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How habitat-modifying organisms structure the food web of two coastal ecosystems

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Abstract

The diversity and structure of ecosystems has been found to depend both on trophic interactions in food webs and on other species interactions such as habitat modification and mutualism that form non-trophic interaction networks. However, quantification of the dependencies between these two main interaction networks has remained elusive. In this study, we assessed how habitat-modifying organisms affect basic food web properties by conducting in-depth empirical investigations of two ecosystems: North American temperate fringing marshes and West African tropical seagrass meadows. Results reveal that habitatmodifying species, through non-trophic facilitation rather than their trophic role, enhance species richness across multiple trophic levels, increase the number of interactions per species (link density), but decrease the realized fraction of all possible links within the food web (connectance). Compared to the trophic role of the most highly connected species, we found this non-trophic effects to be more important for species richness and of more or similar importance for link density and connectance. Our findings demonstrate that food webs can be fundamentally shaped by interactions outside the trophic network, yet intrinsic to the species participating in it. Better integration of non-trophic interactions in food web analyses may therefore strongly contribute to their explanatory and predictive capacity.

Keywords: consumer-resource interactions, non-trophic interactions, facilitation, mutualism, ecological networks, ecosystem engineering, foundation species

Introduction

One of the great challenges in ecology is to elucidate how different types of species interactions drive the structure and dynamics of communities and ecosystems. Ever since Darwin (1859) coined the term 'web of life' [1], food webs have been intensively studied as paradigmatic examples of natural complex systems [2-4]. To date, analyses investigating the stability and structure of species interaction networks have primarily focused on the properties of the network formed by feeding interactions between species [5-12]. Studies typically investigate the topology of trophic interactions (e.g. links per species, connectance) [5, 9], variation in interaction strength (e.g. across trophic levels), and the nature of trophic interactions (e.g. predator-prey, plant-herbivore) [8, 11, 13, 14].

However, species do not only interact through feeding interactions. Non-trophic interactions, such as mutualism and habitat modification, are pervasive in ecosystems and, through their impact on species abundance and the strength of individual trophic links, may transform the topology and dynamics of the overall network [15, 16]. Despite urgent calls from recent studies to integrate non-trophic interactions [4, 17-20], food webs are still typically studied without considering species interactions outside the trophic network, and quantification of the impacts of non-trophic effects on food web structure has thus far remained elusive. Therefore, even after 150 years the question remains: are food webs mostly 'self-shaped' by trophic interactions alone or are they fundamentally contingent on non-trophic interactions?

Here, we empirically test the hypothesis that, in ecosystems dominated by organisms that strongly modify their abiotic environment (hereafter called 'habitat modifiers'), overall food web complexity is enhanced by these modifications, beyond previously documented single-species facilitation effects. Habitat modifiers, also described as 'ecosystem engineers' or 'foundation species', are increasingly recognized as important drivers of ecosystem

functions [16, 21-28]. Although habitat modifiers are part of the food web like any other species (e.g. as prey or predator), they also have non-trophic effects on associated species by creating new habitat, altering resource availability and modifying physical environmental conditions. In theory, these non-trophic effects can be positive for some species (facilitation) [16], and negative for others, meaning the overall impact of non-trophic interactions on food web structure may be positive, negative, or neutral [19]. Despite their ubiquity and pronounced, well-documented direct effects on specific species and individual trophic interactions, it remains unclear (1) how habitat modifiers affect the overall food web, (2) how important non-trophic interactions by habitat modifiers are compared to their own trophic interactions, and (3) how these non-trophic effects compare in importance to those species with the highest number of trophic links in the food web (hereafter called 'most highly connected species') [20].

To investigate whether key food web properties are indeed contingent on non-trophic facilitation by habitat modifiers as hypothesized, we carried out detailed field-based studies in two ecosystems: (1) temperate fringing salt marshes on the cobble beaches of New England (USA, North America) and (2) tropical seagrass meadows on the intertidal flats of the Banc d'Arguin (Mauritania, Africa). Both salt marsh and seagrass ecosystems are essential components of coastal zones worldwide, serving as vital habitats for many species, functioning as carbon and nutrient sinks, and playing an important role in coastal protection [29, 30]. Utilizing the natural dynamics and heterogeneity in each ecosystem, we defined three distinct stages of habitat modification. Within each stage, we intensively sampled all species across trophic levels, and reconstructed the food web structure and non-trophic facilitation linkages using stable isotope analyses, mixing models and literature surveys. This allowed the separation of trophic and non-trophic effects of habitat modifiers on overall food

web structure, and the comparison of these effects to those of the most highly connected species in the food web.

Methods

Study sites. The intertidal zone of New England (North America) is typically composed of a top layer of unconsolidated cobbles (5 to 15-cm diameter) deposited by receding glaciers with a coarse, sandy sediment underneath (median grain size: 386 ± 5 μm (mean ± SE); figure S1a). Heat is an important stressor causing mortality in summer as cobbles on these beaches can heat up to over 40 °C [31]. Additionally, cobble movement during storms can crush any organism present [23]. Bare cobble habitat can become colonized by patches of cordgrass (*Spartina alterniflora*) that stabilize the cobbles between their shoot/root system and shade the substrate with their canopy in summer (figure S1b) [23]. As these colonizing cordgrass patches mature and become more established over time, this habitat modification allows ribbed mussels (*Geukensia demissa*) to form dense aggregations (figure S1c). These aggregations further modify conditions by providing hard and stable substrate and crevice space for attachment (for e.g. algae, barnacles), and by cooling the surface through active evapotranspiration [23].

Whereas cordgrass and mussels occur on relatively narrow fringes (<25 m wide) of intertidal cobble beach and modify their habitat at scales of tens of centimeters, habitat modification by seagrasses and crabs occurs at scales of tens of meters within the much larger (>100 ha) intertidal flats of the Banc d'Arguin. Bare habitat is typified by coarse sandy substrate with many dead shells of the bivalve *Senilia senilis* (median grain size: 175 ± 9 μm; figure S1d). This habitat can become colonized by patches of seagrass (*Zostera noltii*) that trap and accumulate fine suspended sediment from the water layer between their roots [32, 33]. In the first few years (i.e. less than ~5 years old) seagrass habitat typically consists of a

mosaic of seagrass patches alternating with bare sediment (figure S1e). Due to sediment trapping, the sandy substrate within seagrass patches becomes covered by a ~5-cm (measured by a gauge rod) thick silt layer (~71% <63µm silt fraction, ~8% organic matter). As seagrass habitat ages, seagrass cover and the thickness of the silt layer gradually increase over time. In long-term established meadows (>40 years old; figure S1f), seagrass cover increases to around 90% and the silt layer reaches a height of ~90 cm. This thick silt layer allows large numbers of swimming crabs (~3300 ha⁻¹), through their intensive burrowing activities, to create large permanently water-filled pools (size up to ~75 m²) in the silt layer that cover ~30% of these areas (see electronic Supplementary Material, text S1, figure S2).

Occasionally, storms and/or ice scour (in New England) and excessive sediment accumulation after e.g. major dust storms (causing overexposure at low tide) followed by erosion (in Banc d'Arguin) reset cordgrass and seagrass habitats to bare cobbles and sand, respectively [32, 34], yielding mosaics of different ecosystem development stages. The 'natural experiments' formed by the resulting habitat mosaics of different stages of ecosystem development provide an excellent opportunity of study how habitat modifiers affect food webs. Based on if and how long an area had been colonized by cordgrass or seagrass (see 'Habitat selection' below), we defined three distinct stages of habitat modification in both ecosystems: (1) bare areas not yet affected by habitat modifiers, (2) colonizing (1 to 4-year-old) areas characterized by primary habitat modifiers (cordgrass/seagrass), but unaffected by secondary habitat modification, and (3) established cordgrass (>10-year-old) and seagrass (>40-year-old) areas that were also affected by secondary habitat modifiers (mussels/crabs).

Habitat selection. For the fringing marshes of Narragansett Bay, Rhode Island, New England (41°35' N; 71°20' W), we combined Google Earth images taken at low tide in 2002, 2004, 2006, 2007, 2008 and 2010 with ground truthing in 2013 to select 4 paired replicate sampling

stations of three habitat types: (1) cobble habitat that had been bare from 2002 onward, (2) 1 to 2 year-old colonizing habitat growing adjacently at the same tidal elevation, and (3) neighboring established cordgrass habitat that was established before 2002. Colonizing habitat sampling stations were selected onsite from expanding edges of established cordgrass patches. The age of these stations was estimated by measuring the distance to the outer edge and calculating the time of colonization by assuming a growth rate of 0.25 to 0.80 cm/day and a 6-month growing season [35].

At Banc d'Arguin (19°53' N; 16°18' W), we used the normalized differences vegetation index (NDVI) calculated from Landsat 5 and 7 images (U.S. Geological Survey) taken at low tide in 1973, 1985, 1994, 1999 to 2003, 2007, 2009 and 2010 combined with ground truthing in 2011 to select 4 replicates for three types of habitat: (1) bare habitat that had been bare from at least 1973 onwards, (2) 2 to 4-year old colonizing seagrass habitat that had become vegetated after 2007 and (3) at least 40-year old established seagrass meadows that were continuously vegetated since at least 1973. As habitat modification effects occur at much larger scales (see 'Study sites') in these seagrass meadows compared to the New England salt marshes, it was not possible to use a paired design here. Instead, to avoid spatial auto-correlation due to environmental gradients, all sampling stations were selected based on a random spatial distribution, with similar elevation, distance to the gully, maximum fetch length and Exposure Index — an integrative measure of wave exposure [36] (figure S2; table S1).

