

# AI-2 quorum-sensing inhibitors affect the starvation response and reduce virulence in several *Vibrio* species, most likely by interfering with LuxPQ

Gilles Brackman,<sup>1</sup> Shari Celen,<sup>2</sup> Kartik Baruah,<sup>3</sup> Peter Bossier,<sup>3</sup>  
Serge Van Calenbergh,<sup>2</sup> Hans J. Nelis<sup>1</sup> and Tom Coenye<sup>1</sup>

## Correspondence

Tom Coenye

Tom.Coenye@UGent.be

<sup>1</sup>Laboratory of Pharmaceutical Microbiology, Ghent University, Harelbekestraat 72, B-9000 Ghent, Belgium

<sup>2</sup>Laboratory of Medicinal Chemistry, Ghent University, Harelbekestraat 72, B-9000 Ghent, Belgium

<sup>3</sup>Laboratory of Aquaculture and Artemia Reference Center, Ghent University, Rozier 44, 9000 Ghent, Belgium

The increase of disease outbreaks caused by *Vibrio* species in aquatic organisms as well as in humans, together with the emergence of antibiotic resistance in *Vibrio* species, has led to a growing interest in alternative disease control measures. Quorum sensing (QS) is a mechanism for regulating microbial gene expression in a cell density-dependent way. While there is good evidence for the involvement of auto-inducer 2 (AI-2)-based interspecies QS in the control of virulence in multiple *Vibrio* species, only few inhibitors of this system are known. From the screening of a small panel of nucleoside analogues for their ability to disturb AI-2-based QS, an adenosine derivative with a *p*-methoxyphenylpropionamide moiety at C-3' emerged as a promising hit. Its mechanism of inhibition was elucidated by measuring the effect on bioluminescence in a series of *Vibrio harveyi* AI-2 QS mutants. Our results indicate that this compound, as well as a truncated analogue lacking the adenine base, block AI-2-based QS without interfering with bacterial growth. The active compounds affected neither the bioluminescence system as such nor the production of AI-2, but most likely interfered with the signal transduction pathway at the level of LuxPQ in *V. harveyi*. The most active nucleoside analogue (designated LMC-21) was found to reduce the *Vibrio* species starvation response, to affect biofilm formation in *Vibrio anguillarum*, *Vibrio vulnificus* and *Vibrio cholerae*, to reduce pigment and protease production in *V. anguillarum*, and to protect gnotobiotic *Artemia* from *V. harveyi*-induced mortality.

Received 7 July 2009

Revised 23 September 2009

Accepted 23 September 2009

## INTRODUCTION

*Vibrio* species are ubiquitous in marine environments worldwide (Igbino & Okoh, 2008). As opportunistic pathogens they can cause mild to severe infections in humans and marine animals. Vibriosis is one of the most prevalent fish diseases, mainly caused by *Vibrio anguillarum*, *Vibrio alginolyticus*, *Vibrio parahaemolyticus*, *Vibrio harveyi* and *Vibrio campbellii* (Garcia *et al.*, 1997; Austin & Zhang, 2006). Other *Vibrio* species are pathogenic for humans. *Vibrio vulnificus* is associated with gastrointestinal infections primarily following the consumption of raw and undercooked seafood, but it can also cause wound or soft-tissue infections (Bross *et al.*, 2007). In addition,

systemic *V. vulnificus* infections are notorious for their high mortality rate (Chiang & Chuang, 2003). *Vibrio cholerae* is responsible for pandemic and epidemic outbreaks of cholera (Griffith *et al.*, 2006). *V. cholerae* serotype O1 causes the majority of the outbreaks worldwide, while the O139 serotype has only been detected in South-East and East Asia (Sack *et al.*, 2004; Griffith *et al.*, 2006). Cell-cell communication (quorum sensing; QS) in *Vibrio* species plays an important role in virulence. QS in *Vibrio* species involves three types of signal molecules. *N*-Acylhomoserine lactones (AHLs) are used in the LuxM/N QS system, cholera auto-inducer 1 (CAI-1) in the CqsA/S system and auto-inducer 2 (AI-2) in the LuxS/PQ QS system (Bassler *et al.*, 1993, 1997; Higgins *et al.*, 2007; Ryan & Dow, 2008). AI-2 is synthesized starting from *S*-adenosylmethionine, which (through a series of enzymatic reactions, including the reaction catalysed by LuxS) is converted to 4,5-dihydroxy-2,3-pentanedione (DPD) (Surette *et al.*, 1999; Winzer *et al.*, 2002). The spontaneous

Abbreviations: AI-2, auto-inducer 2; CV, crystal violet; DPD, 4,5-dihydroxy-2,3-pentanedione; MCPBA, 4-methoxycarbonyl-phenylboronic acid; QS, quorum sensing; QSI, QS inhibiting.

A set of supplementary methods, three supplementary figures and four supplementary tables are available with the online version of this paper.

cyclization of DPD followed by esterification with a tetrahydroxyborate anion results in the formation of AI-2 (Miller *et al.*, 2004). In *Vibrio* species, sensing of extracellular AI-2 involves two proteins, LuxP and LuxQ (Chen *et al.*, 2002). At a low AI-2 concentration, LuxQ will be autophosphorylated, resulting in the transfer of a phosphate group to LuxO via LuxU (Freeman & Bassler, 1999a, b). Phosphorylation of LuxO leads to its activation and the production of small regulatory RNAs. These small RNAs, together with the chaperone protein Hfq, destabilize mRNA of the response regulator LuxR. In the absence of AI-2, LuxR is not produced and LuxR-dependent genes are not transcribed. Binding of AI-2 to the LuxPQ complex initiates a switch from kinase to phosphatase activity, which results in the dephosphorylation of the downstream proteins LuxU and LuxO. Dephosphorylated LuxO is inactive and does not induce the production of small regulatory RNAs. Hence, the response regulator LuxR is produced and initiates transcription of target genes, including several virulence genes. Therefore, QS inhibitors are promising antipathogenic agents. Due to the presence of the *luxS* gene in diverse bacterial species, AI-2 is considered to be a signal for interspecies communication (Xavier & Bassler, 2003). However, the LuxPQ signal transduction system is restricted to Vibrionales (Sun *et al.*, 2004; Rezzonico & Duffy, 2008). The increase of *Vibrio* disease outbreaks in aquatic organisms as well as in humans (Harvell *et al.*, 2002; Boyd *et al.*, 2008; Kapp, 2009), together with the emergence of antibiotic resistance in *Vibrio* species (Karunasagar *et al.*, 1994; Scarscia *et al.*, 2006), has resulted in a growing interest in alternative disease control measures (Lynch & Wiener-Kronisch, 2008). A novel approach consists of interfering with bacterial communication (Ni *et al.*, 2009). Several cinnamaldehyde and furanone derivatives disrupt AI-2-based QS in *Vibrio* species by decreasing the DNA-binding activity of the response regulator LuxR, and are active both *in vitro* and *in vivo* (Defoirdt *et al.*, 2006, 2007; Brackman *et al.*, 2008). Other compounds, including *S*-anhydroribosyl-L-homocysteine and *S*-homoribosyl-L-cysteine, block the production of AI-2 by inhibiting the key enzyme LuxS (Alfaro *et al.*, 2004; Shen *et al.*, 2006). Based on the concept of molecular mimicry and through virtual screenings using the crystal structure of LuxP, new AI-2 QS inhibitors have been discovered (Li *et al.*, 2008; Ni *et al.*, 2008a, b). However, although these compounds affect bioluminescence in *V. harveyi*, they were evaluated neither for their effect on QS-regulated virulence factors nor for their activity *in vivo*. The goal of the present study was to test whether previously described AI-2 QS inhibitors targeting LuxPQ and various compounds from our collection have the ability to block the production of QS-regulated virulence factors in *Vibrio* species.

