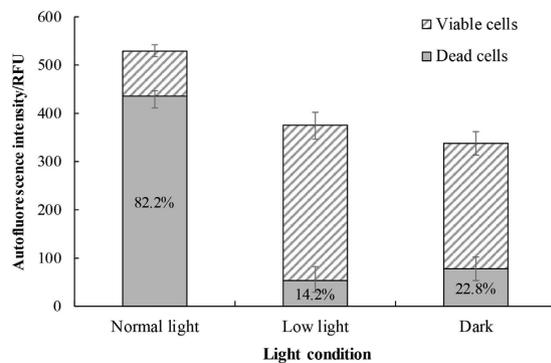


Figure 4. The autofluorescence intensity of algal cells under different light conditions after the algal cells were incubated with 5% LP-10 filtrates for 72 h. Normal light, low light and dark represent light intensities of 50, 20 and 0 $\mu\text{mol photons m}^{-2}\text{s}^{-1}$. The whole bar represents autofluorescence intensity of the control, while the twill part of the bar represents algal autofluorescence intensity of the 5% LP-10 treatment. And the gray part of the bar represents difference between the treatments and control, namely the dead cells under each light conditions.

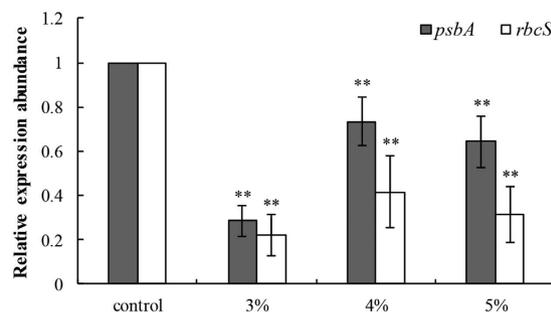


Figure 5. The relative transcriptional expression abundance of two genes (*psbA* and *rbcS*) in algal cells after incubation with LP-10 filtrates for 24 h. All data represent the means \pm S.D. **represents statistical significance at the $p < 0.01$ level.

decrease was observed after the algal cells were incubated with filtrates for more than 36 h, and higher concentration filtrates led to more significant decreases of the ratio. In the late phase, the ratio began to increase and gradually surpassed the control group. These results suggested that the sensitivity of the two pigments to strain LP-10 filtrates was different, and the pigment structures and balance were destroyed by the filtrates.

The photosynthetic efficiency measured from maximum quantum yields (F_v/F_m) showed an apparent decrease after the algal cells were treated with LP-10 filtrates. Higher concentration filtrates and longer treatment time led to more severe damage to the algal cells (Fig. S4A). The F_v/F_m value of the 5% treatment group at 48 h was approximate to 0.3, far less than the normal level. The maximum rETR, which was used to evaluate the maximum photosynthetic capacity, showed a similar change pattern with F_v/F_m (Fig. S4B) and the rETR of all three treatment groups significantly declined after the algal cells were treated with strain LP-10 filtrates.

Effects of light conditions on algal cells. The effects of different light conditions on the algicidal activities of strain LP-10 were presented in Fig. 4. The proportion of dead cells varied based on the light conditions. The algicidal activities of 5% LP-10 filtrates treatment under normal light were 82.2% at 72 h, while those of the low light and dark treatment groups were 14.2% and 22.8% respectively, which were significantly lower than that of the normal light treatment group.

Effects of strain LP-10 on the gene expression of algal cells. Genes *psbA* and *rbcS* play important roles in the photosynthesis of plant and algal cells. Gene *psbA* is responsible for the PSII reaction center protein D1, and gene *rbcS* codes for the small subunit of Rubisco, an important enzyme for CO_2 fixation. Our results demonstrated that the transcriptional expression abundance of *psbA* was 0.29, 0.73 and 0.64-fold of that of control at 24 h when treated by LP-10 filtrates (Fig. 5). And the transcriptional abundance of *rbcS* at 24 h shared a similar pattern with the *psbA* and all three concentrations treatments suppressed the gene expression of *rbcS*.