Food web sampling. For each sampling station, we collected and identified all dominant resident species (representing >95% of the biomass in each trophic group and not migrating with the tides) and planktonic sources (see below and table S2), measured nitrogen and

carbon stable isotope values per species, and constructed the trophic interaction matrices based on literature, databases, abundance and isotope data, and mixing models.

Fringing marshes, New England. At each sampling station (~10×25 m), we randomly sampled six replicate plots using a 25×25-cm quadrat in which we determined the number, abundance and size of resident species. Next, we manually collected all epibenthic organisms by hand-picking within the quadrat and took a 5-cm² 5-cm deep sediment sample using a PVC corer for isotopic analysis of sediment particulate organic matter (sPOM). Finally, we sampled for endobenthic species to a depth of 20 cm using a 38-cm² steel corer after which the samples were sieved over a 1-mm mesh. All fauna was identified to species level in the laboratory. Additionally, we collected benthic microbial mats by scraping from rocks at each station. Water column particulate organic matter samples (wPOM) were sampled into 5-L containers, filtered over a 200-μm zooplankton mesh and finally precipitated onto precombusted Whatman GF/F glass fibre filters. Zooplankton was concentrated using a zooplankton net, and subsequently filtered onto pre-combusted Whatman GF/F filters.

Seagrass meadows, Banc d'Arguin. To standardize sampling at the much larger intertidal mudflats, we established a 50-m diameter circle at each sampling station during low tide. Within this circle, we selected 4 replicate areas for sediment and (endo)benthos samples. Sediment samples were taken with a 5-cm deep, 12.5-cm² PVC corer for isotopic analysis of sPOM. Benthos samples were taken with a 179-cm² stainless steel corer to a depth of 20 cm, after which the samples were sieved over a 1-mm mesh. To determine crustacean densities, we took four 5-m long hauls with a 40-cm wide shrimp net at each station. At the established stations, the water column of 4 intertidal pools was separately sampled. Crustaceans were sampled by taking 1 haul with a shrimp net from the edge to the centre of a pool, while fish were sampled by pulling a beach seine net through each pool. All fauna was identified to species level in the laboratory. Additionally, benthic diatoms were scraped from the sediment

surface at each station. After migration through an 80-μm mesh into combusted sand, they were collected in filtered seawater and precipitated onto pre-combusted Whatman GF/F filters. Water column particulate organic matter samples (wPOM) were sampled into 5-L containers, filtered over a 200-μm zooplankton mesh and precipitated onto pre-combusted Whatman GF/F filters. Zooplankton was concentrated using a zooplankton net (mesh size: 200 μm), and subsequently precipitated onto pre-combusted Whatman GF/F filters.

Stable isotope measurements (δ ¹³C **and** δ ¹⁵N). We took muscle tissue samples from fish and soft tissue from invertebrates wherever possible, but used the whole animal for smaller samples. All samples (including primary producers) were rinsed with demineralized water, oven-dried at 50 °C for 48 h and ground. We took sub-samples for separate carbon and nitrogen analyses when samples contained inorganic calcified structures. Samples for carbon analysis were decalcified prior to analysis by addition of 3 M HCl. Stable isotope ratios were measured using an elemental analyzer coupled to an IRMS (Thermo Scientific).

Food web analyses. Based on abundance data we excluded rare observations to include only ecologically relevant species (representing >95% of the biomass in each trophic group). Next, we used published literature, the WoRMS (World Register of Marine Species) database, FishBase and connected online databases, to determine all potential trophic relations for each species and constructed a theoretical, maximized dichotomous interaction matrix for each sampling station that included all potential trophic links. In other words, we first linked each species to all its potential resources. Next, we used size data (i.e. of some species we only found juveniles with a different diet and suite of consumers than adults), stable isotope biplots $(\delta^{15}N)$ versus $\delta^{13}C$, and Bayesian mixing models (R-package SIAR) to estimate the percentage contribution of each potential resource to a consumer's diet at each station [37, 38].

Biplots and mixing models were constructed for each consumer at each sampling station using δ^{13} C and δ^{15} N stable isotope data with at least two replicate measurements per species. Based on these analyses, we constrained the theoretical, maximized matrices by removing trophic links where a resource contributed less than 5% to the diet of the consumer. As such, we only include regular, empirically important consumer-resource interactions, and omit incidental interactions. Finally, we used the constrained trophic interaction matrix to calculate six commonly used measures of food web structure. We use species richness (number of species or food web nodes; S) as an indicator of diversity, link density (number of links per species; L/S) and connectance (C; realized fraction of all possible links; L/S^2) as metrics of topological complexity of the food web, and the percentages of top (species without consumers), basal (species without resources) and intermediate species as trophic distribution metrics [5, 7, 11]. Food web images presented in figures 1 and 2 were constructed using the software *Network3D* [39].

To investigate whether observed differences in food web structure between bare, colonizing and established habitat indeed resulted from non-trophic facilitation as hypothesized, we first compared food web structure between these three habitats. Next, we examined the abiotic habitat requirements of each species in literature and databases (see deposited data). Based on those requirements, we constructed a second dichotomous interaction matrix for each sampling station that included obligatory non-trophic dependencies for each species. Next, we removed non-trophic links and, consequently, the species depending on these links. In other words, we removed those species from the trophic interaction matrix (both rows and columns) that are obligately dependent on non-trophic facilitation by a habitat modifier and compared habitats again using the resulting new food web matrices (figure 1a and b). Specifically, we first focused on secondary habitat modifiers (mussels/crabs) and removed species from the matrix that depend on attachment to mussels in

the established salt marsh habitat and on the intertidal pools formed by crabs in the established seagrass meadows and recalculated all food web metrics. Second, we removed species from the original matrix that depend on primary habitat modifiers (cordgrass/seagrass) in both established and colonizing habitat. In colonizing habitat, these are the species that directly depend on shading or substrate stabilization by cordgrass or on aboveground structure or silt accumulation by seagrass. In established habitat, however, this procedure also resulted in the removal of species that indirectly depend on primary habitat modification through their direct dependence on secondary modification, because primary modification is a prequisite for secondary modification (e.g. pool formation by crabs is impossible without a silt layer accumulated by seagrass).

To test whether food web differences between habitat types could result from trophic rather than non-trophic effects by habitat-modifying species, we removed primary or secondary habitat modifiers themselves from the original matrix and, as a result, also species that fed exclusively on the removed habitat modifier (figure 1a and 1c). Similar to the previous procedure, we then compared the three habitats again using the resulting new matrices.

Finally, to test the importance of non-trophic facilitation by habitat modifiers for food web structure relative to species with a well-documented key trophic position, we compared the trophic role of the most highly connected species in the food web with the non-trophic effects of habitat modifiers [9, 40]. To this end, we removed the species with the highest number of trophic links from the original interaction matrices in colonizing and established habitat respectively, as well as species solely connected to this node (figure 1a and d). Next, we calculated the relative contribution of this species to each food web metric (i.e. 1 – value calculated for modified matrix / value calculated for original matrix). We then did the same calculations for the non-trophic effects of habitat modifiers.

Statistical analyses. To compare between habitats, we used one-way ANOVA for the seagrass meadows for all metrics. For the salt marshes, and to compare relative contributions of the most highly connected species with those of non-trophic effects of habitat modifiers to food web metrics, we applied Generalized Linear Mixed Models (GLMM) with a Gaussian distribution and sampling station as a random factor with Satterthwaite approximation of the degrees of freedom. All model residuals were checked for normality using Shapiro-Wilk tests (p = 0.05). We applied Tukey HSD posthoc tests to detect significant effects between bare, colonizing and established habitat.

Results

Despite obvious differences between the two ecosystems in terms of non-trophic habitat modification and trophic structure, the results showed that food web properties in the salt marsh and seagrass systems had pronounced and remarkably similar responses to the presence of habitat modifiers (figure 2). Both species richness and link density increased as cordgrass and seagrass beds matured (figure 3; tables S3 & S4). Species richness and link density in the salt marsh increased 1.4 and 1.2 times from bare to colonizing cordgrass, respectively, and were another 1.4 and 1.2 times enhanced in established habitats. The seagrass system demonstrated similar and even stronger trends with both species richness and link density increasing by around 1.5 times from bare to colonizing and another 2.5 (species richness) and 1.5 (link density) times from colonizing to established habitats. Connectance followed an opposite trend, decreasing by 0.8 (salt marsh) to 0.6 (seagrass) times from bare to established, probably because the number of links needed to hold the network together relative to all possible links strongly increases as networks become very small. Trophic distribution metrics

were not significantly affected despite the increase in species richness, indicating that habitat modifiers affected species similarly across multiple trophic levels.

The removal procedure of the non-trophic effects of habitat modifiers revealed that 11% of all species depended on primary habitat modification in both systems, and that another 24% and 64% depended on the combined effects of primary and secondary habitat modification in the salt marsh and seagrass system respectively. When non-trophic effects of secondary habitat modifier (mussels/crabs) were excluded, three food web metrics (species richness, link density and connectance) in established habitat became statistically indistinguishable from colonizing habitat, but continued to deviate from bare habitat (figure 3; tables S3 & S4). Moreover, removal of the non-trophic effects of primary habitat modifiers (cordgrass/seagrass) caused these food web metrics in both established and colonizing habitats to converge towards the simplified characteristics of bare, unmodified habitat in both ecosystems, although species richness remained somewhat enhanced in the salt marsh. In contrast, removal of primary or secondary habitat modifiers from the food web caused relatively minor changes in diversity and complexity metrics, demonstrating that the trophic role of habitat modifiers could not explain the observed changes in food web structure from bare to established habitat (tables S3 & S4). Trophic distribution metrics (percentage of top, intermediate and basal species) did not change consistently in response to the removal of nontrophic interactions or trophic interactions from the network.