## METHODS

**Bacterial strains and growth conditions.** All bacterial strains used in this study are listed in Table 1. They were cultured in Marine-Broth

(MB) (BD) in the presence of appropriate antibiotics at 30 °C with shaking, except for *Escherichia coli* DH5 $\alpha$  and *E. coli* K-12, which were grown in Luria–Bertani broth (LB) (BD) at 30 °C and 37 °C, respectively, without shaking.

**Compound library.** The compounds used in the present study consisted of a selected set of known AI-2 QS inhibitors, supplemented with a series of nucleoside (mainly adenosine) analogues (Fig. 1). 3'-Azido-(3'-N<sub>3</sub>-3'-dA) and 3'-amino-3'-deoxyadenosine (3'-NH<sub>2</sub>-3'-dA) were prepared as reported by Azhayeve & Smrt (1978), and are nowadays also commercially available. For the synthesis of the amide analogues derived from 3'-NH<sub>2</sub>-3'-dA (i.e. LMC-23, LMC-20, IK-1, LMC-21, LMC-27 and LMC-28), we followed the procedure described by Soenens *et al.* (1995). Briefly, the 3'-amino group of unprotected 3'-NH<sub>2</sub>-3'-dA was acylated with the appropriate carboxylic acids using dicyclohexylcarbodiimide (DCC) or diisopropylcarbodiimide (DIC) and *N*-hydroxysuccinimide (NHS) as coupling agents in a mixture of dimethylformamide (DMF) and dichloromethane (see Supplementary Methods and Supplementary Fig. S1). The synthetic route followed for the synthesis of the 3' branched-chain analogue SC-23, differing from that for LMC-21 by the insertion of a CH<sub>2</sub> group between C-3' of the ribofuranose ring and the amide moiety started from the previously described intermediate 1 (Kim *et al.*, 2003) (see Supplementary Methods and Supplementary Fig. S2). The 2-modified adenosine analogues LMC-29, LMC-30 and LMC-35 have been recently synthesized and found to be potent adenosine A<sub>3</sub> receptor antagonists/partial agonists (Cosyn *et al.*, 2006), while PVR-121 is an agonist for the same receptor (Ohno *et al.*, 2004). The amides derived from 3-(4-methoxyphenyl)propanoic acid (i.e. SC-1, SC-2 and SC-3) were prepared by ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC)-mediated coupling of the parent carboxylic acid with the appropriate amine in the presence of *N,N,N*-triethylamine (TEA). The synthesis of SC-20 started from the known sugar intermediate 4 (see Supplementary Methods and Supplementary Fig. S3) that was converted to methyl glycoside (intermediate 5) upon reaction with SnCl<sub>4</sub> and dry methanol (Moradei *et al.*, 1991). Remarkably, during this reaction, a larger amount of intermediate 6 was formed. The reaction mixture could be efficiently separated by flash chromatography, and 5 and 6 were separately deprotected upon treatment with NH<sub>3</sub> in methanol, thereby yielding 7.1 and 7.2. NMR analysis revealed that 7.1 and 7.2 differ only at the anomeric position ( $\alpha$ - or  $\beta$ -keto group). Although the configuration of each anomer remains uncertain, we anticipate that 7.1 represents the  $\beta$ -anomer, since it was formed from 5, which still possessed the participating acetate group at C-2. Subsequently, we continued with azide 7.2 (presumably the  $\alpha$ -anomer), which was reduced through a Staudinger reduction. Finally, the resulting amine was coupled to 3-(4-methoxyphenyl)propanoic acid using *N,N,N',N'*-tetramethyl-*O*-(6-chloro-1H-benzotriazol-1-yl)uronium hexafluorophosphate (HCTU) as the coupling agent. All synthesized compounds were structurally confirmed using <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy and exact mass measurements (see Supplementary Methods) and were shown to possess a purity of at least 95% by combustion analysis. The previously described AI-2 QS inhibitor 2-(2-thienylsulfonyl)ethanethioamide (KM-03009) (Li *et al.*, 2008) was purchased from Acros Organics, while pyrogallol (Ni *et al.*, 2008a) and 4-methoxycarbonyl-phenylboronic acid (MCPBA) (Ni *et al.*, 2008b) were purchased from Sigma-Aldrich. If necessary, compounds were diluted in DMSO (final concentration 0.5%, v/v). The stock solutions were stored at -20 °C. Control solutions contained the same amount of DMSO.

**Determination of MICs.** MICs were determined for each compound by using a microdilution assay, as previously described (Brackman *et al.*, 2009). MB and LB media were used for all *Vibrio* species and both *E. coli* strains, respectively. The plates were incubated and the

**Table 1.** Strains used in this study

BCCM/LMG, Belgian Co-ordinated Collections/Laboratory of Microbiology collection (Ghent University, Belgium); HPACC, Health Protection Agency Culture collection.

Strain	Additional information	Reference or source
<b><i>V. harveyi</i> strains</b>		
BB120	Wild-type from which strains BB170, BB886, MM30, JAF553, JAF483, BNL258, JAF375 and JMH597 are derived	Bassler <i>et al.</i> (1997)
BB170	<i>luxN</i> ::Tn5	Bassler <i>et al.</i> (1993)
BB886	<i>luxPQ</i> ::Tn5 Kan <sup>R</sup>	Bassler <i>et al.</i> (1994)
MM30	<i>luxS</i> ::Tn5	Surette <i>et al.</i> (1999)
JAF553	<i>luxU</i> H58A linked to Kan <sup>R</sup>	Freeman & Bassler (1999a)
JAF483	<i>luxO</i> D47A linked to Kan <sup>R</sup>	Freeman & Bassler (1999b)
BNL258	<i>hfq</i> ::Tn5 <i>lacZ</i>	Lenz <i>et al.</i> (2004)
JAF375	<i>luxN</i> ::Cm <sup>R</sup> <i>luxQ</i> ::Kan <sup>R</sup>	Freeman & Bassler (1999b)
JMH597	<i>luxN</i> ::Tn5 <i>cqsS</i> ::Cm <sup>R</sup>	Defoirdt <i>et al.</i> (2006)
<b><i>V. anguillarum</i> strain</b>		
LMG4411	Isolated from young sea trout ( <i>Salmo trutta</i> )	BCCM/LMG
<b><i>V. campbellii</i> strain</b>		
LMG21363	Isolated from <i>Penaeus monodon</i> juvenile, lymphoid organ	BCCM/LMG
<b><i>V. cholerae</i> strain</b>		
NCTC8457	Isolated from human, biotype El Tor	HPACC
<b><i>V. vulnificus</i> strain</b>		
LMG16867	Isolated from tank water on eel farm	BCCM/LMG
<b><i>E. coli</i> strains</b>		
DH5 $\alpha$ pBlueLux	Strain (not producing AI-2) containing pBlueLux polylinker and <i>luxCDABE</i> genes	Brackman <i>et al.</i> (2008)
K12	AI-2-producing strain	Ren <i>et al.</i> (2004)

OD<sub>590</sub> was measured after 24 h using a Victor Wallac<sup>2</sup> multilabel counter (Perkin Elmer Life and Analytical Sciences).