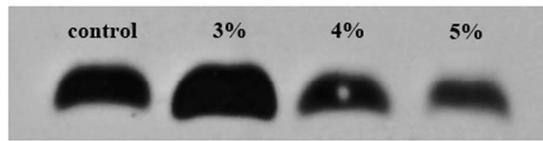


Figure 6. Immunodetection of the PSII reaction center protein D1 after the algal cells were incubated with LP-10 filtrates for 24 h. Control represents the algal cells without addition of LP-10; 3, 4, 5% represent the algal cells treated by different concentrations of LP-10 filtrates respectively.

Effects of strain LP-10 on the expression of PSII reaction center protein D1. The expression of PSII reaction center protein D1 was detected in this study (Fig. 6). The D1 content of the 3% treatment group was higher than that of the control while the content of the 4 and 5% treatment groups was remarkably decreased. The D1 content of the 3, 4 and 5% treatment groups was 169, 71 and 43% of the control, based on the quantification of band intensities using densitometry with Quantity-One software.

Discussion

Marine disasters bring about huge losses to humans every year, and red tides were one of the most notorious calamities. However, oceans also provide huge resources for us as a library which makes it possible to utilize marine microorganisms to regulate the HABs. Marine algicidal bacteria are suggested to be excellent candidates in managing HABs^{8,10}. Up to now, many bacteria have been isolated to be effective in removing harmful algae, but the action mechanism of algicidal bacteria on toxic algae has not been well explained. In this study, we investigated the mode of action of a marine algicidal bacterium *Bacillus* sp. LP-10, which was previously isolated in our laboratory from the surface seawater samples of the East China Sea.

As is known, ROS are common byproducts of aerobic metabolism, predominantly produced in chloroplasts and mitochondria of algal cells¹⁹. Normally, the antioxidant system of algal cells could remove the superfluous ROS to keep the cells from oxidative damage. However, environmental stresses often induce algal cells an unusual rise of ROS level. Liu *et al.* found the addition of allelochemical florfenicol significantly enhanced the intracellular ROS content of *Skeletonema costatum* at 6 h³⁵. Similarly, an obvious increase of ROS was observed in our study when strain LP-10 was inoculated into the algal cultures, which indicated that the algal cells were suffering from oxidative stress. And the MDA content rise also confirmed the oxidative damage in the algal cells.

NAC is a commonly used ROS scavenger, and our results demonstrated that the addition of NAC to the algal cultures significantly improved the survival rate of algal cells compared to algal cells treated by LP-10 filtrates without NAC. And the ROS content determination in ROS scavenger assays hinted that the superfluous ROS was really removed by the ROS scavengers. The tested results of two other antioxidants (GSH and AsA) showed similar consequences and confirmed that the superfluous ROS was reacted with the scavengers rather than surviving the algal cells by other means. Furthermore, the autofluorescence intensity of algal cells in NAC + LP treatment group was approximate to the CK and NAC control groups, suggesting that the ROS was the main reason for algal cell death.

MDA content was an indicator of lipid peroxidation in algal cells. Our results suggested the MDA content were keeping rising during the experiment time, and the rate of lipid peroxidation became more and more serious, implying the antioxidant system of algal cells may not clear the excessive ROS in time and the algal cells suffered from oxidative damage for long. Other reported isolates were also proved to cause the increase of MDA content in algal cells^{15,36}.

Algal cells execute programs to clear the excess ROS via antioxidant systems²¹. Our results showed that addition of LP-10 enhanced SOD, CAT, POD and GSH activities of algal cells, the activities of these four antioxidants at 72 h were significantly higher than the initial levels, suggesting algal cells were kept under the oxidative stress and superfluous ROS could not be cleared in time to resume the normal status. Excess ROS triggered a series of damage processes in algal cells²², leading the dysfunction of many fundamental organelles and structures. MDA assays also revealed that algal cells were under continuous oxidative stress as discussed above, which will oxidize the lipids of membranes to MDA, thus the membrane permeability changed, and the algal cells finally disrupted for dysfunction of algal cells.