The comparison of the relative contribution to food web metrics by non-trophic habitat modification versus the most highly connected species demonstrated that habitat modification was much more important for diversity and typically of more, or similar, importance for food web complexity. In both the salt marsh and seagrass system, species richness was more sensitive to the removal of habitat modifiers than to the removal of the most highly connected species (figure 4, table S5). Link density and connectance in established salt marsh and

seagrass habitat were also more affected by primary habitat modification than by the most highly connected species, although effects on link density in the smaller salt marsh food were similar. Because species that are facilitated by habitat modifiers exhibit relatively low levels of trophic connectance compared to the most highly connected species, habitat modifiers and the most highly connected species had opposite effects on connectance in established habitats. Whereas non-trophic interactions by habitat modifiers reduced connectance, this metric was enhanced by the most highly connected species. In colonizing habitat, link density and connectance were more affected by the most highly connected species, whereas in the seagrass system, effects on connectance were similar and link density was more affected by habitat modification. Finally, the effects on trophic distribution metrics varied depending on both the ecosystem (salt marsh or seagrass) and habitat type (established or colonzing). In general, however, we found these metrics to be more affected by habitat modifiers in the seagrass system, whereas the importance of habitat modifiers and the most highly connected species were similar in the salt marsh (table S5).

Discussion

Overall, our findings demonstrate that habitat modification strongly changes food web structure – not only by facilitating species and thus enhancing diversity, but also by increasing the number of trophic interactions that species have with other species in the food web. We found these non-trophic, indirect effects of habitat modifiers to be much more pronounced than their trophic roles. Furthermore, depending on the metric and habitat type, we found habitat modification of more or similar importance compared to the trophic role of the most highly connected species in the food web. This is important because the most highly connected node has been repeatedly documented to have a key structuring role in food webs as well in networks in general [9, 40], thus emphasizing the importance of habitat

modification for food web structure in our study systems. Finally, although trophic distribution metrics were affected by habitat modifiers, we did not identify consistent shifts in the percentage of top, intermediate and basal species as we removed the non-trophic effects of habitat modifiers. This implies that their modification of environmental conditions alters food web structure across multiple trophic levels, for example by affecting the outcome of trophic and competitive interactions between species.

Studies from a wide range of ecosystems have shown that amelioration of physical stress by habitat-modifying organisms can profoundly impact the associated community by facilitating other species [16], and recent work demonstrated that such effect may not only be local [41]. On cobble beaches, for instance, many studies (including this one) have revealed non-trophic facilitation of the local community [23, 42], and that at a scale of meters to tens of meters, cordgrass patches function as wave breaks to facilitate wave-sensitive forb species [43]. Similarly, intertidal mussel beds in the Wadden Sea were found to facilitate the community at a scale exceeding 100 m by baffling waves and changing sediment characteristics [24, 44]. Although we focused on habitat modifier-effects within a single habitat type, the contrasting spatial scales at which non-trophic facilitation impacted food webs in salt marshes (<1 m) and seagrass meadows (>10 m) respectively, suggest that habitatmodifying species can affect not only a few species, but also whole food web dynamics beyond the habitat in which they grow. Furthermore, the investigated habitat types reflect distinct stages of ecosystem succession and changes in food webs structure can largely be explained by habitat modification (figure 3). This illustrates that non-trophic interactions can play a key role in moderating community assembly as well as in maintaining complex food webs. In addition to habitat provision and alleviation of physical stress, habitat modification may also indirectly stimulate the development of complex food webs by increasing productivity and mediating how energy and nutrients move through ecosystems. Habitat

modification thus affects important factors that control the stability of assembling food webs during succession [12, 45].

Until now, food web structure and stability have been typically analyzed as a function of the properties of the trophic network itself, like the number of species and links, connectance, or the strength of loops [7-12]. Our findings indicate that, in ecosystems dominated by habitat-modifying organisms, those properties themselves can be highly contingent on interactions outside the trophic network, yet intrinsic to the species participating in it. Most likely, this is not only the case in ecosystems where these organisms drive succession as investigated here, but also in systems where habitat modifiers create alternative stable states by facilitating their own growth or survival [46]. We therefore suggest that integrating non-trophic interactions in food web analyses is important in increasing their explanatory and predictive capacity, and consider our approach as a vital first step in that direction. We propose that future work should focus on the development of quantitative hybrid networks of multiple interaction types, firmly grounded in empirical data. One potential problem in this regard is the large diversity of non-trophic interactions observed in real ecosystems. A first conceptual approach to overcome this issue may be to construct dynamic models, in which trophic interactions are modeled as energy flows or consumerresource interactions, and non-trophic interactions are integrated using functional classes defined by how trophic interactions are modified [4, 19].

Increased predictive capacity is urgently needed by society because ecosystems dominated by habitat modifiers like seagrass meadows, salt marshes, coral reefs, peatlands and rainforests are now degrading at alarming rates worldwide due to anthropogenic disturbances with relatively low success of restoration efforts [29, 30, 47]. Our findings suggest that the development of the typically complex species interaction networks of these systems can take decades due to long-term cumulative effects of multiple habitat modifiers.

This implies that ecosystem conservation and restoration efforts should not focus on trophic interactions alone (e.g. only on recovery of top predators), but like future approaches to network analyses, should consider various interaction types and potential synergistic or antagonistic effects between them.

Data accessibility

Supplementary Material is available online. The data are deposited in DRYAD at: http://datadryad.org/submit?journalID=RSPB&manu=RSPB-2015-2326.

Competing interests

The authors declare no competing interests.

Author contributions

EvdZ, TvdH, CA, HO, AHA and BRS generated hypotheses and designed field sampling.

EvdZ, TvdH, CA, LLG, KJvdR, JAvG, MJAC, and MvdG collected field data. EvdZ, TvdH, and CA conducted analyses, and wrote the first draft of the manuscript. All authors contributed substantially to revisions.

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References

- [1] Darwin, C. 1859 On the origin of species by means of natural selection, or the preservation of favoured races in the struggle of life. London, John Murray.
- [2] Dunne, J.A., Williams, R.J. & Martinez, N.D. 2002 Food-web structure and network theory: the role of connectance and size. *Proc. Natl. Acad. Sci. USA* **99**, 12917-12922. (doi:10.1073/pnas.192407699).
- [3] Allesina, S., Alonso, D. & Pascual, M. 2008 A general model for food web structure. *Science* **320**, 658-661. (doi:10.1126/science.1156269).
- [4] Kéfi, S., Berlow, E.L., Wieters, E.A., Navarrete, S.A., Petchey, O.L., Wood, S.A., Boit, A., Joppa, L.N., Lafferty, K.D., Williams, R.J., et al. 2012 More than a meal... integrating non-feeding interactions into food webs. *Ecol. Lett.* **15**, 291-300. (doi:10.1111/j.1461-0248.2011.01732.x).
- [5] May, R.M. 1972 Will a large complex system be stable? *Nature* **238**, 413-414. (doi:10.1038/238413a0).
- [6] Pimm, S.L., Lawton, J.H. & Cohen, J.E. 1991 Food web patterns and their consequences. *Nature* **350**, 669-674. (doi:10.1038/350669a0).
- [7] Williams, R.J. & Martinez, N.D. 2000 Simple rules yield complex food webs. *Nature* **404**, 180-183. (doi:10.1038/35004572).
- [8] Neutel, A.M., Heesterbeek, J.A.P. & de Ruiter, P.C. 2002 Stability in real food webs: weak links in long loops. *Science* **296**, 1120-1123. (doi:10.1126/science.1068326).

- [9] Dunne, J.A., Williams, R.J. & Martinez, N.D. 2002 Network structure and biodiversity loss in food webs: robustness increases with connectance. *Ecol. Lett.* **5**, 558-567. (doi:10.1046/j.1461-0248.2002.00354.x).
- [10] Bascompte, J., Melian, C.J. & Sala, E. 2005 Interaction strength combinations and the overfishing of a marine food web. *Proc. Natl. Acad. Sci. USA* **102**, 5443-5447. (doi:10.1073/pnas.0501562102).
- [11] Gross, T., Rudolf, L., Levin, S.A. & Dieckmann, U. 2009 Generalized models reveal stabilizing factors in food webs. *Science* **325**, 747-750. (doi:10.1126/science.1173536).
- [12] Neutel, A.M., Heesterbeek, J.A.P., van de Koppel, J., Hoenderboom, G., Vos, A., Kaldeway, C., Berendse, F. & de Ruiter, P.C. 2007 Reconciling complexity with stability in naturally assembling food webs. *Nature* **449**, 599-U511. (doi:10.1038/nature06154).
- [13] Stouffer, D.B. & Bascompte, J. 2011 Compartmentalization increases food-web persistence. *Proc. Natl. Acad. Sci. USA* **108**, 3648-3652. (doi:10.1073/pnas.1014353108).
- [14] Dunne, J.A., Lafferty, K.D., Dobson, A.P., Hechinger, R.F., Kuris, A.M., Martinez, N.D., McLaughlin, J.P., Mouritsen, K.N., Poulin, R., Reise, K., et al. 2013 Parasites affect food web structure primarily through increased diversity and complexity. *PLoS. Biol.* 11, e1001579. (doi:10.1371/journal.pbio.1001579).
- [15] Stachowicz, J.J. 2001 Mutualism, facilitation, and the structure of ecological communities. *Bioscience* **51**, 235-246. (doi: 10.1641/0006-3568(2001)051[0235:MFATSO]2.0.CO;2).
- [16] Bruno, J.F., Stachowicz, J.J. & Bertness, M.D. 2003 Inclusion of facilitation into ecological theory. *Trends Ecol. Evol.* 18, 119-125. (doi:10.1016/s0169-5347(02)00045-9).
 [17] Olff, H., Alonso, D., Berg, M.P., Eriksson, B.K., Loreau, M., Piersma, T. & Rooney, N. 2009 Parallel ecological networks in ecosystems. *Philos. Trans. R. Soc. B-Biol. Sci.* 364, 1755-1779. (doi:10.1098/rstb.2008.0222).

