

# Salinity appears to be the main factor shaping spatial COI diversity of *Corbicula* lineages within the Chinese Yangtze River Basin

Cong Zeng<sup>1,2</sup>  | Yangxin Tang<sup>1</sup>  | Martin Vastrade<sup>3</sup> | Neil E. Coughlan<sup>4</sup> | Ting Zhang<sup>5</sup> | Yongjiu Cai<sup>6</sup> | Karine Van Doninck<sup>3,7</sup> | Deliang Li<sup>1</sup>

<sup>1</sup>Hunan Engineering Technology Research Center of Featured Aquatic Resources Utilization, College of Animal Science and Technology, Hunan Agricultural University, Changsha, China

<sup>2</sup>School of Oceanography, Shanghai Jiao Tong University, Shanghai, China

<sup>3</sup>Laboratory of Evolutionary Genetics and Ecology; Research Unit in Environmental and Evolutionary Biology; Institute of Life, Earth and Environment (ILEE), University of Namur, Namur, Belgium

<sup>4</sup>School of Biological, Earth and Environmental Sciences, University College Cork, Cork, Ireland

<sup>5</sup>College of Chemistry and Chemical Engineering, Central South University, Changsha, China

<sup>6</sup>Nanjing Institute of Geography and Limnology, Chinese Academy of Sciences, Nanjing, China

<sup>7</sup>Molecular Biology & Evolution, Université Libre de Bruxelles, Brussels, Belgium

## Correspondence

Deliang Li, Hunan Engineering Technology Research Center of Featured Aquatic Resources Utilization, College of Animal Science and Technology, Hunan Agricultural University, Changsha, China. Email: [lidl@hunau.edu.cn](mailto:lidl@hunau.edu.cn)

## Funding information

National Natural Science Foundation of China, Grant/Award Number: 31772832; National Key Research and Development Program of China, Grant/Award Number: 2019YFD090060306; Hunan Provincial Modern Agricultural Research System, Grant/Award Number: 2019-105;

## Abstract

**Aim:** To date, few studies have examined the phylogenetics of *Corbicula* clams in their native range and the environmental parameters influencing their distribution, although this could provide great insights into the biological adaptation and invasion dynamics of *Corbicula* clams. We sought to identify the genetic lineages of native *Corbicula* clams and elucidate the environmental factors shaping the distributions of identified lineages.

**Location:** China, mainly the Yangtze River Basin.

**Methods:** The alignment comprised 558 COI sequences including samples from China and 222 COI sequences from published studies. This dataset was used to generate phylogenetic trees and compare population diversity. We used dbRDA method to assess the relationship between these COI data and environmental factors measured to identify the important factors affecting *Corbicula*'s distribution.

**Results:** The COI phylogenetic tree delineated the monophyly of 3 major COI clades and 77 distinct COI haplotypes in the Yangtze River Basin. The invasive lineage C/S (FW17) was not identified in our sampling in China, while invasive lineages A/R (FW5 = Hap6), B (FW1 = Hap17) and Rlc (FW4 = Hap43) were identified, abundant and widely distributed in the middle and lower reaches of Yangtze River. Focusing on populations from Yangtze River Basin, both the COI haplotype and nucleotide diversity in the lakes along the Yangtze River increased with longitude, except for the river mouth population. The AMOVA tests showed significant differentiation between the middle and lower reaches of the Yangtze River and among populations. The dbRDA results suggested that the parameter chloride explained most of the spatial COI haplotype distribution variation in the Yangtze River Basin, with the three invasive lineages tolerating broad fluctuations of salinity (chloride levels ranging from 4 to 60 mg/L).

**Conclusions:** The *Corbicula* COI haplotypes found within the Yangtze River Basin had distinct distribution preferences, with the invasive androgenetic lineages being the

Cong Zeng and Yangxin Tang should be considered joint first authors.

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. *Diversity and Distributions* published by John Wiley & Sons Ltd.

Youth Innovation Promotion Association  
CAS, Grant/Award Number: 2020316;  
Irish Research Council, Grant/Award  
Number: GOIPD/2022/861

Editor: Elizabeta Briski

most abundant and widely distributed. Genetic diversity was higher in this native region than in invaded areas in Europe and America, while it decreased with increasing distance from the river mouth. Salinity appeared to be the main environmental factor shaping the *COI* haplotype distribution of *Corbicula* lineages within their native range.

#### KEYWORDS

Asian clam, evolutionary adaptation, genetic diversity, haplotype distribution, lineages, oceanic origin

## 1 | INTRODUCTION

Biological invasions have been unambiguously shown to be among the major causes of global biodiversity decline (Courchamp et al., 2017; Diagne et al., 2021). Genus *Corbicula*, also known as *Corbicula* complex, is considered one of the most ecologically and economically costly freshwater invaders (Haubrock et al., 2022; Pigneur, Falisse, et al., 2014; Strayer, 2010). The original distribution of *Corbicula* (*Bivalvia*, *Cyrenidae*; formerly *Corbiculidae*) seems to be confined to Asia, Africa and Australia (Araujo et al., 1993); since the mid-20th century, they have established a global distribution that spans all continents except the Antarctic (Bespalaya et al., 2018; Crespo et al., 2015). In general, *Corbicula* clams are regarded as high-impact freshwater invaders due to their ability to detrimentally alter community and ecosystem dynamics, through nutrient cycling and energy flow, phytoplankton depletion and competition for resources and substrate modification (Pigneur, Falisse, et al., 2014; Sousa et al., 2014). Its diverse reproductive strategies (androgenetic hermaphrodite, sexual dioecism, cross-fertilization and self-fertilization) is another issue of interest in their invasion biology (Pigneur et al., 2012). Although the rapid expansion of *Corbicula* across its invasive range is mostly linked to anthropogenic-mediated dispersal (Coughlan et al., 2017), the invasive *Corbicula* lineages all appear to be hermaphrodites reproducing through androgenesis, and one individual is sufficient to establish a new population (reviewed in Pigneur et al., 2012). These *Cyrenidae* are therefore considered a potential model taxon for the general study of rapid bivalve radiations into inland environments (Graf, 2013).

Few studies have attempted to assess the phylogenetics and phylogeography of sympatric *Corbicula* clams within their native habitats, in Asia, Africa or Australia. As a result, the evolutionary processes that have shaped historical radiation and adaptation in *Corbicula*, as well as their contemporary interactions with environmental preferences, remain unclear. One of the dominant drivers of species distribution is habitat (McDowell et al., 2014). Because of their global distribution and high invasive efficiency (Crespo et al., 2015), the *Corbicula*'s habitat preference for environmental factors such as temperature, turbidity and salinity has been studied (Avelar et al., 2014; McMahon, 1979; Müller & Baur, 2011; Sousa et al., 2006). Low temperature (<2°C), high temperature (>25°C) and high turbidity (>150 nephelometric turbidity units, NTU) could cause

a decline in their survival rates (Avelar et al., 2014; McMahon, 1979; Müller & Baur, 2011), and it was also reported that changes in salinity could influence their abundance and biomass (Sousa et al., 2006). The consequences of these environmental factors could directly shape their distribution and colonization (Ferreira-Rodríguez et al., 2017), but it remains unclear whether different effects exist between species (lineages) within the genus.

Despite an almost global distribution, the taxonomy of this genus is poorly understood, primarily due to the extraordinary range of morphological variation and unique reproductive modes, including both sexual dioecy and androgenetic hermaphrodite species (Gomes et al., 2016; Komaru et al., 2013; Peñarrubia et al., 2017; Pigneur et al., 2011). In recent years, with the development of sequencing methods, molecular markers have been employed to solve taxonomical difficulties (e.g. Bespalaya et al., 2018; Haponski & Ó Foighil, 2019; Vastrade et al., 2022). Two *Corbicula* mitochondrial lineages are now considered to be widely distributed within Asian freshwater environments using *COI*, forms A/R and B (Park & Kim, 2003; Pigneur, Etoundi, et al., 2014; Wang et al., 2014). However, discrepancies remain between genetic and morphological identifications (Lee et al., 2005; Siripattawan et al., 2000), as well as between nuclear and mitochondrial markers (Hedtke et al., 2008; Lee et al., 2005; Peñarrubia et al., 2017; Pfenninger et al., 2002; Vastrade et al., 2022). The taxonomical identification of *Corbicula* also remains difficult because of their androgenetic mode of reproduction and polyploidy.

Despite the limitations of relying on a single molecular marker, the widely used *COI* marker has shown to be reliable for *Corbicula*, and was used in this study. Hence, in this study, we sampled *Corbicula* clams from the middle and lower Yangtze River Basin and produced mitochondrial *COI* sequences, downloaded available *COI* sequences of *Corbicula* clams from the NCBI database and identified the mitochondrial lineages present in the native and non-native regions. The spatial distribution of these lineages was further investigated in the populations of the middle and lower Yangtze River, and environmental indicators were used to investigate the drivers influencing their distributions. We tested the hypothesis of whether salinity was the main factor influencing the distribution of *Corbicula* in the Yangtze River Basin. These results will improve our understanding of the adaptive mechanisms of their invasion and provide information for biodiversity conservation in invaded areas to control their colonization.

## 2 | METHODS

### 2.1 | Sample collection

Corbicula specimens were collected with hand net or dredge from seven freshwater lakes and the Yangtze River mouth (YzM,  $n = 60$ ). Sampled lakes included five of the largest freshwater lakes in China: Chao Lake (ChL,  $n = 4$ ), Dongting Lake (DoL,  $n = 126$ ), Hongze Lake (HzL,  $n = 20$ ), Poyang Lake (PoL,  $n = 19$ ) and Tai Lake (TaL,  $n = 30$ ), as well as two other lakes situated within the Yangtze River Basin: Datong Lake (DaL,  $n = 55$ ) and Dianshan Lake (DsL,  $n = 22$ ) (see Figure S1 and S2a–g for the location of sampled lakes and estuary and sampling localities within the lakes, respectively, and Table S1 for the sampling time). Lakes Datong (DaL), Dongting (DoL) and Poyang (PoL) are located in the middle reaches of the Yangtze River Basin, while the other four lakes are situated in the lower reaches. Currently, only Dongting Lake and Poyang Lake are directly connected with the Yangtze River, the other lakes are indirectly connected through a network of tributary rivers, except Hongze Lake. Located in the lower reaches of the Huai River Basin, Hongze Lake is directly connected to the Yangtze River by the Beijing–Hangzhou Grand Canal. Specimens were washed on-site using a 0.5 mm mesh sieve with clean water and were immediately preserved in ethanol until DNA extraction.