Environmental factors often affected important physiological functions of algal cells^{23,26}. Domingues *et al.* investigated the response of *Phaeodactylum tricornerutum* to high light exposure, the results showed photosynthetic activities of algal cells declined obviously²⁵. In this study, addition of *Bacillus* sp. LP-10 filtrates significantly impacted the fundamental functions of algal cells. Among which, photosynthesis systems are vulnerable to oxidative stress. Photosynthesis pigments are key components of photosynthesis systems, which are responsible for light harvesting and photosynthetic reaction³⁷. When LP-10 filtrates were inoculated into the algal cultures, an obvious decrease of Chl *a* and *b* contents was observed, and the ratio of these two pigments was significantly changed, suggesting the pigment structures were destroyed, scanning electron microscope (SEM) and transmission electron microscope (TEM) images of algal cells under treatment also confirmed the results (Figs S5, S6). The data of electron microscope indicated the

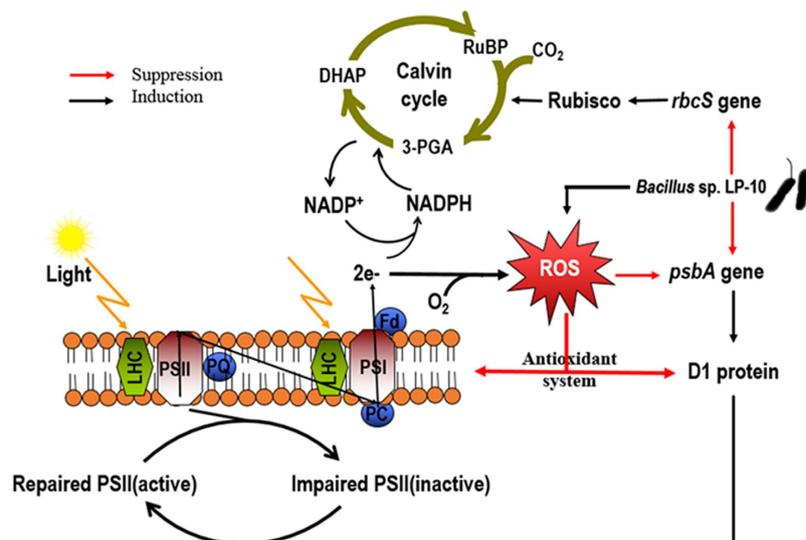


Figure 7. A hypothetical scheme for the algicidal mechanism of the algal cells upon *Bacillus sp. LP-10*.

thylakoid of the algal cells became sparse and structure of the chloroplast structures were changed. The unstable ratio of Chl *a* and *b* contents may be the consequence of sensitivity difference of these two pigments under strain LP-10 stress. The low ratio in the early phase indicates that photosynthetic antenna complexes are more vulnerable to oxidative stress than light-harvesting complexes¹⁷.

The Fv/Fm and maximum rETR are two important parameters used to evaluate the photosynthesis efficiency and capacity³⁸, which were usually disturbed by environmental stresses. Domingues *et al.* found high light exposure will lead declination of the Fv/Fm of algal cells²⁵. Our results revealed that the value of Fv/Fm significantly decreased when the algal cells were under strain LP-10 stress, suggesting the photosynthetic efficiency was hindered by strain LP-10. Similarly, the rETR was also significantly inhibited by strain LP-10, implying that the photosynthetic capacity of algal cells decreased after treatment with strain LP-10. These results indicated that the photosynthesis systems of algal cells were attacked by the active metabolites produced by *Bacillus sp. LP-10*. Combined with the pigment assays, we speculated that strain LP-10 would destroy the structures of photosynthetic system and lower the photosynthesis activities, causing the dysfunction of the photosynthetic system at last.