- [18] Baiser, B., Whitaker, N. & Ellison, A.M. 2013 Modeling foundation species in food webs. *Ecosphere* **4**, art146. (doi:10.1890/ES13-00265.1).
- [19] Sanders, D., Jones, C.G., Thébault, E., Bouma, T., van der Heide, T., van Belzen, J. & Barot, S. 2014 Integrating ecosystem engineering and food webs. *Oikos* 123, 513–524. (doi: 10.1111/j.1600-0706.2013.01011.x).
- [20] Kéfi, S., Berlow, E.L., Wieters, E.A., Joppa, L.N., Wood, S.A., Brose, U. & Navarrete, S.A. 2015 Network structure beyond food webs: mapping non-trophic and trophic interactions on Chilean rocky shores. *Ecology* **96**, 291-303. (doi: 10.1890/13-1424.1).
- [21] Dayton, P. 1972 Toward an understanding of community resilience and the potential effects of enrichments to the benthos at McMurdo Sound, Antarctica. In *Proceedings of the Colloquium on Conservation Problems in Antarctica* (pp. 81–95 Lawrence, Kansas, USA, Allen Press.
- [22] Jones, C.G., Lawton, J.H. & Shachak, M. 1994 Organisms as ecosystem engineers. *Oikos* **69**, 373-386. (doi: 10.2307/3545850).
- [23] Altieri, A.H., Silliman, B.R. & Bertness, M.D. 2007 Hierarchical organization via a facilitation cascade in intertidal cordgrass bed communities. *Am. Nat.* **169**, 195-206. (doi:10.1086/510603).
- [24] Van der Zee, E.M., van der Heide, T., Donadi, S., Eklof, J.S., Eriksson, B.K., Olff, H., van der Veer, H.W. & Piersma, T. 2012 Spatially extended habitat modification by intertidal reef-building bivalves has implications for consumer-resource interactions. *Ecosystems* **15**, 664-673. (doi:10.1007/s10021-012-9538-y).
- [25] Van der Heide, T., Eklof, J.S., van Nes, E.H., van der Zee, E.M., Donadi, S., Weerman, E.J., Olff, H. & Eriksson, B.K. 2012 Ecosystem engineering by seagrasses interacts with grazing to shape an intertidal landscape. *PLoS One* 7, e42060. (doi:10.1371/journal.pone.0042060).

- [26] Angelini, C. & Silliman, B.R. 2013 Secondary foundation species as drivers of trophic and functional diversity: evidence from a tree-epiphyte system. *Ecology* **95**, 185-196 (doi:10.1890/13-0496.1).
- [27] Van der Heide, T., Tielens, E., van der Zee, E.M., Weerman, E.J., Holthuijsen, S., Eriksson, B.K., Piersma, T., van de Koppel, J. & Olff, H. 2014 Predation and habitat modification synergistically interact to control bivalve recruitment on intertidal mudflats. *Biol. Conserv.* **172**, 163-169. (doi:10.1016/j.biocon.2014.02.036).
- [28] Van der Zee, E.M., Tielens, E., Holthuijsen, S., Donadi, S., Eriksson, B.K., van der Veer, H.W., Piersma, T., Olff, H. & van der Heide, T. 2015 Habitat modification drives benthic trophic diversity in an intertidal soft-bottom ecosystem. *J. Exp. Mar. Biol. Ecol.* **465**, 41-48. (doi:10.1016/j.jembe.2015.01.001).
- [29] Waycott, M., Duarte, C.M., Carruthers, T.J.B., Orth, R.J., Dennison, W.C., Olyarnik, S., Calladine, A., Fourqurean, J.W., Heck, K.L., Hughes, A.R., et al. 2009 Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proc. Natl. Acad. Sci. USA* **106**, 12377-12381. (doi:10.1073/pnas.0905620106).
- [30] Gedan, K.B., Silliman, B.R. & Bertness, M.D. 2009 Centuries of human-driven change in salt marsh ecosystems. *Annu. Rev. Mar. Sci.* 1, 117-141. (doi:10.1146/annurev.marine.010908.163930).
- [31] Bertness, M.D. 1989 Intraspecific competition and faciliation in a Northern acron barnacle population. *Ecology* **70**, 257-268. (doi:10.2307/1938431).
- [32] Folmer, E.O., van der Geest, M., Jansen, E., Olff, H., Anderson, T.M., Piersma, T. & van Gils, J.A. 2012 Seagrass-sediment feedback: an exploration using a non-recursive structural equation model. *Ecosystems* **15**, 1380-1393. (doi:10.1007/s10021-012-9591-6).
- [33] Van der Heide, T., Govers, L.L., de Fouw, J., Olff, H., van der Geest, M., van Katwijk, M.M., Piersma, T., van de Koppel, J., Silliman, B.R., Smolders, A.J.P., et al. 2012 A three-

- stage symbiosis forms the foundation of seagrass ecosystems. *Science* **336**, 1432-1434. (doi:10.1126/science.1219973).
- [34] Ewanchuk, P.J. & Bertness, M.D. 2003 Recovery of a northern New England salt marsh plant community from winter icing. *Oecologia* **136**, 616-626. (doi:10.1007/s00442-003-1303-7).
- [35] Newton, C. & Thornber, C. 2013 Ecological impacts of macroalgal blooms on salt marsh communities. *Estuaries Coasts* **36**, 365-376. (doi:10.1007/s12237-012-9565-0).
- [36] Howes, D., Morris, M. & Zacharia, M. 1999 British Columbia estuary mapping system. Coastal Task Force Resource Inventory Committee Secretariat.
- [37] Parnell, A.C., Inger, R., Bearhop, S. & Jackson, A.L. 2010 Source partitioning using stable isotopes: coping with too much variation. *PLoS One* **5**, e9672. (doi:10.1371/journal.pone.0009672).
- [38] Parnell, A. & Jackson, A. 2011 SIAR: Stable Isotope Analysis in R. R package version 4.1. 3.
- [39] Williams, R.J. 2010 Network3D [Software]. Cambridge, UK. Microsoft Research.[40] Jordan, F. 2009 Keystone species and food webs. *Philos. Trans. R. Soc. B-Biol. Sci.* 364,
- 1733-1741. (doi:10.1098/rstb.2008.0335).
- [41] Van de Koppel, J., van der Heide, T., Altieri, A.H., Eriksson, B.K., Bouma, T.J., Olff, H. & Silliman, B.R. 2015 Long-distance interactions regulate the structure and resilience of coastal ecosystems. *Annu. Rev. Mar. Sci.* 7, 139-158. (doi:10.1146/annurev-marine-010814-015805).
- [42] Angelini, C., van der Heide, T., Griffin, J.N., Morton, J.P., Derksen-Hooijberg, M., Lamers, L.P.M., Smolders, A.J.P. & Silliman, B.R. 2015 Foundation species' overlap enhances biodiversity and multifunctionality from the patch to landscape scale in southeastern

United States salt marshes. *Proc. R. Soc. B-Biol. Sci.* **282**, 20150421. (doi:10.1098/rspb.2015.0421).

- [43] Van de Koppel, J., Altieri, A.H., Silliman, B.R., Bruno, J.F. & Bertness, M.D. 2006 Scale-dependent interactions and community structure on cobble beaches. *Ecol. Lett.* **9**, 45-50. (doi:10.1111/j.1461-0248.2005.00843.x).
- [44] Donadi, S., van der Heide, T., van der Zee, E.M., Eklof, J.S., van de Koppel, J., Weerman, E.J., Piersma, T., Olff, H. & Eriksson, B.K. 2013 Cross-habitat interactions among bivalve species control community structure on intertidal flats. *Ecology* **94**, 489-498. (doi: 10.1890/12-0048.1).
- [45] Moore, J.C., Deruiter, P.C. & Hunt, H.W. 1993 Influence of productivity on the stability of real and model ecosystems. *Science* **261**, 906-908. (doi:10.1126/science.261.5123.906). [46] Scheffer, M., Carpenter, S., Foley, J.A., Folke, C. & Walker, B. 2001 Catastrophic shifts in ecosystems. *Nature* **413**, 591-596. (doi:10.1038/35098000).
- [47] Rands, M.R.W., Adams, W.M., Bennun, L., Butchart, S.H.M., Clements, A., Coomes, D., Entwistle, A., Hodge, I., Kapos, V., Scharlemann, J.P.W., et al. 2010 Biodiversity conservation: challenges beyond 2010. *Science* **329**, 1298-1303. (doi:10.1126/science.1189138).