### 2.2 | DNA extraction and sequencing

Genomic DNA was extracted from each specimen's mantle tissue using the TIANamp tissue DNA kit (Tiangen Biotech Co., Ltd.). A 710-base-pair (bp) fragment of the mitochondrial *COI* gene was then amplified using the universal primers LCO1490/HCO2198 (Folmer et al., 1994). PCRs were conducted in a final reaction volume of 25  $\mu$ l consisting of 100 ng template DNA, 25 mM MgCl<sub>2</sub>, 1 $\times$  PCR buffer, 2.5 mM each dNTP, 10  $\mu$ M each of primers, 2.5 U of Taq DNA polymerase (Tiangen) and ddH<sub>2</sub>O. The reactions were conducted in an Applied Biosystems 2720 thermal cycler (Singapore), with an initial denaturation step at 94°C for 5 min, followed by 35 cycles at 94°C for 1 min, 50°C for 1 min and 72°C for 1 min, with a final extension step of 72°C for 5 min. Following this, 3  $\mu$ l of PCR products were electrophoresed and visualized on 1% ethidium bromide (EB)-stained agarose gels to confirm single products. PCR products were purified using a TIANgel Midi Purification Kit (Tiangen Biotech Co., Ltd.). Products were sequenced by Sangon Biotechnology (Shanghai) Co., Ltd. (China) using Applied Biosystems™ 3730xl DNA Analyzer, and the sequences were subjected to quality control independently in Geneious (R11, Biomatters Ltd.), BioEdit (Hall, 1999) and Sequencher 4.1.4 (Gene Codes Corporation).

### 2.3 | Phylogenetic analysis and haplotype network

As morphological traits cannot be relied upon for taxonomical identifications in *Corbicula* due to phenotypic plasticity, *COI*

sequences were used here to identify lineages within this complex. To examine the phylogenetic relationship of *Corbicula* clams, other *COI* sequences from specimens collected throughout China were obtained from the NCBI database and the literature, including a few reference sequences (Table 1). However, as the sequences obtained from GenBank varied in length, only sequences with a length >550 bp were considered in our analyses. All sequences from Chinese *Corbicula* individuals were aligned using the plugin ClustalW Alignment (Thompson et al., 1994) in Geneious (Gap Open Cost = 100 and Gap Extend Cost = 10; R11, Biomatters Ltd.). Moreover, the *COI* sequences of the four identified invasive lineages in Europe and America (Hedtke et al., 2008; Park & Kim, 2003; Pigneur et al., 2011; Pigneur, Etoundi, et al., 2014; Siripattawan et al., 2000; Vastrade et al., 2022), being FW5 (form A/R, NCBI ID: AF196268) (Siripattawan et al., 2000), FW1 (form B, AF196269) (Siripattawan et al., 2000), FW4 (form R/c, AF269096) (Renard et al., 2000) and FW17 (form C/S, AF269095) (Renard et al., 2000), were added to the alignment and phylogenetic reconstruction to compare native Chinese and invasive *Corbicula* specimens using *COI*. Furthermore, *Corbicula* spp. not previously documented in China were also included in the alignment and *COI* phylogeny: *C. sandai* (AF196272), *C. japonica* (AF196271) (Siripattawan et al., 2000), *C. loehensis* (AY275666), *C. matannensis* (AY275663), *C. moltkiana* (AY275657), *C. possoensis* (AY275661) (Glaubrecht et al., 2003) and *Batissa violacea* (Outgroup, DQ837727) (Glaubrecht et al., 2006).

All sequences in the alignment were trimmed to the same length. *COI* haplotypes were generated using DnaSP v 6.12.03 (Rozas et al., 2017), while the haplotype network was produced using the median-joining network method with POPART 1.7 (Leigh & Bryant, 2015). Phylogenetic relationships were studied between the *COI* haplotypes by constructing a maximum-likelihood (ML) phylogenetic tree. The best nucleotide substitution model was selected based on the lowest AICc value by ModelFinder (Kalyaanamoorthy et al., 2017) and was used to build the maximum-likelihood phylogenetic tree using IQ-TREE (Nguyen et al., 2015).

Furthermore, to visualize the spatial genetic variation and the abundance of *Corbicula* clams residing within the middle and lower reaches of the Yangtze River Basin, haplotypes from DnaSP were mapped along the Yangtze River using ArcGIS 10.2 (ESRI Inc).

### 2.4 | Genetic diversity

This study focused on *Corbicula* from the middle and lower reaches of the Yangtze River, and only populations associated with the Yangtze River were retained for the following analyses, including ChL, DaL, DsL, DoL, HzL, PoL, TaL, and YzM. *COI* genetic diversity was evaluated for all the *Corbicula* specimens recovered from the Yangtze River Basin (Table 3), the analyses computed the number of haplotypes ( $h$ ), the haplotypic diversity ( $hd$ ) and the nucleotide diversity ( $\pi$ ) in DnaSP.

Considering the sampling size, which is uneven here, the ChL ( $n = 4$ ) population was further excluded in the structure analysis to

**TABLE 1** Sources of *Corbicula* specimens and sequences used in the present study

Sites	Code	Specimens (collected in this study)	Sequences (downloaded from NCBI)	Total
(a) Samples and sequences from China				
Chao Lake	ChL	4	0	4
Dongting Lake	DoL	126	16 <sup>a</sup>	142
Dianshan Lake	DsL	22	0	22
Datong Lake	DaL	55	0	55
Fujiang River	FuR	0	30 <sup>c</sup>	30
Hong Kong	HoK	0	3 <sup>d</sup>	3
Hongze Lake	HzL	20	78 <sup>e</sup>	98
Poyang Lake	PoL	19	14 <sup>a</sup>	33
Qingshan Lake	QiL	0	13 <sup>a</sup>	13
Tai Lake	TaL	30	36 <sup>c</sup>	66
Taiwan	TaW	0	28 <sup>a</sup>	28
Yalu River	YaR	0	1 <sup>f</sup>	1
Yangtze River Mouth	YzM	60	0	60
Yunan	YuN	0	3 <sup>g</sup>	3
Total		336	222	558
(b) Sequences for references				
<i>Corbicula loehensis</i>	AY275666	0	1	4 <sup>h</sup>
<i>C. matannensis</i>	AY275663	0	1	
<i>C. moltkiana</i>	AY275657	0	1	
<i>C. possoensis</i>	AY275661	0	1	
<i>C. japonica</i>	AF196271	0	1	2 <sup>i</sup>
<i>C. sandai</i>	AF196272	0	1	
FW1 Form B	AF196269	0	1	3 <sup>a</sup>
FW4 Form Rlc	AF269096	0	1	
FW5 Form A/R	AF196268	0	1	
FW17 Form C/S	AF269095	0	1	1 <sup>j</sup>
<i>Batissa violacea</i>	DQ837727	0	1	1 <sup>h</sup>

Note: Specimens sampled and sequences retrieved from GenBank from China, published data are marked with subscript letters, references: <sup>a</sup>Park & Kim, 2003; <sup>b</sup>Wang et al., 2014; <sup>c</sup>Iida et al., 2012; <sup>d</sup>Pfenninger et al., 2002; <sup>e</sup>Li et al., 2015; <sup>f</sup>Yamada et al., 2014; <sup>g</sup>Pigneur, Etoundi, et al., 2014; <sup>h</sup>Glaubrecht et al., 2003; <sup>i</sup>Siripattawan et al., 2000; <sup>j</sup>Renard et al., 2000. Sequences available in GenBank for reference, citations and accession numbers are provided.

avoid bias. The Yangtze River is usually divided into middle (=Middle) and lower (=Lower) reaches by the Hukou (which is the boundary for middle and lower reaches of Yangtze River), so this study assigned DaL, DoL and PoL into Middle Yangtze River Basin and the other lakes (DsL, HzL, TaL and YzM) into Lower Yangtze River Basin. An analysis of molecular variance (AMOVA) was performed to determine the amount of genetic variability using *F*-statistics among geographic populations and among clades. The significance of the covariance components associated with the different possible levels of genetic structure was tested by 10,000 permutations. AMOVA was performed in Arlequin v3.5.2.2 (Excoffier & Lischer, 2010). For the uneven sampling size, we performed another AMOVA analysis considering the populations with sample size >30 for all populations and populations of clade II (see Table S2a,b) and compared

the results with the first analysis. Both analyses yielded similar results and therefore the first analysis was retained within this study (Table 4).

## 2.5 | Environmental parameter measurements and statistical analyses

To assess the extent to which *Corbicula* haplotypes and environmental parameters influence the spatial distribution of *Corbicula* COI lineages, 18 environmental parameters were measured, including temperature (Temp), transparency (Trans), pH, dissolved oxygen (DO), conductivity (Cond), Chlorophyll-a (Chla), turbidity (Turb), chloride (Cl), calcium (Ca), magnesium (Mg), potassium (K), sodium

(Na), total phosphorus (TP), total nitrogen (TN), nitrate ( $\text{NO}_3^-$ -N), nitrite ( $\text{NO}_2^{2-}$ -N), ammonium ( $\text{NH}_4^+$ -N), orthophosphate ( $\text{PO}_4^{3-}$ -P) and permanganate index ( $\text{COD}_{\text{Mn}}$ ), for Chao Lake, Datong Lake, Dianshan Lake, Dongting Lake, Hongze Lake, Poyang Lake and Tai Lake. These 18 environmental parameters for each lake were collected at different times, and the specific sampling details are shown in Table S1 and Figure S2, and finally, the average of all time points was used for subsequent analysis (Table 2). Temp, pH, DO, Cond and Turb were measured using an YSI 6600 V2-4 Multi-Parameter Water Quality Sonde, and Trans was measured in the field using Secchi disk. Chloride (Cl) was determined by ion chromatography, and Ca, Mg, K and Na were measured using ICP-AES. Samples for the determination of  $\text{NH}_4^+$ -N,  $\text{NO}_3^-$ -N,  $\text{PO}_4^{3-}$ -P and Chla concentration were filtered using Whatman GF/F filters. Chla was determined spectrophotometrically after extraction in 90% hot ethanol. TP, TN,  $\text{NO}_3^-$ -N,  $\text{NO}_2^{2-}$ -N,  $\text{NH}_4^+$ -N,  $\text{PO}_4^{3-}$ -P and  $\text{COD}_{\text{Mn}}$  were measured in the laboratory based on standard methods (Rice et al., 2012). The measurements and analyses of these environmental parameters were completed by the Nanjing Institute of Geography and Lakes, Chinese Academy of Sciences. The data are displayed in Table 2.

