



“*Candidatus Colwellia aromaticivorans*” sp. nov., “*Candidatus Halocyntiibacter alkanivorans*” sp. nov., and “*Candidatus Ulvibacter alkanivorans*” sp. nov. Genome Sequences

Mariana E. Campeão,^{a,b} Jean Swings,^{a,b,c} Bruno Sergio Silva,^{a,b} Koko Otsuki,^{a,b} Fabiano L. Thompson,^{a,b} Cristiane C. Thompson^a

^aLaboratory of Microbiology, Instituto de Biologia, Universidade Federal do Rio de Janeiro (UFRJ), Rio de Janeiro, Brazil

^bCenter of Technology-CT2, SAGE-COPPE, Federal Universidade Federal do Rio de Janeiro (UFRJ), Rio de Janeiro, Brazil

^cGhent University, Ghent, Belgium

ABSTRACT Unplanned oil spills during offshore production are a serious problem for the industry and the marine environment. Here, we present the genome sequence analysis of three novel hydrocarbon-degrading bacteria, namely, “*Candidatus Colwellia aromaticivorans*” sp. nov., “*Candidatus Halocyntiibacter alkanivorans*” sp. nov., and “*Candidatus Ulvibacter alkanivorans*” sp. nov.

Metagenomic data obtained from microcosm experiments with oil incubation in seawater from the Foz do Amazonas and Barreirinhas basins indicated the biodegradation potential of deep-sea microbial communities (1). Here, we present the metagenome assembled genome (MAG) sequences of three novel hydrocarbon-degrading bacteria obtained from these previous microcosm experiments, as detailed in our previous work (1). First, read trimming was obtained with prinseq v0.20.4 (-trim_qual_right/trim_qual_left 20, -trim_qual_type min, -trim_qual_rule lt, -trim_qual_window 5, -trim_qual_step 1, -min_len 35, -min_qual_mean 20, -trim_left/-trim_right 10, -out_bad null, -derep 1, and -custom_params\“CTGTCTCTTATACACATCT 1;CTGTCTCTTATACACATCTGACGCTGCCGACGA 1;CTGTCTCTTATACACATCTCCGAGCCCACGAGAC 1\” (2). Subsequently, a binning approach was performed through cross-assembly using eight metagenomes from Foz do Amazonas and eight metagenomes from Barreirinhas with SPAdes v3.6.2 (3); using k-mers of 21, 33, 55, 77, 99, and 127; mapping reads against assembled scaffolds with Bowtie 2 v2.2.5 (4) (-very-fast, -a, -no-unal, -no-discordant, and -no-mixed); converting to bam format with SAMtools v1.7 (5); coverage estimating with jgi_summarize_bam_contig_depths (-outputDepth); and binning with MetaBAT v0.25.4 (-specific, -v, -m 2500, and -minSamples 5) (6). RefineM v0.0.23 (scaffold_stats, outliers, and filter_bins commands) (7) was used to remove contamination, and CheckM v1.0.11 (lineage_wf workflow) (8) was used to assess quality stats. Genomic taxonomy was performed as described previously (9–12). These three novel MAGs showed less than 98.7% 16S rRNA sequence identity with the closest species. The three novel lineages had <95% amino acid identity (AAI) and <70% DNA-DNA hybridization (DDH) toward the closest phylogenetic neighbors, suggesting that the recovered genomes are new species, namely, “*Candidatus Colwellia aromaticivorans*” sp. nov., “*Candidatus Halocyntiibacter alkanivorans*” sp. nov., and “*Candidatus Ulvibacter alkanivorans*” sp. nov. (Table 1). “*Candidatus Ulvibacter alkanivorans*” sp. nov. and “*Candidatus Halocyntiibacter alkanivorans*” sp. nov. possess an alkane monooxygenase, the specific enzyme for alkane degradation, that was lacking in “*Candidatus Colwellia aromaticivorans*” sp. nov. “*Candidatus Colwellia aromaticivorans*” has a phenol 2-monooxygenase gene involved in toluene biodegradation and harbors two mono-

Citation Campeão ME, Swings J, Silva BS, Otsuki K, Thompson FL, Thompson CC. 2019. “*Candidatus Colwellia aromaticivorans*” sp. nov., “*Candidatus Halocyntiibacter alkanivorans*” sp. nov., and “*Candidatus Ulvibacter alkanivorans*” sp. nov. genome sequences. Microbiol Resour Announc 8:e00086-19. <https://doi.org/10.1128/MRA.00086-19>.

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2019 Campeão et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Fabiano L. Thompson, fabianothompson1@gmail.com, or Cristiane C. Thompson, thompsoncristiane@gmail.com.