Figure Legends

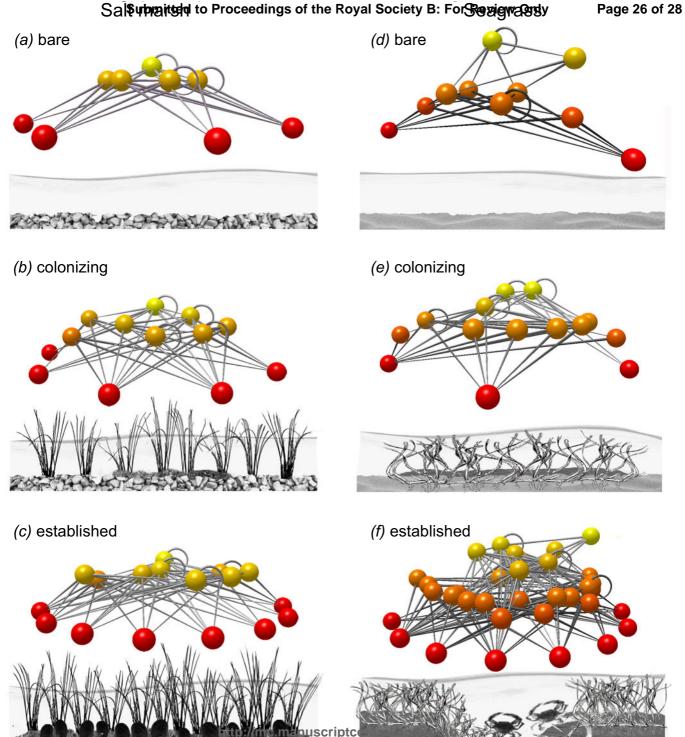
Figure 1. Conceptual representation of performed removal procedures. (*a to b*) To test the hypothesis that observed differences in food web structure between habitats resulted from non-trophic facilitation by a habitat modifier, we first removed its non-trophic effect by removing species that depend obligatorily on its non-trophic facilitation (e.g. as attachment substrate). (*a to c*) Second, to test whether food webs differed due to the trophic effects of the habitat modifier, we deleted it and species exclusively feeding on it from the food web. (*a to*

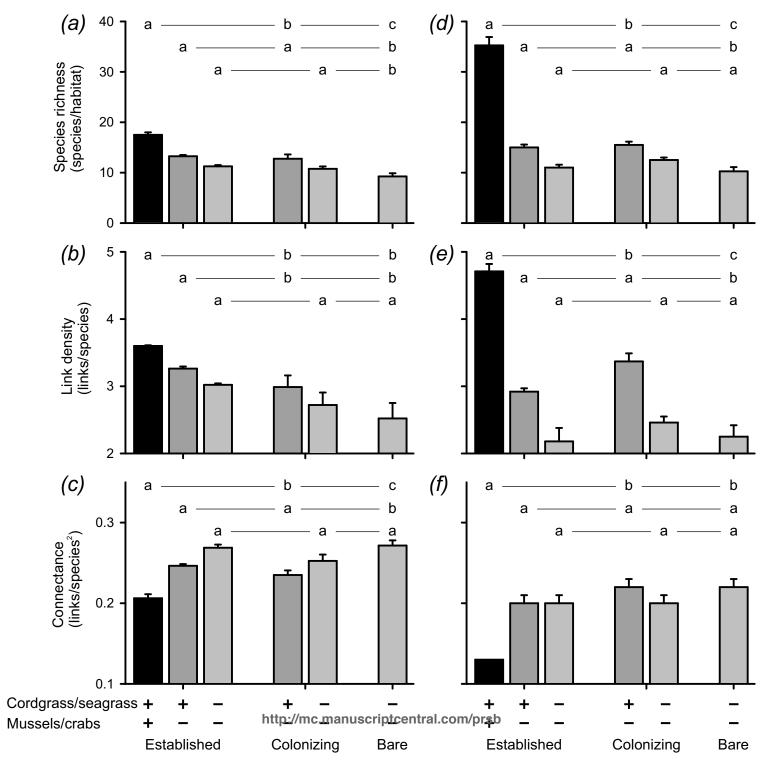
d) Third, we compared the trophic and non-trophic effects by habitat modifiers to those of the most highly connected species.

Figure 2. Salt marsh and seagrass system food webs in the absence and presence of primary (cordgrass/seagrass) and secondary (mussels/crabs) habitat modifiers. Bare sites are typified by relatively simple food webs (a, d). Food webs have higher species richness and link density in colonizing habitats with primary habitat modifiers (b, e) and these effects are further enhanced by secondary habitat modifiers in established habitats (c, f). Node color changes from red (basal species) to yellow with increasing trophic level.

Figure 3. Species richness (a, d), link density (b, e) and connectance (c, f) as conditional on primary and secondary habitat modifiers. For each habitat, the most left bar indicates the natural situation, and subsequent bars depict the outcomes of the removal of species dependent on habitat modification (i.e. removal of non-trophic effects). Letters indicate posthoc grouping; error bars represent SEM. Species richness and link density are significantly higher in modified habitat, whereas connectance is generally lower. Removal of species dependent on habitat modification increased similarity to unmodified, bare habitat.

Figure 4. The relative effect of secondary and primary habitat modification, and the most highly connected species on species richness (*a*, *d*), link density (*b*, *e*) and connectance (*c*, *f*). Letters indicate statistical grouping; error bars represent SEM. The analyses shows that habitat modification was more important for diversity and of more or similar importance for complexity.





Supplementary Material

Text S1 – Origin of the intertidal pools in Banc d'Arguin

Methods. Whereas the habitat modification effects by cordgrass, ribbed mussels and seagrass have all been well investigated [1-10], the mechanism driving intertidal pool formation at Banc d'Arguin was not clear yet. Therefore, we combined GIS analyses, field surveys, and an experiment to determine their origin.

We first investigated the hypothesis that the pools are formed by an interaction between seagrass silt accretion and the abiotic environment. Previous studies suggest that depressions in seagrass meadows may form due to "blow-outs" caused by storm events [11] or that they are simply remnants of old creeks that have not yet filled up [12]. For both hypotheses, this would mean that, after formation, these structures should either (1) migrate through the meadows due to directional disturbance (i.e. erosion) and regrowth dynamics [11] or (2) gradually decrease in size over time due to sediment accretion caused by dense seagrass stands in and around these pools, as previously demonstrated with experimental pools created in these meadows [12]. Therefore, we compared the spatial position and pool size development over a 7-year period based on Google Earth images from Quickbird (October 2004) and Geoeye (March 2011), both taken at low tide. Using Google Earth Pro, we measured the surface area of 60 clearly visible pools at 5 established areas on the October 2004 image and subsequently, measured the surface areas of the same pools again on the March 2011 image. The number of pools per hectare, their size, depth and relative cover was determined at each established site during field surveys.

Second, we tested the hypothesis that bioturbating organisms burrowing in the silt layer created the pools. Literature and explorative field surveys suggested bioturbating fish, waterbirds and crabs as potential candidates. As a first indication, we therefore started with surveys determining their densities.

Fish surveys. Based on literature, we suspected rays to be one of the more important bioturbating fish species [13, 14]. We determined their abundance on the tidal flats by visual surveys from a boat along random transects, within 10 meters of each side of the front of the boat at a maximum speed of 5 km h⁻¹. Transect lengths were determined using GPS. Fourteen transects (~250-300 m long) were carried out by boat at three established sites (E1, E3 & E4; Fig. S2a; 42 transects in total) in October 2012. Rays were identified up to species level whenever possible, but other taxonomic classes (family, order) were occasionally used when identification up to species level was not possible. We used the average number of rays per transect per site as a measure of abundance per hectare on the tidal flats.

To determine the abundance of rays and other potentially bioturbating fish species in the intertidal pools, we placed underwater cameras (GoPro HD HERO2, Woodman Labs Inc, USA) in pools at three established sites. Per site, the size of three pools were measured and subsequently monitored by two cameras taking photographs at 10-s intervals across one full high tide period. All fish species that entered the pools were marked as potentially bioturbating species. Each fish was identified to species level whenever possible and other

taxonomic classes (family, order) were used when identification up to species level was not possible. Next, we calculated the abundance per hectare pool during high tide for each species.

Waterbird surveys. Waterbird abundance was measured in 60×100 m quadrats marked with PVC poles at three established sites (E1, E3 & E4; Fig. S2a). During low tide, we scored the number of feeding waterbirds in and outside pools every 15 minutes in the quadrats from a distance of ~400 m, using a telescope (zoom ocular 20-60×; ATM 80 HD, Swarovski, Absam, Austria). Only bird species that were large enough to walk in the pools or could enter them by swimming or diving were counted during our observations. Counting started when the water had retreated from the marked square until the area was inundated again. Birds were counted during three complete tidal cycles in October 2012. Based on these data, we calculated the abundance per hectare during low tide on the tidal flats and in the pools for each species.

Crab surveys. We started by comparing crab densities at all four selected bare, colonizing and established sites. We set out two 50-m long and 10-m wide transects at each site and counted the number of crab (Callinectes marginatus) holes as a proxy for crab density and activity. At three of our established sites (E1, E3 & E4; Fig. S2a), we measured pool size and depth of 18, 31 and 11 intertidal pools, respectively. Within these pools, the number of crab holes was scored to determine the average crab density per pool and the relation between number of crabs per pool and pool size.

Experiment. Finally, we experimentally tested whether crabs would be capable of initiating pools in undisturbed, healthy seagrass meadows. In three consecutive experiments, we pushed a circular cage 25 cm into the sediment of a healthy, undisturbed meadow (diameter: 125 cm; height: 50 cm; mesh size of 1 cm) and added a crab (carapace 7.6 ± 0.4 cm) that was caught in the direct surroundings of the cage. We determined the length and width of crab holes after 24 hours.