The relationship between our *COI* haplotypes and environmental data was tested using distance-based multivariate linear model analysis (DistLM) in the statistical software Primer-e PERMANOVA+ (Anderson et al., 2008). The relationships between *COI* haplotype frequencies and environmental parameters were initially examined by analysing each predictor separately (marginal tests), and then sequentially using an adjusted  $R^2$  selection procedure while avoiding collinearity between environmental data. Matrices in DistLM analyses were built using Bray-Curtis resemblance matrices of haplotypic abundances and Euclidean distance resemblance matrices of normalized 18 environmental parameters. The  $p$ -values for individual predictor variables were obtained using 9999 permutations (Leduc et al., 2012). Distance-based redundancy analysis (dbRDA) plots were also generated using PERMANOVA+ to visualize the results once the best DistLM model of each population was obtained. Specimens recovered from unmentioned lakes were excluded from this analysis as no corresponding environmental data were available for the locations.

### 3 | RESULTS

A total of 336 specimens were collected from seven lakes, as well as the Yangtze River mouth. All specimens were successfully amplified and sequenced to obtain *COI* barcodes for each individual (Table 1). After combined with sequences extracted from databases and previous studies presented in the literature (Table 1), the final alignment comprised 558 *COI* sequences of 526bp including samples from China and 222 mitochondrial DNA samples documented by other studies from *Corbicula* sampled in other regions of the world. The 526bp *COI* sequences comprised only 77 haplotypes.

### 3.1 | Haplotype phylogenetics

The *COI* phylogenetic tree obtained through ML delineated the monophyly of three major clades with relatively high support values: clades I, II and III (Figure 1). There was also additional *Corbicula spp.* that clustered outside these three major clades based on the examined sequences, such as *C. possoensis*, *C. loehensis*, *C. matannensis* and *C. moltkiana* from Malaysia and *C. sandai* from Japan (Figure 1). The *COI* haplotype FW17 retrieved in invasive lineage C/S also clustered outside the three clades (Peñarrubia et al., 2017; Pigneur, Etoundi, et al., 2014) and was not identified in our sampling in China, neither in other Chinese populations retrieved from other studies. Clade III occupied a basal position relative to clades I and II and clustered together 35 *COI* haplotypes from our study with the brackish water species *C. japonica* from Japan (Figure 1). Clade I included haplotype FW5 of invasive lineage A/R (corresponding to Hap 6 in our study) that is widely distributed in Europe and America (Peñarrubia et al., 2017; Pigneur, Etoundi, et al., 2014) and five additional *COI* haplotypes from China (Figure 1). FW5 (Hap6) was the only haplotype of clade I detected in our sampled lakes along the middle and lower Yangtze River (Figure 2). Clade II included not only 36 *COI* haplotypes from China but also FW1 of invasive *Corbicula* lineage B and FW4 of invasive lineage Rlc.

### 3.2 | Population diversity in middle and lower Yangtze River Basin

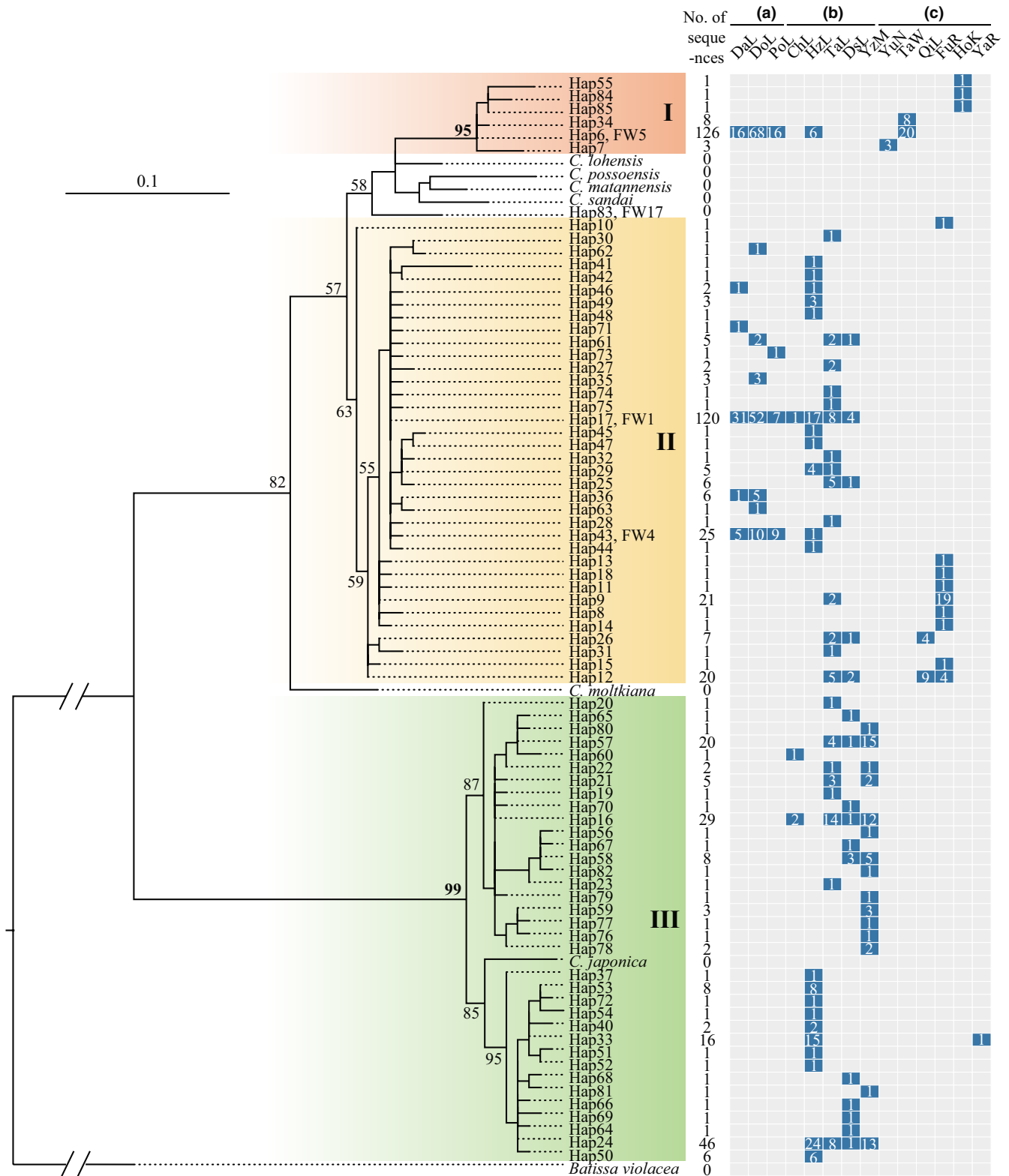
Among all 77 *COI* haplotypes we retrieved, the most common haplotype in our Chinese studied populations was *COI* FW5 (Hap6) of invasive lineage A/R (frequency: 126/558, Figure 1) and *COI* FW1 (Hap17) of invasive lineage B (frequency: 120/558, Figure 1). The most common *COI* haplotype in the estuarine clade III was Hap24 (frequency: 46/558, Figure 1).

Focusing on populations from Yangtze River Basin, *COI* haplotypes from the present study belonging to clade I were only observed in the middle reaches of the Yangtze River Basin and a few specimens in Hongze Lake (HzL), including only FW5 (Figure 2, a and b). Haplotypes from clade III were restricted to the lower reaches of the Yangtze River Basin and the estuary, comprising the estuarine lineages (Figure 2, e and f). In contrast, haplotypes from clade II were found across the whole middle and lower Yangtze River Basin, except in the Yangtze River mouth (YzM) (Figure 2, c and d) and included invasive *Corbicula* lineages B and Rlc, corresponding here to haplotypes Hap 17 (FW1) and Hap 43 (FW4) respectively. *COI* haplotypes FW1 and FW4 exhibited only one nucleotide difference and were both distributed widely in the middle and lower reaches of the Yangtze River (Figure 2, c and d). Haplotypes from clade II were found in sympatry with individuals from the other two clades (I and III). No lakes in the Yangtze River Basin were found to be inhabited by clams with *COI* haplotypes belonging to all three clades, except HzL, which does not belong to the basin but is connected by the Beijing-Hangzhou Grand Canal.