Received 24 January 2019

Accepted 3 March 2019

Published 11 April 2019

TABLE 1 Genomic features of metagenome-associated genomes^a

Genome feature	Data for strain:		
	" <i>Candidatus Colwellia aromaticivorans</i> " sp. nov.	" <i>Candidatus Halocynthiibacter alkanivorans</i> " sp. nov.	" <i>Candidatus Ulvibacter alkanivorans</i> " sp. nov.
Coverage (×)	97	19	19
Completeness (%)	97.4	98.7	93.1
Contamination (%)	1.8	2.15	0.2
Size (Mbp)	3.71	4.7	2.7
No. of contigs	101	171	117
<i>N</i> ₅₀ value (bp)	72,862	55,069	47,750
No. of ORFs ^b	3,278	4,540	2,625
16S rRNA similarity with closest relative (%)	<i>Colwellia rossensis</i> S51-W(gv)1 ^T (98.5)	<i>Halocynthiibacter arcticus</i> PAMC 20958 ^T (97.2)	<i>Ulvibacter antarcticus</i> IMCC3101 ^T (96.2)
GGD ^c with closest relative (%)	<i>Colwellia rossensis</i> S51-W(gv)1 ^T (ND) ^d	<i>Halocynthiibacter arcticus</i> PAMC 20958 ^T (19.4)	<i>Ulvibacter antarcticus</i> DSM 23424 (17.8)
AAI with closest relative (%)	<i>Colwellia rossensis</i> S51-W(gv)1 ^T (ND)	<i>Halocynthiibacter arcticus</i> PAMC 20958 ^T (65.4)	<i>Ulvibacter antarcticus</i> DSM 23424 (71.8)

^a 16S rRNA sequences recovered were retrieved with RNAmmer (14) and compared with BLAST (15) against the Silva database (16, 17). The closest were selected to align through ssu-align v0.1.1 (18, 19) and perform phylogeny through IQ-TREE (20–22). The 16S rRNA similarity was calculated with TaxonDC (23). AAI and DDH values were performed with CompareM (<https://github.com/dparks1134/CompareM>) and GGDC (<http://ggdc.dsmz.de/>), respectively. The genome of *Ulvibacter antarcticus* DSM 23424 was used for AAI and DDH analysis. *In silico*-predicted phenotype analysis reveals that "*Candidatus Colwellia aromaticivorans*" sp. nov. is positive for chitin hydrolysis, propionate utilization, and D-lactate utilization. "*Candidatus Halocynthiibacter alkanivorans*" sp. nov. is positive for acid production from D-ribose, sucrose, and glycerol; acetate utilization; indole production; and nitrate reduction. "*Candidatus Ulvibacter alkanivorans*" sp. nov. is positive for D-lactic acid utilization.

^b ORFs, open reading frames.

^c GGD, genome-to-genome distance.

^d ND, not determined.

oxygenases and nine oxidoreductases. Oxygenases, a class of oxidoreductases, are key enzymes in hydrocarbon degradation (13).

"*Candidatus Colwellia aromaticivorans*" (ar.o.ma'ti.ca. N.L. adj. *aromaticivorans*, aromatic, referring to the property of utilizing aromatic compounds). *In silico*-predicted phenotypes indicate that the novel species hydrolyzes chitin, utilizes propionate and D-lactate, and does not utilize D-fructose, L-arabinose, glycerol, citrate, or L-malate. The G+C content is 38%.

"*Candidatus Halocynthiibacter alkanivorans*" (al.ka.ni'vo.rans. M.L. n. *alkanum* saturated aliphatic hydrocarbon; L. v. *vorare* to eat; L. adj. *alkanivorans* alkane-devouring). *In silico*-predicted phenotypes indicate that the novel species produces acid from D-ribose, sucrose, and glycerol. Acetate is utilized, nitrate is reduced, and indole is produced. The G+C content is 59%.

"*Candidatus Ulvibacter alkanivorans*" (al.ka.ni'vo.rans. M.L. n. *alkanum* saturated aliphatic hydrocarbon; L. v. *vorare* to eat; L. adj. *alkanivorans* alkane-devouring). *In silico*-predicted phenotypes indicate that the novel species utilizes D-lactate and does not hydrolyze starch or carboxymethyl cellulose (CM-cellulose). It is negative for the utilization of D-mannitol, cellobiose, D-fructose, L-fucose, D-galactose, myo-inositol, D-sorbitol, sucrose, trehalose, and propionic acid. The G+C content is 40%.

Data availability. This whole-genome shotgun project has been deposited in GenBank under the BioProject accession no. [PRJNA478776](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA478776).

ACKNOWLEDGMENTS

We thank CNPQ, CAPES, and FAPERJ for the support.