Results. Established meadows were typified by a high number of intertidal pools in the silt layer (82.8 ± 9.2 pools ha⁻¹, 0.19 ± 0.01 m depth (mean \pm SE)) that combined covered 30% (\pm 4%) of these intertidal habitats. To find the mechanism driving intertidal pool formation in the silt layer of established seagrass meadows, we first explored the hypotheses that (1) pools are formed by (migrating) "blow-outs" caused by storm events [11], or (2) that they are remnants of old intertidal creeks that have not yet filled up [12]. Using satellite image analyses, we found that the pools were spatially stagnant between 2004 and 2011, and had increased in size (mean size₂₀₀₄= 37.6 \pm 2.4 m², mean size₂₀₁₁= 47.0 \pm 2.7 m²; t = -3.307; p < 0.01; n = 60). Because experimental pools in these meadows have previously been shown to quickly disappear due to sediment accumulation in the seagrass meadows [12], our outcomes suggest that the pools do not simply form by an interaction between seagrass silt accretion and the abiotic environment.

Second, we investigated whether bioturbating animals play a significant role in the formation and maintenance of the intertidal pools. We started by determining densities of bioturbating crabs, fish and bird species. We found an average ray density of 0.60 ± 0.28 individuals ha⁻¹ during high tide on the intertidal flats and an average density of 0.21 ± 0.07 individuals ha⁻¹ in the pools (Table S6). In addition, observations with underwater cameras in the pools yielded an average of 4.08 ± 1.90 individuals ha⁻¹ pool during high tide of other

potentially bioturbating fish. Average bird densities were 0.63 ± 0.22 individuals ha⁻¹ during low tide on the tidal flats and 0.07 ± 0.02 individuals ha⁻¹ in the tidal pools (Table S7).

In contrast to the rather low numbers of birds and fish in the pools, field surveys revealed very high abundances of burrowing crabs (*Callinectes marginatus*) in the established meadows (3300 ± 427 holes ha⁻¹) – also in comparison to colonizing (308 ± 54 holes ha⁻¹) and bare areas (0 ± 0 holes ha⁻¹). Moreover, we found a strong positive correlation between crab hole numbers within the pools and pool size ($R^2 = 0.74$; t = 17.1; p < 0.001; n = 60; Fig. S3). Finally, our onsite cage experiment demonstrated that, within 24 hours, crabs were able to dig a hole (0.05 ± 0.02 m²; n = 3) in the silt layer of an undisturbed seagrass meadow, illustrating that crabs can create pools rather than 'just' colonizing them. Overall, we conclude that bioturbation by crabs in the thick silt layer, possibly exacerbated by feeding birds and fish, is the most likely mechanism driving the formation of intertidal pools at Banc d'Arguin.

Table S1. Environmental characteristics of the three habitat types at the New England cobble beaches were kept similar through pairwise selection of habitats (distance between habitat types was ~5 to 20 m, and at the same tidal elevation; see *Habitat Selection* in *Methods*). At Banc d'Arguin, distances between sampling areas were much larger (200 to >2000 m between habitat types). Banc d'Arguin sites were therefore selected based on a random spatial distribution, excluding dispersal limitation as a potential explanation for differences in food web structure (Fig. S2a), and similar elevation, distance to the gully, maximum fetch and exposure index. We used one-way ANOVA to test for differences in relative elevation, distance to the gulley, and maximum fetch and a Kruskal Wallis test for exposure index.

				_	
	Mean values ± S		Statistics		
	Established (E)	Colonizing (C)	Bare (B)	F or χ^2	p
Relative elevation (cm)	-255±5	-247±15	-263±11	0.53	0.609
Distance to gully (km)	0.09 ± 0.03	0.14 ± 0.04	0.09 ± 0.03	0.66	0.541
Maximum fetch (km)	20.85±10.45	8.61 ± 1.20	7.77 ± 2.02	1.69	0.238
Exposure index (class 0-5)	1.75 ± 0.63	0.5 ± 0.5	0.5 ± 0.5	3.04	0.219

Df. (error) of ANOVA tests: 2 (9).

Table S2. Overview of the sampling methodology per sampling site.

	Fringing marshes	Intertidal seagrass meadows
Water column	wPOM (0.7-200 μm)	wPOM (0.7-200 μm)
	Zooplankton (>200 μm)	Zooplankton (>200 μm)
		4x beach seine net in intertidal pools
		4x shrimp net haul edge to center in pools
Epibenthos	Flora/fauna: 6x collection in 25×25 quadrat	Flora/fauna: 4x collection from 179 cm ² core
	Microbial mat: 6x collection from rocks	Fauna: 4x 5-m shrimp net haul
		Microbial mat: 4x using migration method
	Additional manual collection of organisms with insufficient biomass for stable isotopes	Additional manual collection of organisms with insufficient biomass for stable isotopes
Endobenthos	Fauna: 6x 20-cm deep, 38-cm ² core sieved	Fauna: 4x 20-cm deep 179 cm ² core sieved
	across 1-mm mesh	across 1-mm mesh
	sPOM: 6x 5-cm deep, 5-cm ² core	sPOM: 4x 5-cm deep, 12.5-cm ² core
	Additional manual collection of organisms	Additional manual collection of organisms
	with insufficient biomass for stable isotopes	with insufficient biomass for stable isotopes

Table S3. Change in food web structure indicators in New England salt marshes along an empirical gradient of food web complexity (left to right: established to bare), and the simulated removal of non-trophic and trophic effects by secondary (mussels) and primary (cordgrass) habitat-modifying species, respectively.

	Mean values ± SE			Statist	ics	
	Established (E)	Colonizing (C)	Bare (B)	F	p	Posthoc groups (E, C, B)
All effects included (i.e	. original matrix)				
Number of species (S)	17.75 ± 0.48	12.75 ± 0.85	9.25 ± 0.63	109.5	***	a, b, c
Links per species (L/S)	3.62 ± 0.02	2.99 ± 0.17	2.52 ± 0.23	14.1	**	a, b, b
Connectance (L/S ²)	0.203 ± 0.005	0.235 ± 0.006	0.271 ± 0.006	35.3	***	a, b, c
Basal species (%)	46.5 ± 0.7	39.7 ± 2.6	43.8 ± 2.8	2.6	ns	
Intermediate spec. (%)	33.9 ± 0.9	42.7 ± 2.4	34.3 ± 4.2	3.3	ns	
Top species (%)	19.6±1.2	17.6±1.0	21.9±1.4	3.2	ns	
Non-trophic effects of 1	nussels removed					
Number of species (S)	13.25 ± 0.25	12.75±0.85	9.25 ± 0.63	14.3	**	a, a, b
Links per species (L/S)	3.26 ± 0.03	2.99 ± 0.17	2.52 ± 0.23	6.0	*	a, ab, b
Connectance (L/S ²)	0.246 ± 0.002	0.235 ± 0.006	0.271 ± 0.006	12.5	***	a, a, b
Basal species (%)	37.8 ± 0.7	39.7 ± 2.6	43.8±2.8	2.2	ns	
Intermediate spec. (%)	45.3±0.8	42.7±2.4	34.3±4.2	5.3	*	a, a, b
Top species (%)	16.9±1.5	17.6±1.0	21.9±1.4	4.3	*	a, ab, b
Non-trophic effects of c	cordgrass remove	ed				
Number of species (S)	11.25 ± 0.25	10.75 ± 0.48	9.25 ± 0.63	8.7	*	a, a, b
Links per species (L/S)	3.02 ± 0.02	2.72 ± 0.19	2.52 ± 0.23	2.6	ns	
Connectance (L/S ²)	0.269 ± 0.004	0.252 ± 0.008	0.271 ± 0.006	4.9	ns	
Basal species (%)	44.5 ± 0.9	46.8 ± 2.0	43.8 ± 2.8	0.8	ns	
Intermediate spec. (%)	35.6 ± 0.7	32.4±1.5	34.3 ± 4.2	0.3	ns	
Top species (%)	20.0±1.7	20.8±1.5	21.9±1.4	0.4	ns	
Trophic effects of muss	sels removed					
Number of species (S)	16.75 ± 0.48	11.75 ± 0.85	8.75 ± 0.85	73.5	***	a, b, c
Links per species (L/S)	3.60 ± 0.02	2.90 ± 0.19	2.41 ± 0.28	11.2	**	a, b, b
Connectance (L/S ²)	0.214 ± 0.005	0.247 ± 0.006	0.274 ± 0.007	27.6	***	a, b, c
Basal species (%)	49.3±0.8	43.2 ± 3.0	47.0 ± 4.4	1.1	ns	
Intermediate spec. (%)	35.8±1.0	46.4 ± 2.4	33.1±4.5	6.5	*	ac, b, c
Top species (%)	14.9±1.4	10.4±1.3	20.0±2.3	10.8	*	ac, b, c
Trophic effects of cord	grass removed					
Number of species (S)	16.75 ± 0.48	11.75 ± 0.85	9.25 ± 0.63	87.5	***	a, b, c
Links per species (L/S)	3.72 ± 0.02	3.07 ± 0.18	2.52 ± 0.23	15.8	**	a, b, c
Connectance (L/S ²)	0.222 ± 0.006	0.262 ± 0.007	0.271 ± 0.001	15.5	**	a, b, b
Basal species (%)	44.8±1.9	34.6 ± 2.4	43.8±2.8	6.3	*	ac, b, c
Intermediate spec. (%)	34.3±1.3	46.4 ± 2.4	34.3 ± 4.2	5.8	**	ac, b, c
Top species (%)	20.1±1.2	19.1±1.0	21.9±2.8	1.4	ns	

Df. (error) of all tests: 2 (7). * p<0.05, ** p<0.01, *** p<0.001.

Table S4. Change in food web structure indicators in the tropical seagrass system Banc d'Arguin along an empirical gradient of food web complexity (left to right: established to bare), and the simulated removal of non-trophic and trophic effects by secondary (crabs) and primary (seagrass) habitat-modifying species, respectively.