TABLE 2 Environmental parameters measured in each lake: The average value  $\pm$  standard deviation of all measurements taken during several sampling months is presented

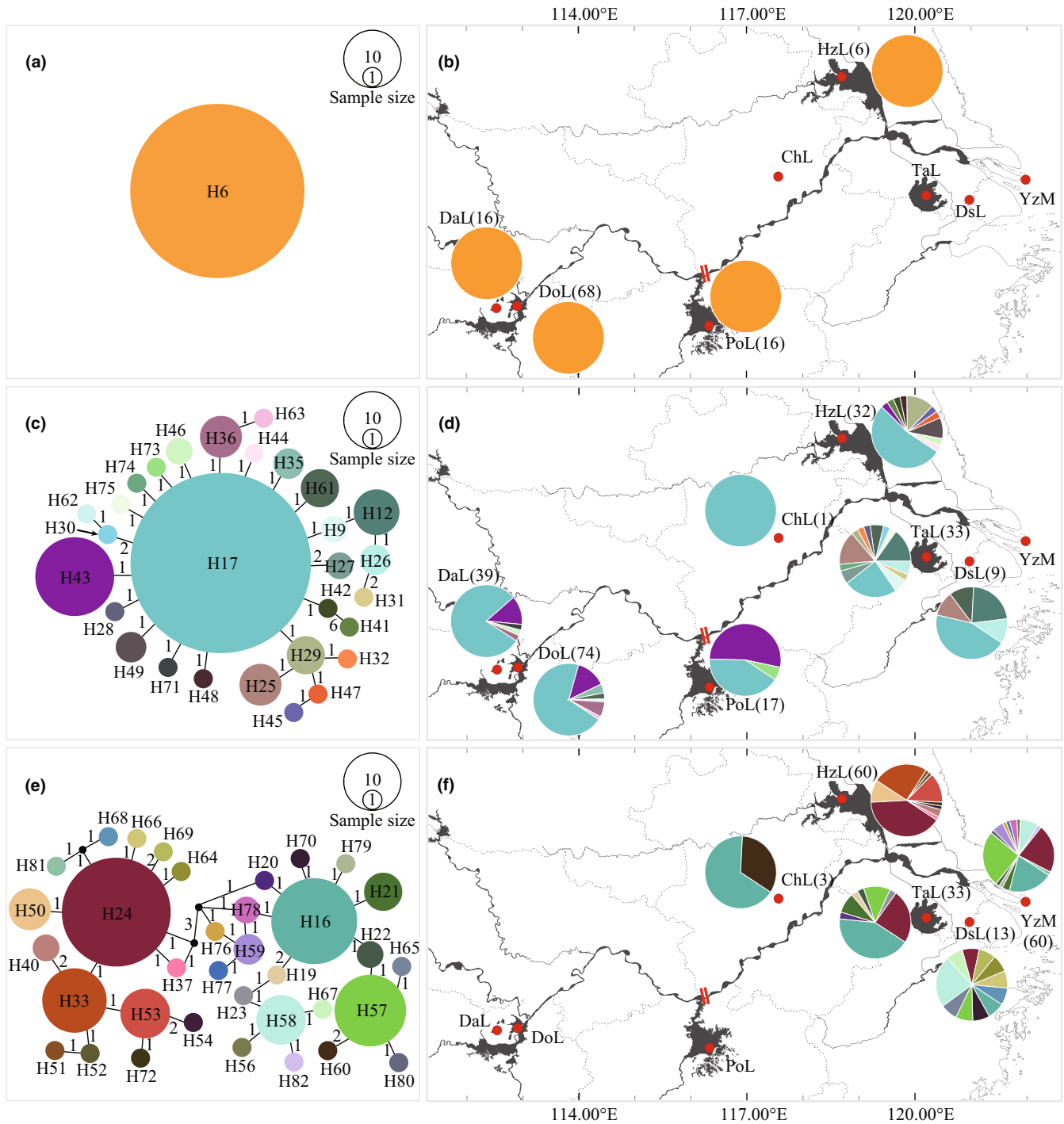
Environmental Parameter	DaL	DoL	Pol	ChL	HZL	TaL	DsL
Temperature/Temp (°C)	27.6 $\pm$ 0.16	28.3 $\pm$ 0.36	25.1 $\pm$ 0.80	29.9 $\pm$ 0.14	28.2 $\pm$ 0.28	28.4 $\pm$ 0.63	26.4 $\pm$ 0.21
Transparency/Trans (m)	0.31 $\pm$ 0.05	0.46 $\pm$ 0.16	0.41 $\pm$ 0.09	0.29 $\pm$ 0.07	0.21 $\pm$ 0.04	0.38 $\pm$ 0.10	0.38 $\pm$ 0.03
pH	8.2 $\pm$ 0.48	8.1 $\pm$ 0.08	7.4 $\pm$ 0.13	9.3 $\pm$ 0.30	9.1 $\pm$ 0.09	8.5 $\pm$ 0.08	8.6 $\pm$ 0.08
Dissolved oxygen/DO (mg/L)	7.6 $\pm$ 1.58	7.5 $\pm$ 0.10	7.3 $\pm$ 0.41	7.4 $\pm$ 0.40	6.7 $\pm$ 0.57	7.3 $\pm$ 0.43	7.1 $\pm$ 0.12
Conductivity/Cond ( $\mu$ s/cm)	319.13 $\pm$ 12.76	228.14 $\pm$ 45.95	97.03 $\pm$ 16.4	258.08 $\pm$ 35.11	479.75 $\pm$ 73.36	476.43 $\pm$ 102.86	672.00 $\pm$ 27.27
Chlorophyll-a/Chla ( $\mu$ g/L)	9.18 $\pm$ 8.45	1.70 $\pm$ 0.77	3.42 $\pm$ 0.66	4.98 $\pm$ 0.91	5.33 $\pm$ 3.06	21.75 $\pm$ 13.25	3.22 $\pm$ 0.69
Turbidity/Turb (mg/L)	29.50 $\pm$ 5.79	40.30 $\pm$ 22.25	39.87 $\pm$ 22.59	46.40 $\pm$ 14.54	93.79 $\pm$ 38.42	31.92 $\pm$ 12.16	23.56 $\pm$ 2.90
Chloride/Cl (mg/L)	14.80 $\pm$ 0.22	5.31 $\pm$ 2.11	4.58 $\pm$ 1.17	21.96 $\pm$ 4.07	42.91 $\pm$ 8.83	43.79 $\pm$ 15.49	63.68 $\pm$ 16.22
Calcium/Ca (mg/L)	29.92 $\pm$ 3.14	27.49 $\pm$ 4.93	10.10 $\pm$ 1.99	21.58 $\pm$ 2.44	29.11 $\pm$ 2.41	28.26 $\pm$ 2.01	43.45 $\pm$ 3.91
Magnesium/Mg (mg/L)	13.20 $\pm$ 0.22	6.79 $\pm$ 1.30	1.89 $\pm$ 0.25	5.84 $\pm$ 0.63	12.98 $\pm$ 2.05	7.52 $\pm$ 1.34	11.58 $\pm$ 0.60
Potassium/K (mg/L)	5.81 $\pm$ 0.17	2.37 $\pm$ 1.77	1.95 $\pm$ 0.30	3.63 $\pm$ 0.50	4.69 $\pm$ 0.82	5.18 $\pm$ 0.65	9.02 $\pm$ 0.17
Sodium/Na (mg/L)	9.38 $\pm$ 0.42	5.60 $\pm$ 2.06	4.33 $\pm$ 0.58	17.99 $\pm$ 3.42	49.72 $\pm$ 17.79	39.40 $\pm$ 13.60	82.94 $\pm$ 7.86
Total phosphorus/TP (mg/L)	0.06 $\pm$ 0.004	0.03 $\pm$ 0.004	0.07 $\pm$ 0.02	0.12 $\pm$ 0.02	0.09 $\pm$ 0.02	0.08 $\pm$ 0.03	0.21 $\pm$ 0.04
Total nitrogen/TN (mg/L)	0.50 $\pm$ 0.06	0.65 $\pm$ 0.04	1.36 $\pm$ 0.19	1.58 $\pm$ 0.28	1.86 $\pm$ 0.49	2.28 $\pm$ 0.87	2.23 $\pm$ 0.30
Nitrate/NO <sub>3</sub> <sup>-</sup> -N(mg/L)	0.25 $\pm$ 0.19	1.70 $\pm$ 0.82	0.87 $\pm$ 0.15	0.13 $\pm$ 0.17	1.23 $\pm$ 0.61	0.63 $\pm$ 0.26	0.64 $\pm$ 0.18
Nitrite/NO <sub>2</sub> <sup>2-</sup> -N (mg/L)	0.04 $\pm$ 0.04	0.01 $\pm$ 0.01	0.03 $\pm$ 0.01	0.01 $\pm$ 0.02	0.01 $\pm$ 0.01	0.05 $\pm$ 0.07	0.15 $\pm$ 0.07
Ammonium/NH <sub>4</sub> <sup>+</sup> -N (mg/L)	0.42 $\pm$ 0.19	0.99 $\pm$ 1.08	0.21 $\pm$ 0.06	0.16 $\pm$ 0.03	0.11 $\pm$ 0.05	0.54 $\pm$ 0.20	0.42 $\pm$ 0.29
Orthophosphate/ PO <sub>4</sub> <sup>3-</sup> -P ( $\mu$ g/L)	129.35 $\pm$ 10.98	43.61 $\pm$ 22.07	14.92 $\pm$ 5.25	4.64 $\pm$ 0.88	28.76 $\pm$ 13.53	8.79 $\pm$ 3.97	157.50 $\pm$ 40.43
Permanganate index/ COD <sub>Mn</sub> (mg/L)	6.04 $\pm$ 0.41	2.93 $\pm$ 0.38	3.73 $\pm$ 1.55	6.20 $\pm$ 2.47	4.13 $\pm$ 0.88	3.87 $\pm$ 0.74	4.17 $\pm$ 0.18



**FIGURE 1** Phylogenetic tree inferred from ML analyses and haplotype geographic distribution of COI haplotypes for *Corbicula* lineages. Bootstrap values (%) are indicated at each node. Each haplotype shared by different populations and its number of sequences displayed in the right panel; a and b being, respectively, middle and lower reaches of Yangtze River, and c representing populations outside of Yangtze River that we did not sample (QIL without specific location).