**Identification of the molecular target of the QS inhibitors.** The assay for the effect on constitutively expressed bioluminescence (using *E. coli* DH5 $\alpha$  containing the pBlueLux plasmid), the bioassay for LuxS inhibition (using *V. harveyi* MM30) and assays to determine the molecular target of the compounds tested (using *V. harveyi* BB120, BB170, BB886, JAF375, JAF553, JAF483, JMH597 and BNL258) were conducted as described previously (Brackman *et al.*, 2008). Each compound was tested at least six times in triplicate ( $n \geq 18$ ).

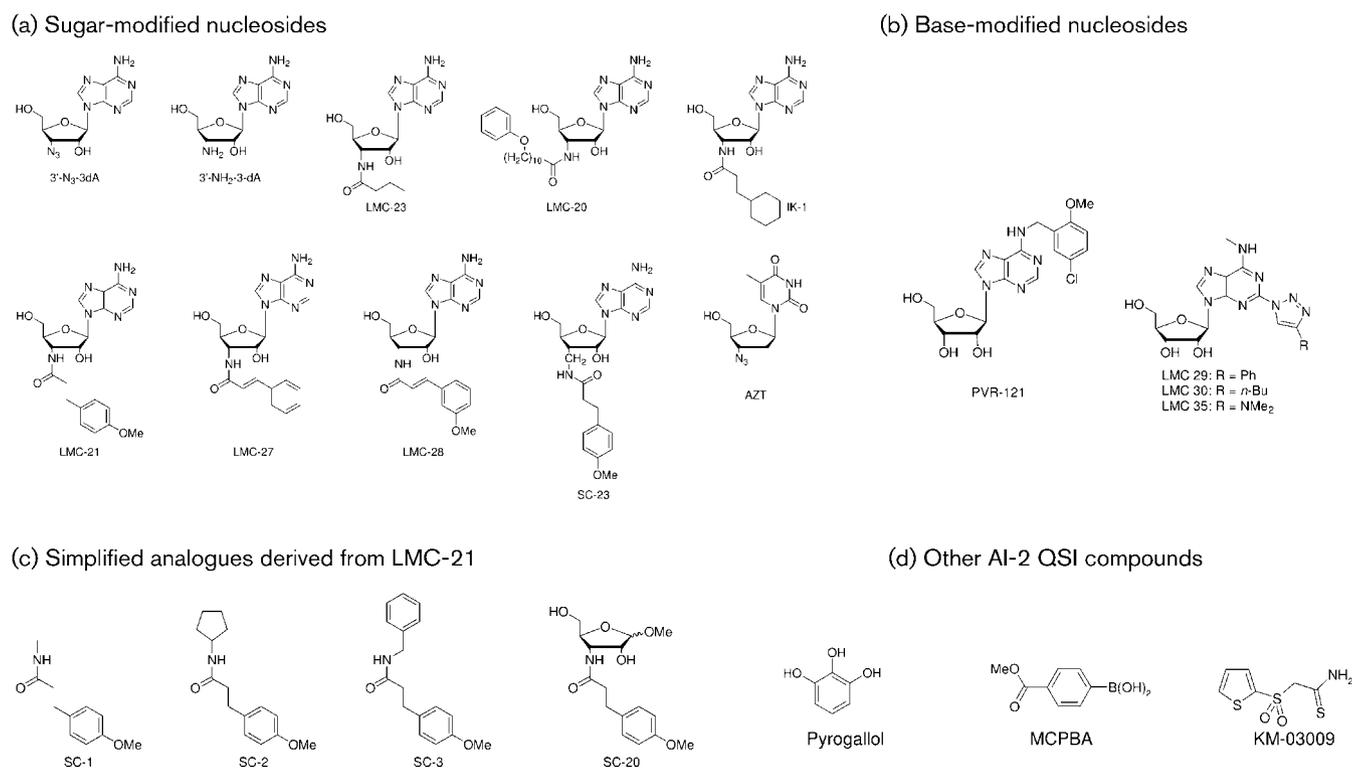
**Effect on QS-regulated virulence phenotypes *in vitro*.** The effect of AI-2 QS inhibitors on pigment production and protease activity in *V. anguillarum* LMG 4411 was determined as described previously (Croatto *et al.*, 2002). Each compound was tested at least twice in triplicate ( $n \geq 6$ ). Biofilms were grown according to Brackman *et al.* (2008). In brief, the *Vibrio* strains were grown overnight in MB and approximately  $10^8$  c.f.u. ml<sup>-1</sup> was added to the wells of a 96-well microtitre plate in the presence or absence of QS inhibiting (QSI) compounds. Bacteria were allowed to adhere and grow without agitation for 4 h at 30 °C. After 4 h, plates were emptied and rinsed with sterile physiological saline (PS). After this rinsing step, fresh MB (with or without compounds) was added and the plate was incubated for 20 h at 30 °C.

Biofilm biomass was quantified by crystal violet (CV) staining (Peeters *et al.*, 2008). The control signal corresponded to an A<sub>590</sub> of  $0.604 \pm 0.108$  and  $0.639 \pm 0.129$  for *V. anguillarum* LMG4411 and *V. vulnificus* LMG16867, respectively. For quantification of the number of metabolically active (i.e. living) cells in the biofilm, a resazurin assay was used (Peeters *et al.*, 2008). Each compound was tested at least six times in triplicate ( $n \geq 18$ ).

**Effect on QS-regulated stress responses *in vitro*.** *Vibrio* species were grown overnight in MB, and cells were collected by centrifugation and resuspended in artificial seawater (ASW) (Bang *et al.*, 2007). A 1 ml sample of the bacterial suspension was transferred to 100 ml glass bottles containing 19 ml ASW (with and without test compound). These suspensions were incubated at 30 °C without shaking. After 48 h, 1 ml samples were taken and the number of culturable cells was determined by plating serial dilutions on tryptic soy agar (TSA) (Oxoid) plates containing 2% (w/v) NaCl. Results were expressed as numbers of viable cells present after 48 h. Each assay was repeated at least three times. The effect of the compounds on the susceptibility towards doxycycline and chloramphenicol of all the *Vibrio* strains tested was determined as described previously (Brackman *et al.*, 2008). Each assay was repeated at least three times. A change in MIC was considered relevant if there was a shift of more than two doubling dilutions in either direction.

**Artemia challenge tests.** All experiments were performed with high-quality hatching cysts of *Artemia franciscana* shrimps (EG type, batch 6940, INVE Aquaculture). Cysts (200 mg) were hydrated in 18 ml tap water over 1 h. The procedure of Marques *et al.* (2004) was used to obtain sterile decapsulated cysts and nauplii. Challenge tests (in triplicate) were performed as described previously (Brackman *et al.*, 2008).