	Mean values ± SE			Statistics		
	Established (E)	Colonizing (C)	Bare (B)	F	p	Posthoc groups (E, C, B)
All effects included (i.e.	. original matrix)					
Number of species (S)	35.25±1.65	15.50 ± 0.65	10.25 ± 0.85	134.5	***	a, b, c
Links per species (L/S)	4.71 ± 0.11	3.37 ± 0.12	2.25 ± 0.17	46.35	***	a, b, c
Connectance (L/S ²)	$0.13\pm 3e-3$	0.22 ± 0.01	$0.22\pm4e-3$	74.18	***	a, b, b
Basal species (%)	21.4±1.2	24.1±2.4	26.6±3.5	1.0	ns	
Intermediate spec. (%)	48.1±3.5	61.5±3.4	41.8 ± 9.8	2.5	ns	
Top species (%)	30.5±2.8	14.4±1.1	31.6±11.2	2.1	ns	
Non-trophic effects of c	erabs removed					
Number of species (S)	14.50 ± 0.50	15.50 ± 0.65	10.25 ± 0.85	16.70	***	a, a, b
Links per species (L/S)	2.92±0.05	3.37±0.12	2.25 ± 0.17	20.23	***	a, a, b
Connectance (L/S ²)	0.20 ± 0.01	0.22 ± 0.01	$0.22\pm4e-3$	2.06	ns	
Basal species (%)	32.8±1.8	24.1±2.4	26.6±3.5	2.9	ns	
Intermediate spec. (%)	53.3±1.9	61.5±3.4	41.8±9.8	2.6	ns	
Top species (%)	17.4 ± 2.3	14.4±1.1	31.6±11.2	1.9	ns	
- Non-trophic effects o	f seagrass remov	ed				
Number of species (S)	11.00±0.58	12.50±0.5	10.25±0.85	3.00	ns	
Links per species (L/S)	2.18±0.09	2.46 ± 0.09	2.25±0.17	1.16	ns	
Connectance (L/S ²)	0.20 ± 0.01	0.20 ± 0.01	$0.22\pm4e-3$	1.29	ns	
Basal species (%)	43.3±2.3	29.8±22.8	26.6±3.5	9.5	**	a, b, b
Intermediate spec. (%)	15.8 ± 4.0	38.0 ± 4.0	41.8±9.8	4.6	*	a, ab, a
Top species (%)	50.0±5.3	40.2 ± 3.7	31.6±11.2	1.5	ns	
Trophic effects of crabs	s removed					
Number of species (S)	34.25±1.65	14.50 ± 0.65	10.25±0.85	127.0	***	a, b, b
Links per species (L/S)	4.27±0.23	2.96 ± 0.10	2.25±0.17	35.33	***	a, b, c
Connectance (L/S^2)	$0.13\pm 3e-3$	0.21 ± 0.01	$0.22\pm4e-3$	117.4	***	a, b, b
Basal species (%)	21.3±1.1	25.7±2.6	26.6±3.5	1.0	ns	
Intermediate spec. (%)	41.7±2.4	58.8±3.5	41.8±9.8	2.6	ns	
Top species (%)	36.3±2.6	15.4±1.2	31.6±11.2	2.7	ns	
Trophic effects of seagi	ass removed					
Number of species (S)	34.25±1.65	14.50 ± 0.65	10.25±0.85	127.0	***	a, b, b
Links per species (L/S)	4.50±0.21	3.46±0.13	2.25±0.17	42.8	***	a, b, c
Connectance (L/S^2)	0.13±2e-3	0.24 ± 0.01	0.22±4e-3	86.2	***	a, b, b
Basal species (%)	19.1±1.1	18.8±2.7	26.6±3.5	2.9	ns	, ,
Intermediate spec. (%)	49.6±3.6	65.8±3.7	41.8±9.8	3.7	ns	
Top species (%)	31.4±2.8	15.4±1.8	31.6±11.2	1.9	ns	

Df. (error) of all tests: 2 (9). * p<0.05, ** p<0.01, *** p<0.001. ns = not significant.

Table S5. Relative effect of non-trophic facilitation by primary and secondary habitat modifiers, and that of the trophic role of the most highly connected species for established and colonizing habitat of both ecosystems.

Colonizing natitation	Mean values ± SE			Statistics		
	Secondary habitat modifier (hm2)		High. conn. species (hc)		p	Posthoc groups (hm2, hm1, hc)
Fringing marshes			species (iie)			(2,,)
Established						
Number of species (S)	0.25±0.02	0.37 ± 0.02	0.06±1e-3	156.9	***	a, b, c
Links per species (L/S)	0.10±5e-3	0.17±3e-3	0.16±3e-3	215.0	***	a, b, b
Connectance (L/S ²)	-0.21 ± 0.03	-0.32 ± 0.03	0.11±3e-3	135.1	***	a, b, c
Basal species	0.19±3e-3	0.04±7e-3	-0.06±2e-3	1570.3	***	a, b, c
Intermediate spec.	-0.34 ± 0.04	-0.05 ± 0.03	$0.47\pm1e-3$	345.5	***	a, b, c
Top species	0.14 ± 0.05	-0.01±0.05	-0.68 ± 0.05	126.8	***	a, b, c
Colonizing						
Number of species (S)		0.15±0.02	0.08±5e-3	9.8	*	
Links per species (L/S)		0.09 ± 0.01	0.19 ± 0.01	161.6	***	
Connectance (L/S^2)		-0.08 ± 0.03	-0.12±7e-3	47.3	**	
Basal species		-0.18±0.03	-0.09±6e-3	9.2	*	
Intermediate spec.		0.24 ± 0.03	0.50 ± 0.03	938.1	***	
Top species		-0.18 ± 0.03	-1.04 ± 0.13	68.4	**	
1 1						
Seagrass meadows						_
Established						
Number of species (S)	0.59 ± 0.01	0.69 ± 0.01	$0.03\pm1e-3$	1155.5	***	a, b, c
Links per species (L/S)	0.35 ± 0.03	0.52 ± 0.03	$0.06\pm 9e-3$	121.8	***	a, b, c
Connectance (L/S^2)	-0.54±0.06	-0.52±0.08	0.04 ± 0.01	78.7	***	a, a, b
Basal species	-0.54±0.10	-1.04±0.14	$-0.03\pm1e-3$	40.5	***	a, b, c
Intermediate spec.	-0.04±0.05	0.84 ± 0.10	$-0.03\pm1e-3$	75.4	***	a, b, a
Top species	0.42 ± 0.10	-0.73±0.33	0.07 ± 0.01	9.8	*	a, b, a
Colonizing						
Number of species (S)		0.19±0.02	0.06±3e-3	31.6	**	
Links per species (L/S)		0.19 ± 0.02 0.26 ± 0.04	0.16 ± 0.02	5.7	ns	
Connectance (L/S^2)		0.09 ± 0.03	0.10 ± 0.02 0.10 ± 0.02	0.2	ns	
Basal species		-0.24 ± 0.04	$-0.07\pm3e-3$	23.6	**	
Intermediate spec.		0.51 ± 0.06	0.13 ± 0.09	11.6	*	
Top species		-1.87 ± 0.44	-0.47 ± 0.41	5.4	ns	
1 op species		1.0/-0.77	J. 17-U.TI	٥.١	110	

Df. (error) of all tests: 2 (9). * p<0.05, ** p<0.01, *** p<0.001. ns = not significant.

Table S6. Ray and other fish densities (number ha⁻¹ during high tide) at three established sites at Banc d'Arguin. Densities were determined for the number of rays on the tidal flats and for the number of rays and other fish species present in the pools at the same tidal flat.

	Mean values ± SE					
Ray-Fish species						
Tidal flat	Site E1	Site E3	Site E4	Total		
Dasyatis marmorata	0.11±0.11	0±0	0.85±0.85	0.32±0.27		
Rhinobatos sp	1.09 ± 0.43	0.79 ± 0.37	0.78 ± 0.53	0.89 ± 0.10		
Total				0.60 ± 0.28		
Pool						
Dasyatis marmorata	0.23 ± 0.15	0.43±0.43	0.18±0.18	0.28±0.08		
Rhinobatos sp	0.31±0.25	0.04 ± 0.04	0.09 ± 0.06	0.15 ± 0.08		
Total				0.21±0.07		
Arius heudelotii	0.19±0.11	2.88±2.77	7.31±3.99	2.28±1.07		
Dicentrarchus punctatus	1.34 ± 1.34	2.88 ± 1.95	0.15 ± 0.15	1.46 ± 0.79		
Diplodus sp	4.80 ± 3.80	6.55 ± 6.55	0.03 ± 0.03	3.80 ± 2.00		
Ephippion guttiferum	0.27 ± 0.12	0.60 ± 0.60	0.02 ± 0.02	0.30 ± 0.17		
Epinephelus aeneus	0.31 ± 0.26	0.94 ± 0.94	7.05 ± 3.55	2.76 ± 2.5		
Epinephelus sp	0 ± 0	0 ± 0	1.18 ± 0.54	0.39 ± 0.39		
Mugil cephalus	3.51 ± 3.45	33.36±32.38	26.97 ± 19.87	21.28 ± 9.07		
Rhyzoprionodon acutus	0.04 ± 0.04	0 ± 0	0 ± 0	0.01 ± 0.01		
Sciaena umbra	0.04 ± 0.04	0.04 ± 0.04	1.13 ± 0.88	0.40 ± 0.36		
Tilapia guineesis	0 ± 0	28.60±28.40	0 ± 0	9.54 ± 9.54		
Umbrina canariensis	0±0	0±0	7.90 ± 6.60	2.63 ± 2.63		
Total				4.08±1.90		

Table S7. Waterbirds densities (number ha⁻¹ during low tide) at three established sites at Banc d'Arguin. Densities were determined for the number of waterbirds foraging on the tidal flats and for the number of birds foraging in the pools within the same tidal flat.