Among different *Corbicula* populations, Dianshan Lake (DsL) displayed the highest COI haplotype (0.957) and nucleotide diversity (0.0464) in the Yangtze River Basin (Table 3). The lowest COI

haplotype diversity (0.599) and the lowest nucleotide diversity (0.0080) were observed, respectively, in the Datong Lake (DaL) and the Yangtze River mouth (YzM), located at both extremities



**FIGURE 2** COI haplotype distributions of clades I (a and b), II (c and d) and III (e and f) in each *Corbicula* population along the middle and lower Yangtze River with populations outside of Yangtze River excluded. (a), (c) and (e) were haplotype networks per clade, (b), (d) and (f) were maps of part of the Yangtze River with the populations/sampling sites pointed with the haplotype distribution and abundance. In (a), (c) and (e), “H,” standing for “Hap,” with numbers corresponding to haplotype names in Figure 1; circle size being proportional to the number of sequences per haplotype, black dots and numbers next to solid lines representing undetected or hypothetical haplotypes and mutational steps between haplotypes respectively. In (b), (d) and (f), numbers in parentheses represent the number of sequences. The double solid red lines represent Hukou, which is the boundary for middle and lower reaches of Yangtze River.

of our sampling region. Both the haplotype and nucleotide diversity in the lakes along the Yangtze River increased with longitude, except for the river mouth population (Table 3), reflecting an increase in COI genetic diversity in the downstream direction of the

river course (i.e. flowing from west to east) (Figure 2). The AMOVA tests showed significant differentiation between the middle and lower reaches of the Yangtze River and among populations using the COI data ( $p < .001$ , Tables 4a and S2a). For *Corbicula* in the



Population	N	h	Hd	$\pi$	Longitude
DaL	55	6	0.599 ± 0.052	0.0117 ± 0.0013	112.54° E
DoL	142	8	0.634 ± 0.025	0.0139 ± 0.0002	112.93° E
PoL	33	4	0.665 ± 0.050	0.0146 ± 0.0006	116.27° E
ChL	4	3	0.833 ± 0.222	0.0440 ± 0.0210	117.57° E
HZL	98	22	0.877 ± 0.018	0.0423 ± 0.0022	118.72° E
TaL	66	22	0.916 ± 0.018	0.0441 ± 0.0009	120.16° E
DsL	22	16	0.957 ± 0.029	0.0464 ± 0.0033	120.97° E
YzM	60	15	0.851 ± 0.024	0.0080 ± 0.0005	121.96° E

Abbreviations: *h*, number of haplotypes; *hd*, haplotype diversity; *N*, sample size;  $\pi$ , nucleotide diversity.

Source of variation	Df	Sum of squares	Components	Percentage
(a) All geographic populations				
Between region	1	2003.496	7.816	50.46***
Among populations	6	458.213	1.392	8.98***
Within populations	472	2964.699	6.281	40.55*
Total	479	5426.408	15.489	
(b) Geographic populations in clade II				
Between region	1	6.587	0.04693	7.69
Among populations	5	7.514	0.04096	6.71*
Within populations	198	103.431	0.52238	85.60***
Total	204	117.532	0.61027	
(c) Geographic populations in clade III				
Among populations	4	135.321	1.07106	34.95***
Within populations	164	326.880	1.99317	65.05
Total	168	462.201	3.06423	

Note: Populations of the AMOVA analysis: DaL, DoL, PoL, ChL, HzL, TaL, DsL and YzM and the regions included in the analysis: middle reaches and lower reaches of Yangtze River. Only one haplotype (Hap6) was detected in the collected geographic populations in clade I, and the AMOVA test was therefore not applicable. Samples in clade III were only found in the lower region of the Yangtze River, and the hierarchical level of the region was not applicable. \* $p < 0.05$ ; \*\*\* $p < 0.001$ .

Yangtze Basin, all samples in clade I belonged to the same *COI* Hap 6, displaying no differentiation. In clade II, including haplotypes from almost the entire middle and lower basin regions, 85.60% of the genetic *COI* differentiation came from within populations ( $p < .001$ , Table 4b). In clade III, restricted to the lower basin region, a significant differentiation was observed among populations in the region ( $p < 0.001$ , Table 4c).

### 3.3 | Environmental factors shaping the haplotype's spatial distribution

Following the removal of the 12 variables that did not explain the distribution (Temp, Trans, pH, DO, Ca, K, Na, TN,  $\text{NO}_3^-$ -N,  $\text{NO}_2^-$ -N,  $\text{NH}_4^+$ -N and  $\text{COD}_{\text{Mn}}$ ), marginal tests resulting from DistLM analysis of *COI* population genetic variation at the regional scale showed that the 98.72% of the spatial *COI* haplotype distribution variation could be explained by chloride (Cl, 50.93%), turbidity (Turb, 23.00%), total phosphorus (TP, 17.37%) and magnesium (Mg, 7.42%) (Figure 3).

Chloride, whose concentrations were shown to increase with sodium and conductivity, was positively correlated with the *COI* distribution of *Corbicula* from Tai Lake (TaL) and Dianshan Lake (DsL) and negatively with Datong Lake (DaL), Dongting Lake (DoL) and Poyang Lake (PoL). The composition of *COI* haplotypes in Hongze Lake (HzL) was possibly impacted by the high turbidity. Accordingly, the *COI* variation observed between geographic populations and the significant *p*-value of chloride suggested that salinity likely affected *Corbicula*'s distribution and dispersal along the river.

## 4 | DISCUSSION

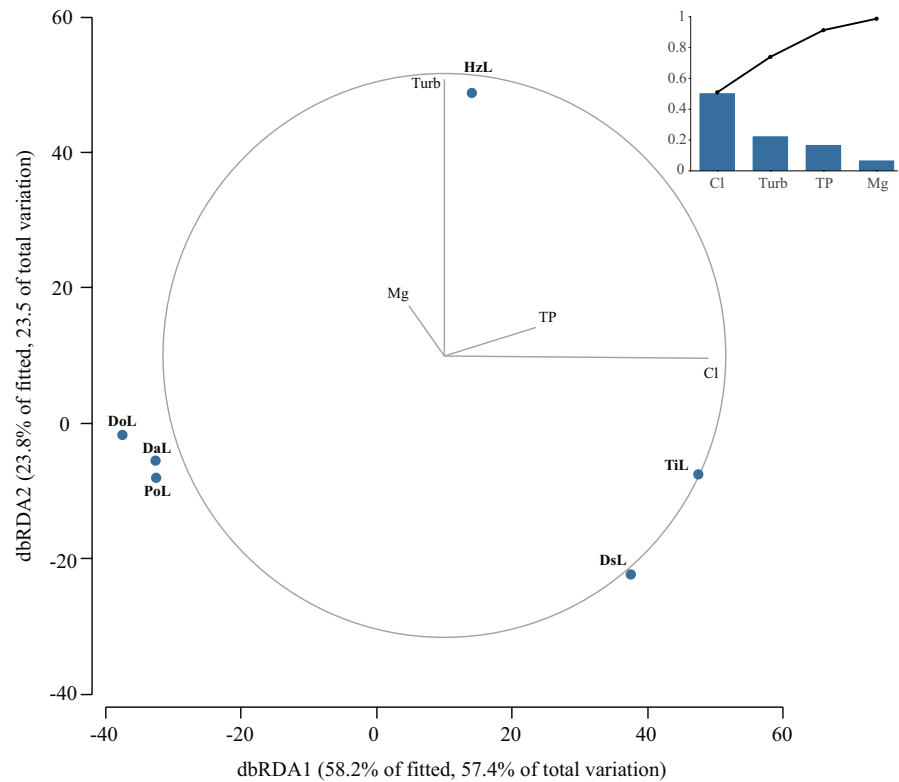
### 4.1 | Three *COI* corbicula clades detected in China

Based on the phylogenetic tree and haplotype networks constructed using the mitochondrial *COI* marker, only three major *Corbicula* clades were grouped within the Yangtze River Basin (Figures 1 and 2). Previous studies documented seven freshwater *Corbicula* species

TABLE 3 Genetic diversity of *Corbicula* among different geographic populations

TABLE 4 AMOVA analysis of different geographic populations in the Yangtze Basin

**FIGURE 3** dbRDA analysis plot showing the relationship between population distribution and environmental variables. Environmental variables are indicated by the grey line pointing in the direction of increasing values, including chloride (Cl), turbidity (Turb), total phosphorus (TP) and magnesium (Mg). Populations are indicated by blue dots. Blue bar (top-right) represents proportion of explained variation by each environmental variable, and line chart represents cumulative proportion.



in China based on morphological characteristics: *C. fluminea*, *C. nitens*, *C. aurea*, *C. largillierti*, *C. fluminalis*, *C. tenuis* and *C. scholastica* (Wang et al., 2014). *C. fluminea* was described with the broadest distribution, inhabiting most river basins and lakes in China (Shu et al., 2014). Similarly, *C. nitens*, *C. aurea* and *C. largillierti* had a wider distribution than *C. fluminalis* which had only been reported within the Yangtze River Basin (Lin, 1962; Tchang et al., 1965). Specimens of *C. tenuis* and *C. scholastica* had also been detected within the Yuan River and the Huai River respectively (Huang & Li, 2003). High polymorphism and phenotypic plasticity within *Corbicula* have resulted in the delimitation of distinct morphotypes that appear highly similar genetically, making identification of distinct *Corbicula* species problematic and often resulting in multiple synonyms for a single lineage (Lin, 1962; Tchang et al., 1965). Similarly, specimens sampled from Dongting Lake, Poyang Lake and Qingshan Lake were previously grouped into two separate subclades and were regarded as *C. fluminea* and *C. leana* (Park & Kim, 2003). However, the macrozoobenthos surveys around these areas only found *C. fluminea* based on morphological examination (Shu et al., 2014).

Although only three major *COI* *Corbicula* clades were detected by the present study of Chinese lakes and Yangtze River mouth, several distinct *COI* haplotypes were retrieved within clades II and III with 77 distinct *COI* haplotypes in total. Since only the *COI* marker was used here, we did not study the nuclear genetic diversity and we could not detect potential cytonuclear mismatches or hybrids occurring in *Corbicula* when androgenetic lineages occur in sympatry. The biological species concept, which defines species as units that exhibit barriers to reproduction, is widely used to delimit taxonomical units. The different reproductive strategies found in *Corbicula*

(androgenetic hermaphrodite, sexual dioecism, cross-fertilization and self-fertilization (Gomes et al., 2016; Peñarrubia et al., 2017; Pigneur, Etoundi, et al., 2014)), as well as possible hybridization between distinct lineages (Hedtke et al., 2008; Lee et al., 2005; Pigneur, Etoundi, et al., 2014; Vastrade et al., 2022), make species delimitation in general using the biological species concept problematic in this genus. Genetic lineages defined using the *COI* marker, as in the present study, can identify distinct mitochondrial lineages within the genus *Corbicula* as evidenced by other studies (e.g. Park & Kim, 2003; Peñarrubia et al., 2017; Pigneur, Etoundi, et al., 2014), but species cannot be identified as discussed in Hedtke et al. (2011) or Vastrade et al. (2022) who included nuclear markers and identified discordances due to cross-species hybridizations.