More than that, *Bacillus sp. LP-10* also inhibited the transcriptional expression of many important genes. Wu *et al.* noted that allelochemical pyrogallol significantly affects the expression of genes *psbA*, *grpE*, *fabZ*, *recA*, *cmpA*, *ftsZ* and *cyrJ*³⁹. In our study, two photosynthesis-related genes (*psbA* and *rbcS*) were investigated. At 24 h, transcriptional expression of both gene *psbA* and *rbcS* were down-regulated. We speculated that the active metabolites produced by strain LP-10 may affected the transcription processes or severe impairment occurred in the genetic molecules of the cells at the time. Gene *psbA* was responsible for synthesis of protein D1, one of the subunits of the reaction center of PSII. Impairment of protein D1 will decrease the photosynthetic capacity, leading to the photoinhibition of the algal cells²⁴. The gene *rbcS* code for the small subunit of Rubisco, which plays a critical role in CO₂ fixation and photorespiration⁴⁰. The inhibition of the *rbcS* in the algal cells implied that the CO₂ fixation cycle (Calvin cycle) was interrupted by *Bacillus sp. LP-10*, which would accelerate the photoinhibition of the algal cells⁴¹, because limitation of CO₂ fixation will decrease the absorption capacity for solar energy of the algal cells and affect the electron transportation of photosystems. The excess energy causes the production of numerous ROS at last, which inhibits the synthesis of protein D1²³.

Our study also found that light was an indispensable factor in algal cell death. The algicidal activity of strain LP-10 under low light was very low (~15%) while normal light led to a higher algicidal activity (~80%), which meant the light intensity of “normal light” was too high for the stressed algal cells and the stressed algal cells could not utilize the “normal light”, so that the relative high light leads to photoinhibition of the algal cells, which indirectly revealed that strain LP-10 impacted the CO₂ fixation and the utilization of solar energy. The results implied that dysfunction of the photosynthesis system (photoinhibition) was the most fundamental reason for algal cell death. Qian *et al.* and Yang *et al.* reported that the allelochemicals N-phenyl-2-naphthylamine and hydroquinone could cause photoinhibition of *Chlorella vulgaris* and *Phaeodactylum tricorutum*^{17,18}. Though some other reasons were able to cause the algal cell death, such as destroying the cell structures, our results demonstrated the role of other factors were minor reasons. Because when the algal cultures were placed in the dark or under low light conditions, the algicidal rate was very low (~15%).

The western blot analysis of D1 protein showed that the D1 protein content of algal cells at 4 and 5% treatment concentrations of strain LP-10 decreased obviously at 24 h. This results were consistent with the

gene expression profiles of *psbA*, which confirmed that photoinhibition was taken place in the algal cells. Photoinhibition of algal cells indicated the photosynthesis would be hampered, and energy-consuming functions would be inhibited. Long-time photoinhibition would be unable to support the survival of the algal cells, leading to the algal cells death eventually.

From the above, we conclude that algicidal bacterium *Bacillus* sp. LP-10 induce a significant increase of ROS in algal cells, and continuous stressing make the superfluous ROS not being cleared in time, which disturb the physiological functions and destruct the structures of algal cells. More important, excess ROS lead to photoinhibition of algal cells by influencing the expression of the photosynthesis-related genes and protein, and impaired photosystem apparatus cannot be repaired, leading to the dysfunction and causing disruption of algal cells eventually (Fig. 7).