Mean values ± SE						
Waterbird species						
Tidal flat	Site E1	Site E3	Site E4	Total		
Phoenicopterus roseus	0±0	0±0	0.04 ± 0.04	0.01±0.01		
Pelecanus onocrotalus	0.04 ± 0.04	0 ± 0	0 ± 0	0.01 ± 0.01		
Egretta gularis	0.59 ± 0.21	0.16 ± 0.16	0.48 ± 0.14	0.36 ± 0.18		
Ardea cinerea monicae	0.07 ± 0.04	0 ± 0	0.12 ± 0.06	0.06 ± 0.03		
Egretta garzetta	0.09 ± 0.09	0.09 ± 0.06	0.04 ± 0.03	0.07 ± 0.01		
Platalea leucorodia (balsaci)	0 ± 0	0.03 ± 0.02	0.14 ± 0.09	0.06 ± 0.04		
Numenius arquata	0.69 ± 0.11	0.44 ± 0.19	1.27 ± 0.40	0.80 ± 0.24		
Limosa lapponica	2.71 ± 0.66	1.09 ± 0.37	7.07 ± 2.67	3.62 ± 1.79		
Total				0.63±0.22		
Pool						
Phoenicopterus roseus	0 ± 0	0 ± 0	0.04 ± 0.04	0.01 ± 0.01		
Pelecanus onocrotalus	0 ± 0	0 ± 0	0 ± 0	0 ± 0		
Egretta gularis	0.09 ± 0.06	0 ± 0	0.11 ± 0.04	0.07 ± 0.03		
Ardea cinerea monicae	0 ± 0	0 ± 0	0 ± 0	0 ± 0		
Egretta garzetta	0 ± 0	0.05 ± 0.03	0 ± 0	0.02 ± 0.02		
Platalea leucorodia (balsaci)	0 ± 0	0.03 ± 0.02	0.07 ± 0.07	0.03 ± 0.02		
Numenius arquata	0.22 ± 0.09	0.13 ± 0.07	0.16 ± 0.09	0.17 ± 0.03		
Limosa lapponica	0.18 ± 0.05	0.13 ± 0.05	0.57±0.49	0.29 ± 0.14		
Total				0.07±0.02		

Supporting figures

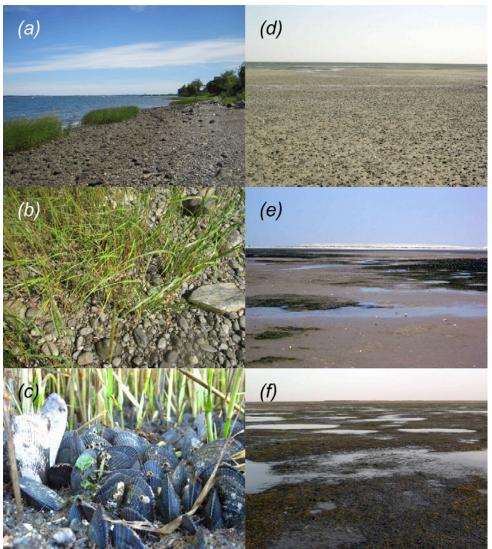


Figure S1. Illustration of habitat types in fringing marshes (New England, US) and intertidal seagrass meadows (Banc d'Arguin, Mauritania). (a) New England salt marshes occur along relatively narrow fringes of cobble beach (<25 m wide). (b) Unconsolidated cobbles become stabilized and shaded in colonizing patches of cordgrass. (c) In established patches, ribbed mussels provide attachment substrate for sessile organisms and crevice space. (d) Banc d'Arguin consists of large intertidal, sandy mudflats of over 100 ha. (e) Colonizing patches of seagrass trap silty sediment from the water layer between their roots. (f) In established areas, crabs burrow large intertidal pools in the silt layer accumulated by seagrass.

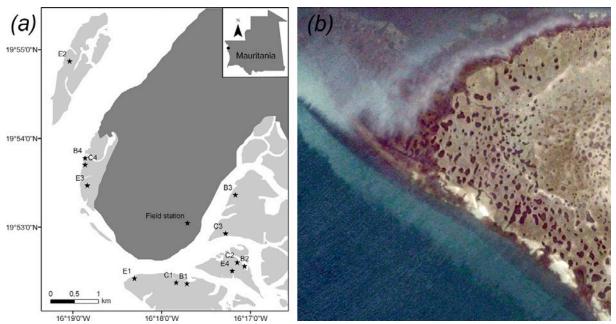


Figure S2. (a) Map of the study area in the Banc d'Arguin, Mauritania. White areas indicate water, intermediate grey areas indicate mudflats exposed during low water and land is represented by dark grey. Stars represent the positions of the different study sites and the PNBA field station. (b) Satellite image showing large numbers of intertidal pools (black) during low tide in the established (>40 years) seagrass meadows at E1.

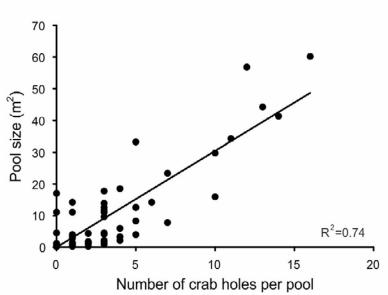


Figure S3. Positive linear relationship (y=3.04x; t=17.1; R^2 =0.74; p<0.001; n=60) between number of crab holes per pool and pool size.

References

- [1] Bertness, M.D. 1989 Intraspecific competition and faciliation in a Northern acron barnacle population. *Ecology* **70**, 257-268. (doi:10.2307/1938431).
- [2] Stephens, E.G. & Bertness, M.D. 1991 Mussel facilitation of barnacle survival in a sheltered bay habitat. *J. Exp. Mar. Biol. Ecol.* **145**, 33-48. (doi:10.1016/0022-0981(91)90004-g).
- [3] Bertness, M.D., Leonard, G.H., Levine, J.M., Schmidt, P.R. & Ingraham, A.O. 1999 Testing the relative contribution of positive and negative interactions in rocky intertidal communities. *Ecology* **80**, 2711-2726.
- (doi:10.1890/0012-9658(1999)080[2711:ttrcop]2.0.co;2).
- [4] Sousa, W.P. 1979 Disturbance in marine intertidal boulder fields: the nonequilibrium maintenance of species diversity. *Ecology* **60**, 1225-1239. (doi:10.2307/1936969).
- [5] Altieri, A.H., Silliman, B.R. & Bertness, M.D. 2007 Hierarchical organization via a facilitation cascade in intertidal cordgrass bed communities. *Am. Nat.* **169**, 195-206. (doi:10.1086/510603).
- [6] Van der Heide, T., van Nes, E.H., Geerling, G.W., Smolders, A.J.P., Bouma, T.J. & van Katwijk, M.M. 2007 Positive feedbacks in seagrass ecosystems implications for success in conservation and restoration. *Ecosystems* **10**, 1311-1322. (doi: 10.1007/s10021-007-9099-7).
- [7] Altieri, A.H., van Wesenbeeck, B.K., Bertness, M.D. & Silliman, B.R. 2010 Facilitation cascade drives positive relationship between native biodiversity and invasion success. *Ecology* **91**, 1269-1275. (doi:10.1890/09-1301.1).
- [8] Van der Heide, T., van Nes, E.H., van Katwijk, M.M., Olff, H. & Smolders, A.J.P. 2011 Positive feedbacks in seagrass ecosystems evidence from large-scale empirical data. *PLoS One* **6**, e16504. (doi:10.1371/journal.pone.0016504).
- [9] Van der Heide, T., Govers, L.L., de Fouw, J., Olff, H., van der Geest, M., van Katwijk, M.M., Piersma, T., van de Koppel, J., Silliman, B.R., Smolders, A.J.P., et al. 2012 A three-stage symbiosis forms the foundation of seagrass ecosystems. *Science* **336**, 1432-1434. (doi:10.1126/science.1219973).
- [10] Folmer, E.O., van der Geest, M., Jansen, E., Olff, H., Anderson, T.M., Piersma, T. & van Gils, J.A. 2012 Seagrass-sediment feedback: an exploration using a non-recursive structural equation model. *Ecosystems* **15**, 1380-1393. (doi:10.1007/s10021-012-9591-6).
- [11] Patriquin, D.G. 1975 Migration of blowouts in seagrass beds at Barbados and Carriacou, West Indies and its ecological and geological implications. *Aquat. Bot.* **1**, 163-189. (doi: 10.1016/0304-3770(75)90021-2).
- [12] Van der Laan, B. & Wolff, W.J. 2006 Circular pools in the seagrass beds of the Banc d'Arguin, Mauritania, and their possible origin. *Aquat. Bot.* **84**, 93-100. (doi:10.1016/j.aquabot.2005.07.009).
- [13] Orth, R.J. 1975 Destruction of eelgrass, *Zostera marina*, by the cownose ray, *Rhinoptera bonasus* in the Chesapeake Bay. *Chesapeake Sci.* **16**, 205-208. (doi: 10.2307/1350896).
- [14] Townsend, E.C. & Fonseca, M.S. 1998 Bioturbation as a potential mechanism influencing spatial heterogeneity of North Carolina seagrass beds. *Mar. Ecol.-Prog. Ser.* **169**, 123-132. (doi:10.3354/meps169123).