**Cytostatic activity assay.** The murine (L1210) and human (CEM, HeLa) cells were seeded at a concentration of  $5.0\text{--}7.5 \times 10^4$  cells per 200  $\mu$ l in wells of a 96-well microtitre plate in the presence of serial (fivefold) dilutions of the test compound, using RPMI 1640 culture medium supplemented with 2 mM L-glutamine, 0.075% (w/v) NaHCO<sub>3</sub>, and 10% (w/v) fetal bovine serum. After 48 h (L1210) or 72 h (CEM, HeLa), the cell numbers were determined using a Coulter Counter (Analisis).



**Fig. 1.** Overview of all analogues used in this study and not previously investigated in the context of QS or biofilm inhibitory activity (a, b and c), and compounds previously only investigated for their effect on AI-2 QS (d). (a) Sugar-modified nucleosides. (b) Base-modified nucleosides. (c) Simplified analogues derived from LMC-21. (d) Other AI-2 QSI compounds.

**Statistics.** The normal distribution of the data was checked using the Shapiro–Wilk test. Normally and non-normally distributed data were analysed using an independent samples *t* test and the Mann–Whitney *U* test, respectively. Statistics were performed using SPSS software, version 17.0.

## RESULTS

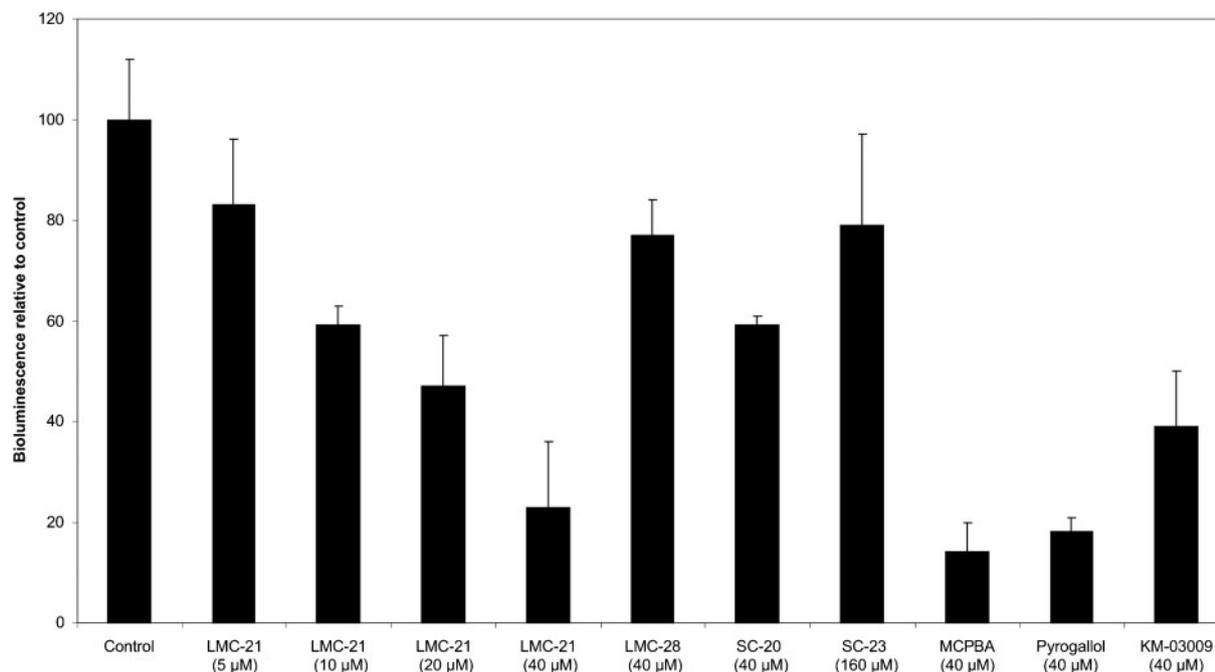
### Inhibition of AI-2 controlled bioluminescence

The antimicrobial activity of all compounds was evaluated against all strains used in the present study, and MICs were found to be higher than 320  $\mu$ M (160  $\mu$ M for pyrogallol). Unless otherwise mentioned, the compounds were used at a concentration of 40  $\mu$ M, which is well below the MIC for all strains tested. Bioluminescence in a constitutively bioluminescent strain *E. coli* DH5 $\alpha$ pBlueLux was not inhibited by any of the compounds tested (see Supplementary Table S1). The effect on AI-2 QS was assessed using *V. harveyi* BB170. LMC-21 was the most active adenosine derivative and a concentration-dependent inhibitory effect was observed (Fig. 2). Its isomer LMC-28, which only differed in the substitution site of the methoxy group, and SC-20, a truncated ribofuranosyl analogue, also inhibited AI-2 QS (Fig. 2), but proved significantly weaker

than LMC-21. SC-23 yielded a significant inhibition of QS in the *V. harveyi* BB170 reporter strain only when tested at a concentration above 40  $\mu$ M (Fig. 2). In addition, MCPBA, KM-03009 and pyrogallol were able to block the AI-2 QS system (Fig. 2). None of the other compounds resulted in a reduction in bioluminescence, even when used in higher concentrations (up to 160  $\mu$ M).

### Molecular target of the phenylpropionamidofuranosyl derivatives

To identify the molecular target of the 3'-deoxy-3'-(4-methoxyphenylpropionamido)ribofuranosyl derivatives, bioluminescence assays were conducted using several AI-2 QS mutants. No inhibitory effects were observed using the *V. harveyi* JAF375 and *V. harveyi* BB886 mutants, while inhibition was observed using the *V. harveyi* JMH597 mutant, suggesting an effect on AI-2 QS. The supernatants of *E. coli* K-12 treated with the compounds revealed no difference in AI-2 activity compared with the control. Furthermore, LMC-21 blocked bioluminescence in *V. harveyi* MM30, but not in *V. harveyi* JAF553, JAF483 or BNL258, suggesting that the target is located upstream of the mutations in the AI-2 signal transduction pathway and most likely is the LuxPQ complex in *V. harveyi*. Similar



**Fig. 2.** Bioluminescence in *V. harveyi* BB170 in the absence (control) and presence of QS inhibitors. Bioluminescence measurements were performed 6 h after the addition of the compounds. Bioluminescence of the control (without addition of compound) was set at 100% and the responses for other samples were normalized accordingly. Error bars, SD. Bioluminescence was significantly lower than that of the untreated control for all compounds ( $P < 0.05$ ).

results were obtained with SC-23, LMC-28, MCPBA and KM03009, suggesting that these molecules also target LuxPQ.

### Effect on protease activity and pigment production

LMC-21 significantly decreased pigment production by *V. anguillarum* LMG4411 after 48 h of growth, but none of the other compounds tested was able to significantly alter pigment production (Table 2). Addition of LMC-21,

MCPBA or pyrogallol resulted in a significantly decreased *V. anguillarum* LMG4411 protease activity (Table 2).

### Effect on *in vitro*-grown biofilms

The effect of the AI-2 QS inhibitors on the number of metabolically active cells in the biofilms of several *Vibrio* strains was evaluated using a resazurin assay. This assay revealed no significant decrease in the number of metabolically active cells in the biofilms of the different *Vibrio* strains following treatment (see Supplementary

**Table 2.** Effect of the AI-2 QS inhibitors (40 µM) on QS-regulated phenotypes

Compound	Protease activity,* <i>V. anguillarum</i> LMG4411	Pigment production,† <i>V. anguillarum</i> LMG4411	Biofilm formation‡	
			<i>V. anguillarum</i> LMG4411	<i>V. vulnificus</i> LMG16867
LMC-21	23 ± 3 %§	19 ± 10 %§	35 ± 11 %§	17 ± 15 %§
KM-03009	5 ± 12 %	2 ± 13 %	2 ± 22 %	5 ± 24 %
MCPBA	20 ± 2 %§	5 ± 16 %	36 ± 8 %§	18 ± 16 %§
Pyrogallol	18 ± 5 %§	10 ± 22 %	10 ± 10 %	40 ± 9 %§

\*Percentage reduction in protease activity compared with an untreated control ( $A_{590} 1.230 \pm 0.132$ ) (mean ± SD).