References

- Landsberg, J. H. The effects of harmful algal blooms on aquatic organisms. *Reviews in Fisheries Science* **10**, 113–390 (2002).
- Hoagland, P. & Scatista, S. *Ecology of harmful algae*. 391–402 (Springer, 2006).
- Van Dolah, F. M. Marine algal toxins: origins, health effects, and their increased occurrence. *Environmental health perspectives* **108**, 133 (2000).
- Anderson, D. M. Turning back the harmful red tide. *Nature* **388**, 513–514 (1997).
- Zingone, A. & Oksfeldt Enevoldsen, H. The diversity of harmful algal blooms: a challenge for science and management. *Ocean & Coastal Management* **43**, 725–748 (2000).
- Sigeo, D. *et al.* *The Ecological Bases for Lake and Reservoir Management*. 161–172 (Springer, 1999).
- Gumbo, R. J., Ross, G. & Cloete, E. T. Biological control of *Microcystis* dominated harmful algal blooms. *African Journal of Biotechnology* **7**, 4765–4773 (2008).
- Cole, J. J. Interactions between bacteria and algae in aquatic ecosystems. *Annual Review of Ecology and Systematics* **13**, 291–314 (1982).
- Doucette, G. J. Interactions between bacteria and harmful algae: a review. *Natural toxins* **3**, 65–74 (1995).
- Mayali, X. & Azam, F. Algicidal bacteria in the sea and their impact on algal blooms. *Journal of Eukaryotic Microbiology* **51**, 139–144 (2004).
- Ma, H. *et al.* Mode of action of membrane-disruptive lytic compounds from the marine dinoflagellate *Alexandrium tamarense*. *Toxicon* **58**, 247–258 (2011).
- Lee, S.O. *et al.* Involvement of an Extracellular Protease in Algicidal Activity of the Marine Bacterium *Pseudoalteromonas* sp. Strain A28. *Applied and Environmental Microbiology* **66**, 4334–4339 (2000).
- Prince, E. K., Poulson, K. L., Myers, T. L., Sieg, R. D. & Kubanek, J. Characterization of allelopathic compounds from the red tide dinoflagellate *Karenia brevis*. *Harmful Algae* **10**, 39–48 (2010).
- Kim, D. *et al.* Red to red—the marine bacterium *Hahella chejuensis* and its product prodigiosin for mitigation of harmful algal blooms. *Journal of microbiology and biotechnology* **18**, 1621–1629 (2008).
- Zhang, H. *et al.* Effect of Oxidative Stress Induced by *Brevibacterium* sp. BS01 on a HAB Causing Species-*Alexandrium tamarense*. *PLoS one* **8**, e63018, doi: 10.1371/journal.pone.0063018 (2013).
- Li, Y. *et al.* Towards molecular, physiological, and biochemical understanding of photosynthetic inhibition and oxidative stress in the toxic *Alexandrium tamarense* induced by a marine bacterium. *Applied microbiology and biotechnology* **98**, 4637–4652 (2014).
- Qian, H. *et al.* Allelochemical stress causes oxidative damage and inhibition of photosynthesis in *Chlorella vulgaris*. *Chemosphere* **75**, 368–375 (2009).
- Yang, C. *et al.* Allelochemical induces growth and photosynthesis inhibition, oxidative damage in marine diatom *Phaeodactylum tricoratum*. *Journal of Experimental Marine Biology and Ecology* **444**, 16–23 (2013).
- Apel, K. & Hirt, H. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.* **55**, 373–399 (2004).
- Atkinson, N. J. & Urwin, P. E. The interaction of plant biotic and abiotic stresses: from genes to the field. *Journal of experimental botany* **63**, 3523–3543 (2012).
- Yu, B. P. Cellular defenses against damage from reactive oxygen species. *Physiological reviews* **74**, 139 (1994).
- Avery, Simon V. Molecular targets of oxidative stress. *Biochemical Journal* **434**, 201–210 (2011).
- Takahashi, S. & Murata, N. How do environmental stresses accelerate photoinhibition. *Trends in Plant Science* **13**, 178–182 (2008).
- Takahashi, S. & Badger, M. R. Photoprotection in plants: a new light on photosystem II damage. *Trends in Plant Science* **16**, 53–60 (2011).
- Domingues, N., Matos, A. R., Marques da Silva, J. & Cartaxana, P. Response of the Diatom *Phaeodactylum tricoratum* to Photooxidative Stress Resulting from High Light Exposure. *PLoS One* **7**, e38162, doi: 10.1371/journal.pone.0038162 (2012).
- Murata, N., Takahashi, S., Nishiyama, Y. & Allakhverdiev, S. Photoinhibition of photosystem II under environmental stress. *Biochimica et Biophysica Acta (BBA) - Bioenergetics* **1767**, 414–421 (2007).
- Schoemann, V., Becquevort, S., Stefels, J., Rousseau, V. & Lancelot, C. *Phaeocystis* blooms in the global ocean and their controlling mechanisms: a review. *Journal of Sea Research* **53**, 43–66 (2005).
- Guan, C. *et al.* Novel algicidal evidence of a bacterium *Bacillus* sp. LP-10 killing *Phaeocystis globosa*, a harmful algal bloom causing species. *Biological Control* **76**, 79–86 (2014).
- Guillard, R. R. In *Culture of marine invertebrate animals* 29–60 (Springer, 1975).
- Schut, F. *et al.* Isolation of typical marine bacteria by dilution culture: growth, maintenance, and characteristics of isolates under laboratory conditions. *Applied and environmental microbiology* **59**, 2150–2160 (1993).
- LeBel, C. P., Ischiropoulos, H. & Bondy, S. C. Evaluation of the probe 2', 7'-dichlorofluorescein as an indicator of reactive oxygen species formation and oxidative stress. *Chemical research in toxicology* **5**, 227–231 (1992).
- Marr, I. L., Suryana, N., Lukulay, P. & Marr, M. I. Determination of chlorophyll a and b by simultaneous multi-component spectrophotometry. *Fresenius' journal of analytical chemistry* **352**, 456–460 (1995).
- Grebe, M. *et al.* An Integrated Analysis of Molecular Acclimation to High Light in the Marine Diatom *Phaeodactylum tricoratum*. *PLoS One* **4**, e7743 (2009).
- Livak, K. J. & Schmittgen, T. D. Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the 2^{-ΔΔCT} Method. *Methods* **25**, 402–408 (2001).
- Liu, W., Ming, Y., Huang, Z. & Li, P. Impacts of florfenicol on marine diatom *Skeletonema costatum* through photosynthesis inhibition and oxidative damages. *Plant Physiology and Biochemistry* **60**, 165–170 (2012).