## 4.2 | Genetic spatial distribution and diversity

The haplotype and nucleotide diversity of sampled *Corbicula* clams residing within the Yangtze River Basin in the present study ranged between 0.599 and 0.957 and 0.0080 and 0.0464, respectively (Table 3), being higher than in the invasive populations in Europe and America where only four distinct *COI* *Corbicula* haplotypes were detected: form A/R (FW5), form B (FW1), form Rlc (FW4) and form C/S (FW17) (Gomes et al., 2016; Peñarrubia et al., 2017; Pigneur, Etoundi, et al., 2014), and recently form D (Haponski & Ó Foighil, 2019). The highest *COI* diversity in our study was detected in Dianshan Lake (DsL) located at the lower reaches of the Yangtze River: 16 distinct haplotypes belonging to clades II (5 haplotypes) and III (11 haplotypes) were found in this lake. Hongze Lake (HzL),

outside but connected with the Yangtze River Basin, also displayed a high diversity with haplotypes of the three clades found in this lake. A similar pattern of *COI* genetic diversity was found in the *Corbicula* populations of Ebro River (Iberian Peninsula, Europe), with higher haplotype and nucleotide diversity in estuarine populations than inland populations, showing an increased diversity downstream (Peñarrubia et al., 2017). The *COI* haplotype and nucleotide diversity of Dongting Lake (DoL) in the present study were similar to those evaluated using the *COI* marker by Wang, Li, et al. (2018) but higher than those assessed by the *Cytb* marker (Wang, Zhu, et al., 2018), which may be attributed to differences in molecular markers or sampling. The *COI* genetic diversity of HzL was also similar to previously reported data (Li et al., 2015).

The low genetic divergence of *Corbicula* clams within their invaded range when compared to the native regions likely results from either a founder effect, whereby only a limited number of specimens were introduced from their native range, or through a short-lived bottleneck event followed by a fast population expansion of a few lineages through androgenesis (Pigneur, Etoundi, et al., 2014). The monomorphism found here in clade I (only haplotype 6 = FW5) represents the clonal or androgenetic invasive lineage A/R, which was widespread in the invasive range but also in the native region and within the Yangtze River Basin. Polymorphism in clade II was represented by 36 *COI* haplotypes found in the Yangtze Basin, including the invading lineages B (haplotype 17 or FW1) and Rlc (haplotype 43 or FW4). Invasive lineage B, as abundant as lineage A/R in Chinese lakes, was found in all the lakes sampled in this study except Yangtze River Mouth, while invasive lineage Rlc was sampled in four distinct lakes (Figure 1). A high diversity was also detected in clade III, with 35 distinct *COI* haplotypes retrieved in the Yangtze River and clustering with the brackish sexual species *C. japonica* that was only found in Japan and Korea (Okamoto & Arimoto, 1986; Park et al., 2002). The relatively high *COI* genetic diversity observed in clades II and III could be linked to possible sexual reproduction or frequent origins of asexual *Corbicula* lineages from sexually reproducing populations (Pigneur, Etoundi, et al., 2014). Biflagellate sperm is a diagnostic marker of androgenesis in *Corbicula* (Pigneur et al., 2012), and previous reports detected it in *Corbicula* populations in the middle and upper Yangtze River (Qiu et al., 2001; Wang, Li, et al., 2018), but not in population in the Yangtze River mouth (Zhan, 2020). This spatial variation of reproduction may provide indirect evidence to explain genetic diversity differences.

### 4.3 | Radiation of *Corbicula* from marine into freshwater habitats

Previous studies on the phylogeny of the family Cyrenidae revealed invasions of freshwater environments by marine/brackish *Corbicula* lineages (Glaubrecht et al., 2006; Graf, 2013). Similar results were detected by the present study, supporting the hypothesis of *Corbicula* radiation from marine into freshwater habitats. First, clade III was mostly found in estuarine lakes such as Tai (TaL) and Dianshan (DsL)

lakes in the lower reaches of the Yangtze River Basin (where the chloride concentration was  $43.79 \pm 15.49$  mg/L and  $63.68 \pm 16.22$  mg/L, respectively), as well as the Yangtze River mouth. Both lakes have originated from the closely residing East China Sea over time. As a geologically recent waterbody, Tai Lake was a large embayment of the East China Sea as recent as 1 million years ago and gradually became separated, while Dianshan Lake remains connected with the East China Sea through the Huangpu River (Gu et al., 2019). Two haplotypes of clade III also invaded Chao Lake (ChL) where the chloride is much lower ( $21.96 \pm 4.07$  mg/L), while distinct haplotypes of clade III (10 haplotypes in total) invaded Hongze Lake (HzL) where the chloride reaches  $42.91 \pm 8.83$  mg/L. Second, marginal tests resulting from DistLM analysis of *COI* population genetic variation at the regional scale suggested that environmental parameters largely explained the spatial *COI* haplotype distribution, with chloride levels explaining most variations. We, however, did not study the nuclear genetic diversity of *Corbicula* and therefore the impact of chloride on the spatial genetic variation within the Yangtze River Basin should be considered preliminary. At the global scale, the temperature has been suggested as the critical factor shaping *Corbicula* distribution (Crespo et al., 2015; Gama et al., 2016). However, at local scales, salinity is considered a major abiotic factor influencing the success and velocity of invasion into new environments in *Corbicula* as suggested also by other studies (Crespo et al., 2017; Sousa et al., 2006). Notably, salinity has been observed to directly affect the physiological processes of *Corbicula* (Baba et al., 1999; Crespo et al., 2017). Third, the two invasive lineages A/R and B, abundant in the Chinese lakes studied here, were sampled from freshwater and estuarine lakes (with chloride ranging from  $4.58 \pm 1.17$  to  $42.91 \pm 8.83$  mg/L for lineage A/R and  $4.58 \pm 1.17$  to  $63.68 \pm 16.22$  mg/L for lineage B). Invasive lineage Rlc was found in lakes where chloride ranged between 5.31 mg/L and 42.91 mg/L. The three invasive lineages, therefore, seem to show a wide tolerance towards varying salinity levels, explaining their abundance and widespread distribution within the Yangtze River Basin (Figure 2; Table 2) and also abroad.

Environmental factors other than salinity may have shaped the colonization of clams during dispersal. High turbidity has the potential for decreased feeding, reduced reproduction and even to reduce shellfish survival (Avelar et al., 2014; Tuttle-Raycraft & Ackerman, 2020; Wilber & Clarke, 2001). Besides, as filter-feeding bivalves, the survival of *Corbicula* depends heavily on the abundance of some phytoplankton species (Sousa et al., 2014). The availability of phosphorus is an important factor that directly controls the phytoplankton growth, biomass and species composition (reviewed in Xu et al., 2010), and therefore may indirectly affect *Corbicula* populations. Finally, Mg, being essential for the shell formation of *Corbicula* (Zhao et al., 2017), could also impact the distribution of *Corbicula* clams. Although these factors may only have a low contribution to the spatial distribution of clams when compared to salinity, their impact should be considered in further analyses.

Environmental factors may vary with regional changes, but salinity usually varies with distance from the estuary, both in invaded and native areas. Salinity was observed here to have the highest impact

on the success of *Corbicula* radiation and may support genetic barriers that shape an uneven distribution through the limitation of gene flow among different *Corbicula* lineages, depending on their salinity tolerance (e.g. Crespo et al., 2017). The present study, however, emphasizes that invasive *Corbicula* lineages found in Europe and America, are also abundant in the native range, combining an androgenetic mode of reproduction with a high salinity tolerance, corroborating their high invasion capacity.

#### 4.4 | Implications for freshwater biodiversity conservation

Control and management of invasive species are highlighted as a priority for biodiversity conservation (Coughlan et al., 2020; Guareschi et al., 2021). *Corbicula* lineages A/R and B are abundant and widely distributed in native and invaded regions (Gomes et al., 2016; Park & Kim, 2003; Peñarrubia et al., 2017; Pigneur, Etoundi, et al., 2014), and their peculiar reproductive mode (including androgenesis, self- and cross-fertilization) may be one of the main reasons for their invasive capacities in freshwater ecosystems (Gomes et al., 2016; Kraemer et al., 1986; Lee et al., 2005; Vastrade et al., 2022). Importantly, our findings here revealed that these two lineages had a wide tolerance to salinity being able to colonize from estuary to inland rivers and lakes, increasing their invasion potential in different water systems. Conservation managers should pay close attention to populations inhabiting environments with a high or fluctuating salinity, as these populations may cryptically radiate upstream into freshwaters overtime. However, whether those further upstream can disperse in a downstream direction and show tolerance to increased salinity levels still requires further investigation to confirm. Distribution and dispersal modelling for these invasive lineages should be updated to reflect this improved understanding of *Corbicula* tolerance to salinity, as study findings based on field data corroborate and expand upon what has so far only been observed under laboratory conditions. Therefore, the two invasive lineages require special caution to control and manage, as these lineages have the potential to establish a new population through only one individual (reviewed in Pigneur et al., 2012), cause biomass and production reduction in primary producers but also habitat competition with native species in the invaded ecosystems (Pigneur, Falisse, et al., 2014; Sousa et al., 2014; Strayer, 2010). Given that these two lineages have a high capacity for invasion, it is critical to detect them early during their invasion so that the loss of native species and the cost of management could be reduced. Although detecting *Corbicula* at an early stage is difficult due to few individuals and small larvae, techniques such as environmental DNA provide a viable method to achieve rapid and sensitive detection (Simberloff et al., 2013). Design of suitable detection primers and kits in the future would directly benefit the biodiversity conservation in invasive areas, as well as the implementation of improved biosecurity protocols (Coughlan et al., 2020).

#### ACKNOWLEDGEMENT

We gratefully thank Guiping Wang and Jianping Wang for collecting specimens. This study was supported by National Natural Science Foundation of China (31772832), National Key Research and Development Program of China (No. 2019YFD0900603), Hunan Provincial Modern Agricultural Research System (No. 2019-105) and Youth Innovation Promotion Association CAS (2020316). NEC graciously acknowledges support from the Irish Research Council (GOIPD/2022/861).

#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

#### DATA AVAILABILITY STATEMENT

Newly generated COI haplotype data in this study were deposited in NCBI under the accession numbers ON504951-95, see the Supporting Information Table S3 for details. Additional files could be found in the Supporting Information and accessed on a figshare deposit: <https://doi.org/10.6084/m9.figshare.21154609>.