†Percentage reduction in pigment production compared with an untreated control ( $A_{405} 0.480 \pm 0.090$ ) (mean ± SD).

‡Percentage reduction in biofilm biomass (CV staining) compared with an untreated control (mean ± SD).

§Significantly different compared with an untreated control ( $P < 0.05$ ; independent samples *t* test).

Table S2). In contrast, several compounds decreased the CV staining of *V. anguillarum* LMG 4411 and *V. vulnificus* LMG16867 biofilms (Table 2). However, no significant anti-biofilm effects were observed for *V. harveyi* BB120 and *V. campbellii* LMG21363. In addition, the use of LMC-21 yielded a minor but significant increase in the CV signal for *V. cholerae* El Tor NCTC8457 ( $15 \pm 8\%$  compared with the untreated control).

### Effect on susceptibility of *Vibrio* species to stress

The effect of the different compounds on the starvation response and on the antimicrobial susceptibility of the different *Vibrio* species was investigated. Upon treatment with LMC-21, cell numbers significantly decreased in all *Vibrio* species (Table 3). Treatment with MCPBA, pyrogallol and KM-03009 reduced the number of culturable cells in some *Vibrio* species only (Table 3). There were no significant differences in the MICs for all *Vibrio* strains tested for chloramphenicol and doxycycline when used alone or in combination with a QS inhibitor (see Supplementary Tables S3 and S4).

### Effect on virulence *in vivo* and cytotoxicity

High mortality rates were observed when *Artemia* shrimps were exposed to *V. harveyi* BB120, but LMC-21 was able to completely protect *Artemia* during bacterial challenge (Fig. 3). LMC-21 alone had an effect neither on *Artemia* (Fig. 3) nor on *V. harveyi* BB120 (data not shown). In addition, LMC-21 was found to have  $IC_{50}$  values  $\geq 250 \mu\text{M}$  (L1210 cells) or  $\geq 125 \mu\text{M}$  (CEM and HeLa cells).

## DISCUSSION

QS is an important regulator of bacterial virulence in some bacterial species. Accordingly, QS inhibition is gaining interest as a potential alternative strategy for the treatment of bacterial infections. Although LuxS appears to be omnipresent in the bacterial world, the LuxPQ signal

transduction system is restricted to Vibrionales (Sun *et al.*, 2004; Rezzonico & Duffy, 2008). This makes the AI-2 receptor complex of Vibrionales an interesting target for the selective control of *Vibrio* species QS-regulated virulence.

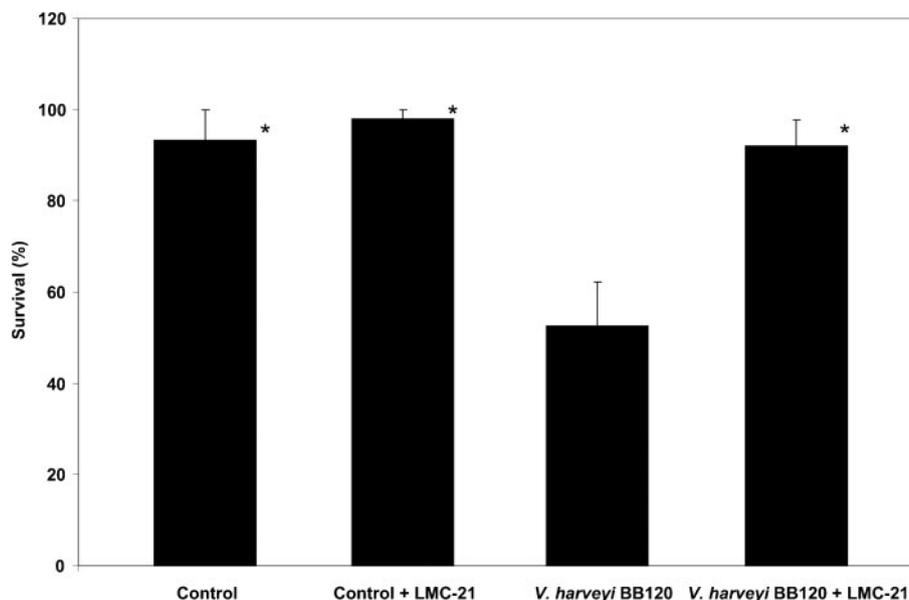
In this study, we not only confirmed the QS inhibitory activity of several established AI-2 QS inhibitors but also discovered several new inhibitors. To identify their molecular target, we evaluated the effect of the most active compound (LMC-21) on different *V. harveyi* QS mutants. Although we originally anticipated that certain adenosine analogues might disturb the biosynthesis of DPD, due to their structural similarity to *S*-adenosylmethionine, our data indicate that LMC-21 exerts its effect at the level of the AI-2 transduction system rather than that of AI-2 production. For these experiments, several *V. harveyi* QS mutants with mutations in the AI-2 signal transduction system were used. *V. harveyi* JAF553 and JAF483 contain a point mutation in the *luxU* and *luxO* genes, respectively, thereby preventing the phosphorelay capacity of LuxU and LuxO. *V. harveyi* BNL258 has a Tn5 insertion in the *hfq* gene, resulting in a non-functional Hfq protein. Since *V. harveyi* strains JAF553, JAF483 and BNL258 are all constitutively luminescent, a lack of inhibition of bioluminescence in one of these strains indicates that the inhibitor acts upstream of the mutated protein. Our compound proved incapable of blocking bioluminescence in these three QS mutants. This suggests that the target of the 3-(methoxyphenylpropionamido)ribofuranosyl derivatives is the upstream component of the AI-2 signalling transduction pathway, LuxPQ. In addition, no effect was observed when testing the compound in *V. harveyi* BB886, a mutant which lacks the LuxP receptor required for the AI-2 response, and in *V. harveyi* JAF375, a mutant which lacks LuxQ. Although several compounds inhibit the AI-2 QS system, there are few reports of QS inhibitors targeting LuxPQ. Phenylboronic acids, pyrogallol derivatives and 2-(2-thienylsulfonyl)ethanethioamide, previously reported to block the AI-2 QS system at the level of LuxPQ (Li *et al.*, 2008; Ni *et al.*, 2008a, b), were at best as active as LMC-21.

**Table 3.** Effect of the AI-2 QS inhibitors (40  $\mu\text{M}$ ) on the QS-regulated starvation response

Data are presented as numbers of viable cells ( $\times 10^8$ ) present after 48 h.