36. Kong, Y., Xu, X. & Zhu, L. Cyanobactericidal Effect of *Streptomyces* sp. HJC-D1 on *Microcystis aeruginosa*. *PLoS One* **8**, e57654, doi: 10.1371/journal.pone.0057654 (2013).
37. Rowan, K. S. *Photosynthetic pigments of algae* (CUP Archive, 1989).
38. Schreiber, U., Bilger, W. & Neubauer, C. In *Ecophysiology of photosynthesis* 49–70 (Springer, 1995).
39. Wu, Z., Shi, J. & Yang, S. The effect of pyrogallol acid on growth, oxidative stress, and gene expression in *Cylindrospermopsis raciborskii* (Cyanobacteria). *Ecotoxicology* **22**, 271–278 (2012).
40. Spreitzer, R. J. & Salvucci, M. E. Rubisco: structure, regulatory interactions, and possibilities for a better enzyme. *Annual review of plant biology* **53**, 449–475 (2002).
41. Takahashi, S. & Murata, N. Interruption of the Calvin cycle inhibits the repair of Photosystem II from photodamage. *Biochim Biophys Acta* **1708**, 352–61 (2005).

Acknowledgements

This work was financially supported by the Joint Project of the National Natural Science Foundation of China and Shandong Province: Marine ecology and environmental sciences (U1406403), the National Natural Science Foundation of China (41376119, 41576109, 40930847), Mr. An Tao is thanked for his help with the western blot techniques. We would like to thank Prof. I. J. Hodgkiss from The University of Hong Kong for help with English.

Author Contributions

Conceived and designed the experiments: C.G., X.G., Z.Y. and T.Z. Performed the experiments: C.G., X.G., Y.L., H.Z., X.L., G.C., J.G. and Z.Y. Analyzed the data: C.G., X.G. and T.Z. Wrote the paper: C.G., X.G. and T.Z.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Guan, C. *et al.* Photoinhibition of *Phaeocystis globosa* resulting from oxidative stress induced by a marine algicidal bacterium *Bacillus* sp. LP-10. *Sci. Rep.* **5**, 17002; doi: 10.1038/srep17002 (2015).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>