#### ORCID

Cong Zeng  <https://orcid.org/0000-0003-1003-9382>

Yangxin Tang  <https://orcid.org/0000-0003-3015-2729>

#### REFERENCES

- Anderson, M., Gorley, R., & Clarke, K. (2008). *PERMANOVA+ for PRIMER: Guide to software and statistical methods*. PRIMER-e.
- Araujo, R., Moreno, D., & Ramos, M. (1993). The Asiatic clam *Corbicula fluminea* (Müller, 1774) (Bivalvia: Corbiculidae) in Europe. *American Malacological Bulletin*, 26, 39–49. <https://doi.org/10.1016/j.actao.2004.05.005>
- Avelar, W., Neves, F., & Lavrador, M. (2014). Modelling the risk of mortality of *Corbicula fluminea* (Müller, 1774) (Bivalvia: Corbiculidae) exposed to different turbidity conditions. *Brazilian Journal of Biology*, 74(2), 509–514. <https://doi.org/10.1590/1519-6984.21612>
- Baba, K., Tada, M., Kawajiri, T., & Kuwahara, Y. (1999). Effects of temperature and salinity on spawning of the brackish water bivalve *Corbicula japonica* in Lake Abashiri, Hokkaido, Japan. *Marine Ecology Progress Series*, 180, 213–221. <https://doi.org/10.3354/meps180213>
- Bespalaya, Y., Bolotov, I., Aksanova, O., Kondakov, A., Gofarov, M., Laenko, T., Sokolova, S. E., Shevchenko, A. R., & Travina, O. V. (2018). Aliens are moving to the Arctic frontiers: An integrative approach reveals selective expansion of androgenic hybrid *Corbicula* lineages towards the north of Russia. *Biological Invasions*, 20, 2227–2243. <https://doi.org/10.1007/s10530-018-1698-z>
- Coughlan, N. E., Cuthbert, R. N., & Dick, J. T. A. (2020). Aquatic biosecurity remains a damp squib. *Biodiversity and Conservation*, 29, 3091–3309. <https://doi.org/10.1007/s10531-020-02011-8>
- Coughlan, N. E., Stevens, A. L., Kelly, T. C., Dick, J. T., & Jansen, M. A. (2017). Zoochorous dispersal of freshwater bivalves: An overlooked vector in biological invasions? *Knowledge & Management of Aquatic Ecosystems*, 418, 42. <https://doi.org/10.1051/kmae/201703>
- Courchamp, F., Fournier, A., Bellard, C., Bertelsmeier, C., Bonnaud, E., Jeschke, J. M., & Russell, J. C. (2017). Invasion biology: Specific problems and possible solutions. *Trends in Ecology & Evolution*, 32(1), 13–22. <https://doi.org/10.1016/j.tree.2016.11.001>



- Crespo, D., Dolbeth, M., Leston, S., Sousa, R., & Pardal, M. Â. (2015). Distribution of *Corbicula fluminea* (Müller, 1774) in the invaded range: A geographic approach with notes on species traits variability. *Biological Invasions*, 17(7), 2087–2101.
- Crespo, D., Leston, S., Martinho, F., Pardal, M. A., & Dolbeth, M. (2017). Survival of *Corbicula fluminea* (Müller, 1774) in a natural salinity and temperature gradient: A field experiment in a temperate estuary. *Hydrobiologia*, 784(1), 337–347. <https://doi.org/10.1007/s10530-015-0862-y>
- Diagne, C., Leroy, B., Vaissière, A.-C., Gozlan, R. E., Roiz, D., Jarić, I., Salles, J. M., Bradshaw, C. J. A., & Courchamp, F. (2021). High and rising economic costs of biological invasions worldwide. *Nature*, 592(7855), 571–576. <https://doi.org/10.1038/s41586-021-03405-6>
- Excoffier, L., & Lischer, H. E. (2010). Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and windows. *Molecular Ecology Resources*, 10(3), 564–567. <https://doi.org/10.1111/j.1755-0998.2010.02847.x>
- Ferreira-Rodríguez, N., Fernández, I., Varandas, S., Cortes, R., Cancela, M. L., & Pardo, I. (2017). The role of calcium concentration in the invasive capacity of *Corbicula fluminea* in crystalline basins. *Science of the Total Environment*, 580, 1363–1370. <https://doi.org/10.1016/j.scitotenv.2016.12.100>
- Folmer, O., Black, M., Hoeh, W., Lutz, R., & Vrijenhoek, R. (1994). DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology*, 3(5), 294–299.
- Gama, M., Crespo, D., Dolbeth, M., & Anastácio, P. (2016). Predicting global habitat suitability for *Corbicula fluminea* using species distribution models: The importance of different environmental datasets. *Ecological Modelling*, 319, 163–169. <https://doi.org/10.1016/j.ecolmodel.2015.06.001>
- Glaubrecht, M., Fehér, Z., & Von Rintelen, T. (2006). Brooding in *Corbicula madagascariensis* (Bivalvia, Corbiculidae) and the repeated evolution of viviparity in corbiculids. *Zoologica Scripta*, 35(6), 641–654. <https://doi.org/10.1111/j.1463-6409.2006.00252.x>
- Glaubrecht, M., Von Rintelen, T., & Korniuschin, A. (2003). Toward a systematic revision of brooding freshwater Corbiculidae in Southeast Asia (Bivalvia, Veneroidea): On shell morphology, anatomy and molecular phylogenetics of endemic taxa from islands in Indonesia. *Malacologia*, 45(1), 1–40.
- Gomes, C., Sousa, R., Mendes, T., Borges, R., Vilares, P., Vasconcelos, V., Guilhermino, L., & Antunes, A. (2016). Low genetic diversity and high invasion success of *Corbicula fluminea* (Bivalvia, Corbiculidae) (Müller, 1774) in Portugal. *PLoS One*, 11(7), e0158108. <https://doi.org/10.1371/journal.pone.0158108>
- Graf, D. L. (2013). Patterns of freshwater bivalve global diversity and the state of phylogenetic studies on the Unionoidea, Sphaeriidae, and Cyrenidae. *American Malacological Bulletin*, 31(1), 135–153. <https://doi.org/10.4003/006.031.0106>
- Gu, X., Zeng, Q., Mao, Z., Chen, H., & Li, H. (2019). Water environment change over the period 2007–2016 and the strategy of fishery improve the water quality of Lake Taihu. *Journal of Lake Sciences*, 31(2), 305–318. <https://doi.org/10.18307/2019.0201>
- Guareschi, S., Laini, A., England, J., Barrett, J., & Wood, P. J. (2021). Multiple co-occurring alien invaders constrain aquatic biodiversity in rivers. *Ecological Applications*, 31(6), e02385. <https://doi.org/10.1002/eap.2385>
- Hall, T. (1999). BioEdit: A user-friendly biological sequence alignment editor and analysis program for windows 95/98/NT. *Nucleic Acids Symposium Series*, 41, 95–98. [https://doi.org/10.14601/Phytopathol\\_Mediterr-14998u1.29](https://doi.org/10.14601/Phytopathol_Mediterr-14998u1.29)
- Haponski, A. E., & Ó Foighil, D. (2019). Phylogenomic analyses confirm a novel invasive north American corbicula (Bivalvia: Cyrenidae) lineage. *PeerJ*, 7, e7484. <https://doi.org/10.7717/peerj.7484>
- Haubrock, P. J., Cuthbert, R. N., Ricciardi, A., Diagne, C., & Courchamp, F. (2022). Economic costs of invasive bivalves in freshwater ecosystems. *Diversity and Distributions*, 28(5), 1010–1021. <https://doi.org/10.1111/ddi.13501>
- Hedtke, S. M., Glaubrecht, M., & Hillis, D. M. (2011). Rare gene capture in predominantly androgenetic species. *Proceedings of the National Academy of Sciences*, 108(23), 9520–9524. <https://doi.org/10.1073/pnas.1106742108>
- Hedtke, S. M., Stanger-Hall, K., Baker, R. J., & Hillis, D. M. (2008). All-male asexuality: Origin and maintenance of androgenesis in the Asian clam corbicula. *Evolution*, 62(5), 1119–1136. <https://doi.org/10.1111/j.1558-5646.2008.00344.x>
- Huang, B., & Li, Y. (2003). Bivalves in the Zhengyangguan part of the Huai River, Shou County, Anhui Province. *Chinese Journal of Zoology*, 38(4), 80–85+130–131. <https://doi.org/10.13859/j.cjz.2003.04.018>
- Iida, M., Kanno, M., & Kijima, A. (2012). Genetic population structure of *Corbicula japonica* around East Asia estimated by mtDNA COI sequence analysis. *Nippon Suin Gakkaishi (Japanese Edition)*, 78(5), 934–944. <https://doi.org/10.2331/suisan.78.934>
- Kalyaanamoorthy, S., Minh, B. Q., Wong, T. K., Von Haeseler, A., & Jermin, L. S. (2017). ModelFinder: Fast model selection for accurate phylogenetic estimates. *Nature Methods*, 14(6), 587–589. <https://doi.org/10.1038/nmeth.4285>
- Komaru, A., Yamada, M., & Houki, S. (2013). Relationship between two androgenetic clam species, *Corbicula leana* and *Corbicula fluminea*, inferred from mitochondrial cytochrome b and nuclear 28S rRNA markers. *Zoological Science*, 30(5), 360–365. <https://doi.org/10.2108/zsj.30.360>
- Kraemer, L., Swanson, C., Galloway, M., & Kraemer, R. (1986). Biological basis of behaviour in *Corbicula fluminea*, II. Functional morphology of reproduction and development and review of evidence for self-fertilization. *American Malacological Bulletin, Special edition*, 2, 193–201.
- Leduc, D., Rowden, A. A., Bowden, D. A., Probert, P. K., Pilditch, C. A., & Nodder, S. D. (2012). Unimodal relationship between biomass and species richness of deep-sea nematodes: Implications for the link between productivity and diversity. *Marine Ecology Progress Series*, 454, 53–64. <https://doi.org/10.3354/meps09609>
- Lee, T., Siripatrawan, S., Ituarte, C. F., & Ó Foighil, D. (2005). Invasion of the clonal clams: *Corbicula* lineages in the New World. *American Malacological Bulletin*, 20(1/2), 113–122.
- Leigh, J. W., & Bryant, D. (2015). POPART: Full-feature software for haplotype network construction. *Methods in Ecology and Evolution*, 6(9), 1110–1116. <https://doi.org/10.1111/2041-210X.12410>
- Li, D. M., Zhang, T. Q., Tang, S. K., Zhong, L. Q., & Liu, X. W. (2015). Analysis of genetic diversity and population structure of *Corbicula fluminea* in Hongze Lake in mitochondrial COI gene sequence. *Oceanologia et Limnologia Sinica*, 46(6), 1339–1346. <https://doi.org/10.11693/hyhz20141200353>
- Lin, Z. (1962). Unionidae (Mollusca) of Poyang Lake, Kiangsi Province China. *Acta Zoologica Sinica*, 14(2), 249–260.
- McDowell, W. G., Benson, A. J., & Byers, J. E. (2014). Climate controls the distribution of a widespread invasive species: Implications for future range expansion. *Freshwater Biology*, 59(4), 847–857. <https://doi.org/10.1111/fwb.12308>
- McMahon, R. F. (1979). Response to temperature and hypoxia in the oxygen consumption of the introduced Asiatic freshwater clam *Corbicula fluminea* (Müller). *Comparative Biochemistry and Physiology Part A: Physiology*, 63(3), 383–388. [https://doi.org/10.1016/0300-9629\(79\)90607-8](https://doi.org/10.1016/0300-9629(79)90607-8)
- Müller, O., & Baur, B. (2011). Survival of the invasive clam *Corbicula fluminea* (Müller) in response to winter water temperature. *Malacologia*, 53(2), 367–371. <https://doi.org/10.4002/040.053.0207>