Compounds	Number of viable cells ( $\times 10^8$ ) (mean $\pm$ SD)				
	<i>V. anguillarum</i> LMG4411	<i>V. campbellii</i> LMG21363	<i>V. cholerae</i> NCTC8457	<i>V. harveyi</i> BB120	<i>V. vulnificus</i> LMG16867
Initial number of cells	1.05 $\pm$ 0.30	1.00 $\pm$ 0.23	1.16 $\pm$ 0.11	1.15 $\pm$ 0.14	1.11 $\pm$ 0.21
Control	0.77 $\pm$ 0.25	0.91 $\pm$ 0.18	1.10 $\pm$ 0.07	1.19 $\pm$ 0.45	1.09 $\pm$ 0.21
LMC-21	0.08 $\pm$ 0.07*	0.47 $\pm$ 0.09*	0.86 $\pm$ 0.12*	0.53 $\pm$ 0.43*	0.67 $\pm$ 0.01*
KM-03009	0.58 $\pm$ 0.07	0.93 $\pm$ 0.46	0.94 $\pm$ 0.45	0.85 $\pm$ 0.16*	1.11 $\pm$ 0.17
MCPBA	0.32 $\pm$ 0.17*	1.02 $\pm$ 0.32	1.24 $\pm$ 0.66	1.17 $\pm$ 0.37	0.93 $\pm$ 0.11
Pyrogallol	0.21 $\pm$ 0.04*	0.56 $\pm$ 0.15*	0.91 $\pm$ 0.13	1.22 $\pm$ 0.67	1.03 $\pm$ 0.05

\*Significantly different from the number of cells present after 48 h in the control ( $P < 0.05$ ; Mann-Whitney *U* test).



**Fig. 3.** Effect of LMC-21 on the survival of *Artemia*. Control: survival of *Artemia* without challenge with *V. harveyi* BB120 and without LMC-21. Control+LMC-21: survival of *Artemia* without challenge with *V. harveyi* BB120 in the presence of LMC-21 (40  $\mu$ M). *V. harveyi* BB120: survival of *Artemia* after challenge with *V. harveyi* BB120 in the absence of LMC-21. *V. harveyi* BB120+LMC-21: survival of *Artemia* after challenge with *V. harveyi* BB120 and treatment with LMC-21 (40  $\mu$ M). Asterisks indicate survival significantly different from treatment with pathogen alone ( $P < 0.001$ ).

None of these compounds has been previously evaluated for its effect on AI-2-related virulence. One molecule from each group of LuxPQ inhibitors was selected for further experiments. LMC-21 was able not only to reduce pigment production in *V. anguillarum* LMG4411 but also to decrease protease activity in this strain. In contrast, none of the established QS inhibitors targeting LuxPQ was able to block pigment production or to reduce protease more than did LMC-21. In addition, LMC-21 decreased the biofilm biomass of *V. anguillarum* and *V. vulnificus*, without reducing the number of viable cells present in the biofilms. Pyrogallol only decreased biofilm biomass in *V. vulnificus*, but to a higher extent than LMC-21. These data confirm the finding that pigment and protease production in *V. anguillarum* and biofilm formation in *V. anguillarum*, *V. vulnificus* and *V. cholerae* are (at least partially) controlled by the AI-2 QS system (Croxatto *et al.*, 2002; Zhu *et al.*, 2002; Hammer & Bassler, 2003; Lee *et al.*, 2007; Brackman *et al.*, 2008). Mutations in the LuxR homologues of *V. anguillarum* (VanT) and *V. vulnificus* (SmcR) have been shown to reduce biofilm formation in these species, indicating that AI-2 QS promotes biofilm formation in these species (Croxatto *et al.*, 2002; Lee *et al.*, 2007). In contrast, *V. cholerae* HapR represses the expression of *vps* genes (involved in the production of exopolysaccharides) and biofilm formation (Zhu *et al.*, 2002; Hammer & Bassler, 2003), indicating that AI-2 QS negatively influences biofilm formation in this species. However, the main QS signalling molecule in *V. cholerae* is CAI-1, and this may explain the limited effect of AI-2 QS

inhibitors on *V. cholerae* biofilm formation. Whether the increase in *V. cholerae* biomass, due to LMC-21, would impose problems in *in vivo* situations remains to be determined. In addition, *Vibrio* species are also known to regulate stress adaptation by means of their QS system. AI-2 is capable of regulating different stress responses, including starvation in *V. cholerae*, *V. vulnificus*, *V. anguillarum* and *Vibrio angustum* (McDougald *et al.*, 2001, 2003; Larsen *et al.*, 2004; Joelsson *et al.*, 2007; Lee *et al.*, 2007; Weber *et al.*, 2008). Our data indicate that LMC-21 suppresses the QS-regulated starvation response in all *Vibrio* species examined, while the other compounds increase susceptibility to starvation-associated stress conditions in some *Vibrio* species only, and to a lesser extent than LMC-21. However, our results indicate that AI-2 inhibition in five *Vibrio* species did not change their antimicrobial susceptibility. Of all the compounds tested, LMC-21 was the most interesting, since it was clearly at least as active in inhibiting *in vitro* virulence as the other active compounds tested in this study. Although a decrease in virulence *in vitro* is not always linked to a decrease in virulence *in vivo*, LMC-21 was shown to be a potent suppressor of *V. harveyi* BB120 virulence *in vivo*. LMC-21 alone had no effect on *Artemia* survival, and its lack of cytotoxicity, when used at 40  $\mu$ M, was confirmed using murine and human cell lines. It is interesting to note that halogenated furanones, well-documented QS inhibitors, have toxic side-effects in concentrations comparable to those used in the present study (Defoirdt *et al.*, 2006; Janssens, 2008).

In a preliminary search for the active pharmacophore of LMC-21, we synthesized two compounds based on the phenylpropionamidofuranosyl backbone. Based on their effect on AI-2-regulated bioluminescence in *V. harveyi* BB120, we identified the most important structural elements required for achieving QS inhibition. Minor changes, e.g. removing the methoxy group from the *para* (LMC-21) to the *meta* position (LMC-28), or the insertion of an extra CH<sub>2</sub> group between the phenylpropionamido substituent and the ribose moiety (SC-23), resulted in a decreased activity. Other molecules strongly resembling LMC-21, e.g. LMC-20 (longer side chain), LMC-23 (lacking the methoxy-substituted aromatic ring), LMC-27 (lacking the methoxy substitution on the aromatic ring) and IK-1, failed to inhibit the AI-2 QS system, and together point toward a specific (receptor-mediated) effect. We also investigated the importance of the adenine moiety present in LMC-21 by evaluating the effect of SC-1, SC-2, SC-3 and SC-20. Only SC-20 inhibited AI-2 QS, clearly showing that the ribofuranose moiety is required for activity. In addition, these results show that although an adenine group is not essential for activity, its presence results in more active compounds. However, the molecular interaction of these compounds with LuxPQ remains to be determined.

## ACKNOWLEDGEMENTS

This work was supported by the Institute for the Promotion of Innovation through Science and Technology in Flanders (IWT-Vlaanderen) and by the special research fund (BOF) of Ghent University.