REFERENCES

1. Campeão ME, Reis L, Leomil L, de Oliveira L, Otsuki K, Gardinali P, Pelz O, Valle R, Thompson FL, Thompson CC. 2017. The deep-sea microbial community from the Amazonian basin associated with oil degradation. *Front Microbiol* 8:1019. <https://doi.org/10.3389/fmicb.2017.01019>.
2. Schmieder R, Edwards R. 2011. Quality control and preprocessing of metagenomic datasets. *Bioinformatics* 27:863–864. <https://doi.org/10.1093/bioinformatics/btr026>.
3. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pyshkin AV. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
4. Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods* 9:357–359. <https://doi.org/10.1038/nmeth.1923>.
5. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis

- G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25: 2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>.
6. Kang DD, Froula J, Egan R, Wang Z. 2015. MetaBAT, an efficient tool for accurately reconstructing single genomes from complex microbial communities. *PeerJ* 3:e1165. <https://doi.org/10.7717/peerj.1165>.
 7. Parks DH, Rinke C, Chuvochina M, Chaumeil P-A, Woodcroft BJ, Evans PN, Hugenholtz P, Tyson GW. 2017. Recovery of nearly 8,000 metagenome-assembled genomes substantially expands the tree of life. *Nat Microbiol* 2:1533–1542. <https://doi.org/10.1038/s41564-017-0012-7>.
 8. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
 9. Amaral GRS, Dias GM, Wellington-Oguri M, Chimetto L, Campeão ME, Thompson FL, Thompson CC. 2014. Genotype to phenotype: identification of diagnostic vibrio phenotypes using whole genome sequences. *Int J Syst Evol Microbiol* 64:357–365. <https://doi.org/10.1099/ijs.0.057927-0>.
 10. Amaral GRS, Campeão ME, Swings J, Thompson FL, Thompson CC. 2015. Finding diagnostic phenotypic features of *Photobacterium* in the genome sequences. *Antonie Van Leeuwenhoek* 107:1351–1358. <https://doi.org/10.1007/s10482-015-0414-6>.
 11. Thompson CC, Amaral GR, Campeão M, Edwards RA, Polz MF, Dutilh BE, Ussery DW, Sawabe T, Swings J, Thompson FL. 2015. Microbial taxonomy in the post-genomic era: rebuilding from scratch? *Arch Microbiol* 197: 359–370. <https://doi.org/10.1007/s00203-014-1071-2>.
 12. De Vos P, Thompson F, Thompson C, Swings J. 2017. A flavor of prokaryotic taxonomy: systematics revisited, p 29–44. In Kurtböke I (ed), *Microbial resources: from functional existence in nature to applications*. Elsevier B.V., Amsterdam, Netherlands.
 13. Vandecasteele JP, Jones T. 2008. *Petroleum microbiology: concepts, environmental implications, industrial applications*. Editions Technips, Paris, France.
 14. Lagesen K, Hallin P, Rødland EA, Stærfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <https://doi.org/10.1093/nar/gkm160>.
 15. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
 16. Pruesse E, Quast C, Knittel K, Fuchs BM, Ludwig W, Peplies J, Glöckner FO. 2007. SILVA: a comprehensive online resource for quality checked and aligned ribosomal RNA sequence data compatible with ARB. *Nucleic Acids Res* 35:7188–7196. <https://doi.org/10.1093/nar/gkm864>.
 17. Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, Peplies J, Glöckner FO. 2013. The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic Acids Res* 41: D590–D596. <https://doi.org/10.1093/nar/gks1219>.
 18. Nawrocki EP, Kolbe DL, Eddy SR. 2009. Infernal 1.0: inference of RNA alignments. *Bioinformatics* 25:1335–1337. <https://doi.org/10.1093/bioinformatics/btp157>.
 19. Cannone JJ, Subramanian S, Schnare MN, Collett JR, D'Souza LM, Du Y, Feng B, Lin N, Madabusi LV, Müller KM, Pande N, Shang Z, Yu N, Gutell RR. 2002. The Comparative RNA Web (CRW) site: an online database of comparative sequence and structure information for ribosomal, intron, and other RNAs. *BMC Bioinformatics* 3:2. <https://doi.org/10.1186/1471-2105-3-2>.
 20. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. 2015. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Mol Biol Evol* 32:268–274. <https://doi.org/10.1093/molbev/msu300>.
 21. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: improving the ultrafast bootstrap approximation. *Mol Biol Evol* 35:518–522. <https://doi.org/10.1093/molbev/msx281>.
 22. Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermini LS. 2017. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat Methods* 14:587–589. <https://doi.org/10.1038/nmeth.4285>.
 23. Tarlachkov SV, Starodumova IP. 2017. TaxonDC: calculating the similarity value of the 16S rRNA gene sequences of prokaryotes or ITS regions of fungi. *J Bioinform Genom* 3:1–4. <https://doi.org/10.18454/jbg.2017.3.5.1>.