## REFERENCES

- Alfaro, J. F., Zhang, T., Wynn, D. P., Karschner, E. L. & Zhou, Z. S. (2004). Synthesis of LuxS inhibitors targeting bacterial cell-cell communication. *Org Lett* **6**, 3043–3046.
- Austin, B. & Zhang, X. H. (2006). *Vibrio harveyi*: a significant pathogen of marine vertebrates and invertebrates. *Lett Appl Microbiol* **43**, 119–124.
- Azhayev, A. V. & Smrt, J. (1978). Nucleic-acids components and their derivatives. 193. Synthesis of 3'-azido-3'-deoxyadenosine and 3'-amino-3'-deoxyadenosine. *Collect Czech Chem Commun* **43**, 1520–1530.
- Bang, W., Drake, M. A. & Jaykus, L. A. (2007). Recovery and detection of *Vibrio vulnificus* during cold storage. *Food Microbiol* **24**, 664–670.
- Bassler, B. L., Wright, M., Showalter, R. E. & Silverman, M. R. (1993). Intercellular signalling in *Vibrio harveyi*: sequence and function of genes regulating expression of luminescence. *Mol Microbiol* **9**, 773–786.
- Bassler, B. L., Wright, M. & Silverman, M. R. (1994). Multiple signalling systems controlling expression of luminescence in *Vibrio harveyi*: sequence and function of genes encoding a second sensory pathway. *Mol Microbiol* **13**, 273–286.
- Bassler, B. L., Greenberg, E. P. & Stevens, A. M. (1997). Cross-species induction of luminescence in the quorum-sensing bacterium *Vibrio harveyi*. *J Bacteriol* **179**, 4043–4045.
- Boyd, E. F., Cohen, A. L., Naughton, L. M., Ussery, D. W., Binnewies, T. T., Stine, O. C. & Parent, M. A. (2008). Molecular analysis of the emergence of pandemic *Vibrio parahaemolyticus*. *BMC Microbiol* **8**, 110.
- Brackman, G., Defoirdt, T., Miyamoto, C., Bossier, P., Van Calenberg, S., Nelis, H. & Coenye, T. (2008). Cinnamaldehyde and cinnamaldehyde derivatives reduce virulence in *Vibrio* spp. by decreasing the DNA-binding activity of the quorum sensing response regulator LuxR. *BMC Microbiol* **8**, 149.
- Brackman, G., Hillaert, U., Van Calenberg, S., Nelis, H. J. & Coenye, T. (2009). Use of quorum sensing inhibitors to interfere with biofilm formation and development in *Burkholderia multivorans* and *Burkholderia cenocepacia*. *Res Microbiol* **160**, 144–151.
- Bross, M. H., Soch, K., Morales, R. & Mitchell, R. B. (2007). *Vibrio vulnificus*: diagnosis and treatment. *Am Fam Physician* **76**, 539–544.
- Chen, X., Schauder, S., Potier, N., Van Dorsselaer, A., Pelczar, I., Bassler, B. L. & Hughson, F. M. (2002). Structural identification of a bacterial quorum-sensing signal containing boron. *Nature* **415**, 545–549.
- Chiang, S. R. & Chuang, Y. C. (2003). *Vibrio vulnificus* infection: clinical manifestations, pathogenesis and antimicrobial therapy. *J Microbiol Immunol Infect* **36**, 81–88.
- Cosyn, L., Palaniappan, K. K., Kim, S. K., Duong, H. T., Gao, Z. G., Jacobson, K. A. & Van Calenberg, S. (2006). 2-Triazole-substituted adenosines: a new class of selective A3 adenosine receptor agonists, partial agonists, and antagonists. *J Med Chem* **49**, 7373–7383.
- Croxatto, A., Chalker, V. J., Lauritz, J., Jass, J., Hardman, A., Williams, P., Camara, M. & Milton, D. L. (2002). VanT, a homologue of *Vibrio harveyi* LuxR, regulates serine, metalloprotease, pigment, and biofilm production in *Vibrio anguillarum*. *J Bacteriol* **184**, 1617–1629.
- Defoirdt, T., Crab, R., Wood, T. K., Sorgeloos, P., Verstraete, W. & Bossier, P. (2006). Quorum sensing-disrupting brominate furanones protect the gnotobiotic brine shrimp *Artemia franciscana* from pathogenic *Vibrio harveyi*, *Vibrio campbellii* and *Vibrio parahaemolyticus* isolates. *Appl Environ Microbiol* **72**, 6419–6423.
- Defoirdt, T., Miyamoto, C. M., Wood, T. K., Meighen, E. A., Sorgeloos, P., Verstraete, W. & Bossier, P. (2007). The natural furanone (5Z)-4-bromo-5-(bromomethylene)-3-butyl-2(5H)-furanone disrupts quorum sensing-regulated gene expression in *Vibrio harveyi* by decreasing the DNA-binding activity of the transcriptional regulator protein LuxR. *Environ Microbiol* **9**, 2486–2495.
- Freeman, J. A. & Bassler, B. L. (1999a). Sequence and function of LuxU: a two component phosphorelay protein that regulates quorum sensing in *Vibrio harveyi*. *J Bacteriol* **181**, 899–906.
- Freeman, J. A. & Bassler, B. L. (1999b). A genetic analysis of the function of LuxO, a two component response regulator involved in quorum sensing in *Vibrio harveyi*. *Mol Microbiol* **31**, 665–667.
- Garcia, T., Otto, K., Kjelleberg, S. & Nelson, D. R. (1997). Growth of *Vibrio anguillarum* in salmon intestinal mucus. *Appl Environ Microbiol* **63**, 1034–1039.
- Griffith, D. C., Kelly-Hope, L. A. & Miller, M. A. (2006). Review of reported cholera outbreaks worldwide, 1995–2005. *Am J Trop Med Hyg* **75**, 973–977.
- Hammer, B. K. & Bassler, B. L. (2003). Quorum sensing controls biofilm formation in *Vibrio cholerae*. *Mol Microbiol* **50**, 101–104.
- Harvell, C. D., Mitchell, C. E., Ward, J. R., Altizer, S., Dobson, A. P., Ostfeld, R. S. & Samuel, M. D. (2002). Climate warming and disease risks for terrestrial and marine biota. *Science* **296**, 2158–2162.
- Higgins, D. A., Pomianek, M. E., Kraml, C. M., Taylor, R. K., Semmelhack, M. F. & Bassler, B. L. (2007). The major *Vibrio cholerae* autoinducer and its role in virulence factor production. *Nature* **450**, 883–886.

- Igbinosa, E. O. & Okoh, A. I. (2008). Emerging *Vibrio* species: an unending threat to public health in developing countries. *Res Microbiol* **159**, 495–506.
- Janssens, J. C. (2008). *Chemical synthesis of N-acyl homoserine lactone analogues and brominated furanones, and their biological activities with Salmonella enterica serovar Typhimurium and Vibrio spp.* Doctoral thesis, KU Leuven, Leuven, Belgium.
- Joelsson, A., Kan, B. & Zhu, J. (2007). Quorum sensing enhances the stress response in *Vibrio cholerae*. *Appl Environ Microbiol* **73**, 3742–3746.
- Kapp, C. (2009). Zimbabwe's humanitarian crisis worsens. *Lancet* **373**, 447.
- Karunasagar, I., Pai, R., Malahti, G. R. & Karunasagar, I. (1994). Mass mortality of *Panaeus monodon* larvae due to antibiotic-resistant *Vibrio harveyi* infection. *Aquaculture* **128**, 203–209.
- Kim, S. K., Gao, Z. G., Van Rompaey, P., Gross, A. S., Chen, A., Van Calenbergh, S. & Jacobson, K. A. (2003). Modeling the adenosine receptors: comparison of the binding domains of A<sub>2A</sub> agonist and antagonists. *J Med Chem* **46**, 4847–4859.
- Larsen, M. H., Blackburn, N., Larsen, J. L. & Olsen, J. E. (2004). Influences of temperature, salinity and starvation on the motility and chemotactic response of *Vibrio anguillarum*. *Microbiology* **150**, 1283–1290.
- Lee, J. H., Rhee, J. E., Park, U., Ju, H. M., Lee, B. C., Kim, T. S., Jeong, H. S. & Choi, S. H. (2007). Identification and functional analysis of *Vibrio vulnificus* SmcR, a novel global regulator. *J Microbiol Biotechnol* **17**, 325–334.
- Lenz, D. H., Mok, K. C., Lilley, B. N., Kulkarni, R. V., Wingreen, N. S. & Bassler, B. L. (2004). The small RNA chaperone Hfq and multiple small RNAs control quorum sensing in *Vibrio harveyi* and *Vibrio cholerae*. *Cell* **118**, 69–82.
- Li, M., Ni, N., Chou, H. T., Lu, C. D., Tai, P. C. & Wang, B. (2008). Structure-based discovery and experimental verification of novel AI-2 quorum sensing inhibitors against *Vibrio harveyi*. *ChemMedChem* **3**, 1242–1249.
- Lynch, S. V. & Wiener-Kronisch, J. P. (2008). Novel strategies to combat bacterial virulence. *Curr Opin Crit Care* **14**, 593–599.
- Marques, A., Francois, J. M., Dhont, J., Bossier, P. & Sorgeloos, P. (2004). Influence of yeast quality on performance of gnotobiotically grown *Artemia*. *J Exp Mar Biol Ecol* **310**, 247–264.
- McDougald, D., Rice, S. A. & Kjelleberg, S. (2001). SmcR-dependent regulation of adaptive phenotypes in *Vibrio vulnificus*. *J Bacteriol* **183**, 758–762.
- McDougald, D., Srinivasan, S., Rice, S. A. & Kjelleberg, S. (2003). Signal-mediated cross-talk regulates stress adaptation in *Vibrio* species. *Microbiology* **149**, 1923–1933.
- Miller, S. T., Xavier, K. B., Campagna, S. R., Taga, M. E., Semmelhack, M. F., Bassler, B. L. & Hughson, F. M. (2004). *Salmonella typhimurium* recognizes a chemically distinct form of the bacterial quorum-sensing signal AI-2. *Mol Cell* **15**, 677–687.
- Moradei, O., De Mortier, C., Varela, O. & De Lederkremer, R. M. (1991). Synthesis of furanose glycoside of abequeose (3,6-dideoxy-D-xylo-hexose). *J Carbohydr Chem* **10**, 469–479.
- Ni, N., Choudhary, G., Li, M. & Wang, B. (2008a). Pyrogallol and its analogs can antagonize bacterial quorum sensing in *Vibrio harveyi*. *Bioorg Med Chem Lett* **18**, 1567–1572.
- Ni, N., Chou, H. T., Wang, J., Li, M., Lu, C. D., Tai, P. C. & Wang, B. (2008b). Identification of boronic acids as antagonists of bacterial quorum sensing in *Vibrio harveyi*. *Biochem Biophys Res Commun* **369**, 590–594.
- Ni, N., Li, M., Wang, J. & Wang, B. (2009). Inhibitors of bacterial quorum sensing. *Med Res Rev* **29**, 65–124.
- Ohno, M., Gao, Z. G., Van Rompaey, P., Tchilibon, S., Kim, S. K., Harris, B. A., Gross, A. S., Duong, H. T., Van Calenbergh, S. & Jacobson, K. A. (2004). Modulation of adenosine receptor affinity and intrinsic efficacy in adenine nucleosides substituted at the 2-position. *Bioorg Med Chem* **12**, 2995–3007.
- Peeters, E., Nelis, H. J. & Coenye, T. (2008). Comparison of multiple methods for quantification of microbial biofilms grown in microtiter plates. *J Microbiol Methods* **72**, 157–165.
- Ren, D., Bedzyk, L. A., Ye, R. W., Thomas, S. M. & Wood, T. K. (2004). Differential gene expression shows natural brominated furanones interfere with the autoinducer-2 bacterial signalling system of *Escherichia coli*. *Biotechnol Bioeng* **88**, 630–642.
- Rezzonico, F. & Duffy, B. (2008). Lack of genomic evidence of AI-2 receptors suggests a non-quorum sensing role for *luxS* in most bacteria. *BMC Microbiol* **8**, 154.
- Ryan, R. P. & Dow, J. M. (2008). Diffusible signals and interspecies communication in bacteria. *Microbiology* **154**, 1845–1858.
- Sack, D. A., Sack, R. B., Nair, G. B. & Siddique, A. K. (2004). Cholera. *Lancet* **363**, 223–233.
- Scrascia, M., Maimone, F., Mohamud, K. A., Materu, S. F., Grimont, F., Grimont, P. A. & Pazzani, C. (2006). Clonal relationship among *Vibrio cholerae* O1 El Tor strains causing the largest cholera epidemic in Kenya in the late 1990s. *J Clin Microbiol* **44**, 3401–3404.
- Shen, G., Rajan, R., Zhu, J., Bell, C. E. & Pei, D. (2006). Design and synthesis of substrate and intermediate analogue inhibitors of S-ribosylhomocysteinase. *J Med Chem* **49**, 3003–3011.
- Soenens, J., Francois, G., Vandeneeckhout, E. & Herdewijn, P. (1995). Synthesis of 3'-amino-3'-deoxyadenosine derivatives as potential drugs for the treatment of malaria. *Nucleosides Nucleotides* **14**, 409–411.
- Sun, J., Daniel, R., Wagner-Döbler, I. & Zeng, A. P. (2004). Is autoinducer-2 a universal signal for interspecies communication: a comparative genomic and phylogenetic analysis of the synthesis and signal transduction pathways. *BMC Evol Biol* **4**, 36.
- Surette, M. G., Miller, M. B. & Bassler, B. L. (1999). Quorum sensing in *Escherichia coli*, *Salmonella typhimurium*, and *Vibrio harveyi*: a new family of genes responsible for autoinducer production. *Proc Natl Acad Sci U S A* **96**, 1639–1644.
- Weber, B., Croxatto, A., Chen, C. & Milton, D. L. (2008). RpoS induces expression of the *Vibrio anguillarum* quorum-sensing regulator VanT. *Microbiology* **154**, 767–780.
- Winzer, K., Hardie, K. R., Burgess, N., Doherty, N., Kirke, D., Holden, M. T., Linforth, R., Cornell, K. A., Taylor, A. J. & other authors (2002). LuxS: its role in central metabolism and the *in vitro* synthesis of 4-hydroxy-5-methyl-3(2H)-furanone. *Microbiology* **148**, 909–922.
- Xavier, K. B. & Bassler, B. L. (2003). LuxS quorum sensing: more than just a numbers game. *Curr Opin Microbiol* **6**, 191–197.
- Zhu, J., Miller, M. B., Vance, R. E., Dziejman, M., Bassler, B. L. & Mekalanos, J. J. (2002). Quorum-sensing regulators control virulence gene expression in *Vibrio cholerae*. *Proc Natl Acad Sci U S A* **99**, 3129–3134.

---

Edited by: J. Anné