- Nguyen, L. T., Schmidt, H. A., Von Haeseler, A., & Minh, B. Q. (2015). IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution*, 32(1), 268–274. <https://doi.org/10.1093/molbev/msu300>
- Okamoto, A., & Arimoto, B. (1986). Chromosomes of *Corbicula japonica*, *C. sandai* and *C. (Corbiculina) leana* (Bivalvia: Corbiculidae). *Venus (Japanese Journal of Malacology)*, 45(3), 194–202. [https://doi.org/10.18941/venusjmm.45.3\\_194](https://doi.org/10.18941/venusjmm.45.3_194)
- Park, J. K., & Kim, W. (2003). Two corbicula (Corbiculidae: Bivalvia) mitochondrial lineages are widely distributed in Asian freshwater environment. *Molecular Phylogenetics and Evolution*, 29(3), 529–539. [https://doi.org/10.1016/S1055-7903\(03\)00138-6](https://doi.org/10.1016/S1055-7903(03)00138-6)
- Park, J. K., Lee, J. S., & Kim, W. (2002). A single mitochondrial lineage is shared by morphologically and allozymatically distinct freshwater corbicula clones. *Molecules and Cells*, 14(2), 318–322.
- Peñarrubia, L., Araguas, R. M., Vidal, O., Pla, C., Viñas, J., & Sanz, N. (2017). Genetic characterization of the Asian clam species complex (corbicula) invasion in the Iberian Peninsula. *Hydrobiologia*, 784(1), 349–365. <https://doi.org/10.1007/s10750-016-2888-2>
- Pfenninger, M., Reinhardt, F., & Streit, B. (2002). Evidence for cryptic hybridization between different evolutionary lineages of the invasive clam genus corbicula (Veneroidea, Bivalvia). *Journal of Evolutionary Biology*, 15(5), 818–829. <https://doi.org/10.1046/j.1420-9101.2002.00440.x>
- Pigneur, L. M., Etoundi, E., Aldridge, D. C., Marescaux, J., Yasuda, N., & Van Doninck, K. (2014). Genetic uniformity and long-distance clonal dispersal in the invasive androgenetic corbicula clams. *Molecular Ecology*, 23(20), 5102–5116. <https://doi.org/10.1111/mec.12912>
- Pigneur, L. M., Falisse, E., Roland, K., Everbecq, E., Deliège, J. F., Smits, J. S., Van Doninck, K., & Descy, J. P. (2014). Impact of invasive Asian clams, *corbicula* spp., on a large river ecosystem. *Freshwater Biology*, 59(3), 573–583. <https://doi.org/10.1111/fwb.12286>
- Pigneur, L. M., Hedtke, S., Etoundi, E., & Van Doninck, K. (2012). Androgenesis: A review through the study of the selfish shellfish *corbicula* spp. *Heredity*, 108(6), 581–591. <https://doi.org/10.1038/hdy.2012.3>
- Pigneur, L. M., Marescaux, J., Roland, K., Etoundi, E., Descy, J. P., & Van Doninck, K. (2011). Phylogeny and androgenesis in the invasive corbicula clams (Bivalvia, Corbiculidae) in Western Europe. *BMC Evolutionary Biology*, 11(1), 147. <https://doi.org/10.1186/1471-2148-11-147>
- Qiu, A. D., Shi, A. J., & Komaru, A. (2001). Yellow and brown shell colour morphs of *Corbicula fluminea* (Bivalvia: Corbiculidae) from Sichuan Province, China, are triploids and tetraploids. *Journal of Shellfish Research*, 20, 323–328.
- Renard, E., Bachmann, V., Cariou, M., & Moreteau, J. (2000). Morphological and molecular differentiation of invasive freshwater species of the genus corbicula (Bivalvia, Corbiculidae) suggest the presence of three taxa in French rivers. *Molecular Ecology*, 9(12), 2009–2016. <https://doi.org/10.1046/j.1365-294X.2000.01104.x>
- Rice, E. W., Baird, R. B., Eaton, A. D., & Clesceri, L. S. (2012). *Standard methods for the examination of water and wastewater*. American Public Health Association.
- Rozas, J., Ferrer-Mata, A., Sánchez-DelBarrio, J. C., Guirao-Rico, S., Librado, P., Ramos-Onsins, S. E., & Sánchez-Gracia, A. (2017). DnaSP 6: DNA sequence polymorphism analysis of large data sets. *Molecular Biology and Evolution*, 34(12), 3299–3302. <https://doi.org/10.1093/molbev/msx248>
- Shu, F. Y., Wang, H. J., Cui, Y. D., & Wang, H. Z. (2014). Diversity and distribution pattern of freshwater molluscs in the Yangtze River basin. *Acta Hydrobiologica Sinica*, 38(1), 19–26. <https://doi.org/10.7541/2014.03>
- Simberloff, D., Martin, J.-L., Genovesi, P., Maris, V., Wardle, D. A., Aronson, J., Courchamp, F., Galil, B., García-Berthou, E., Pascal, M., Pyšek, P., Sousa, R., Tabacchi, E., & Vilà, M. (2013). Impacts of biological invasions: what's what and the way forward. *Trends in Ecology & Evolution*, 28(1), 58–66. <https://doi.org/10.1016/j.tree.2012.07.013>
- Siripattawan, S., Park, J. K., & Ó Foighil, D. (2000). Two lineages of the introduced Asian freshwater clam corbicula occur in North America. *Journal of Molluscan Studies*, 66(3), 423–429. <https://doi.org/10.1093/mollus/66.3.423>
- Sousa, R., Antunes, C., & Guilhermino, L. (2006). Factors influencing the occurrence and distribution of *Corbicula fluminea* (Müller, 1774) in the river Lima estuary. *Annales de Limnologie - International Journal of Limnology*, 42, 165–171. <https://doi.org/10.1051/limn/2006017>
- Sousa, R., Novais, A., Costa, R., & Strayer, D. L. (2014). Invasive bivalves in fresh waters: Impacts from individuals to ecosystems and possible control strategies. *Hydrobiologia*, 735(1), 233–251. <https://doi.org/10.1007/s10750-012-1409-1>
- Strayer, D. L. (2010). Alien species in fresh waters: Ecological effects, interactions with other stressors, and prospects for the future. *Freshwater Biology*, 55(S1), 152–174. <https://doi.org/10.1111/j.1365-2427.2009.02380.x>
- Tchang, S., Li, S. C., & Liu, Y. Y. (1965). Bivalves (Mollusca) of Tung-ting Lake and its surrounding waters, Hunan Province, China. *Acta Zoologica Sinica*, 17(2), 197–216.
- Thompson, J. D., Higgins, D. G., & Gibson, T. J. (1994). CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22(22), 4673–4680. <https://doi.org/10.1093/nar/22.22.4673>
- Tuttle-Raycraft, S., & Ackerman, J. D. (2020). Evidence of phenotypic plasticity in the response of unionid mussels to turbidity. *Freshwater Biology*, 65(11), 1989–1996. <https://doi.org/10.1111/fwb.13595>
- Vastrade, M., Etoundi, E., Bournonville, T., Colinet, M., Debortoli, N., Hedtke, S. M., Emilien, N., Lise-Marie, P., Julie, V., Jean-François, F., & Van Doninck, K. (2022). Substantial genetic mixing among sexual and androgenetic lineages within the clam genus corbicula. *Peer Community Journal*, 2, e73. <https://doi.org/10.24072/pcjournal.180>
- Wang, G. P., Zhang, T., Zhang, J., Li, D. L., & Xiao, T. Y. (2014). Morphological and molecular differentiation of genus corbicula suggests that two species are sympatrically distributed in Datong Lake in the Central Yangtze River basin. *Zoological Studies*, 53(1), 64. <https://doi.org/10.1186/s40555-014-0064-9>
- Wang, J. P., Li, D. L., Zeng, C., Zhu, P. F., & Zhan, J. F. (2018). Genetic diversity and reproductive characteristics of *Corbicula fluminea* from the Laodao River in Liuyang City. *Acta Hydrobiologica Sinica*, 42(5), 913–918. <https://doi.org/10.7541/2018.112>
- Wang, J. P., Zhu, P. F., Wang, J., Gui, Y. T., & Li, D. L. (2018). Genetic diversity analysis of *Corbicula fluminea* in Dongting Lake based on the mitochondrial Cyt b gene. *Journal of Hydroecology*, 39(5), 89–94. <https://doi.org/10.15928/j.1674-3075.2018.05.013>
- Wilber, D. H., & Clarke, D. G. (2001). Biological effects of suspended sediments: A review of suspended sediment impacts on fish and shellfish with relation to dredging activities in estuaries. *North American Journal of Fisheries Management*, 21(4), 855–875. [https://doi.org/10.1577/1548-8675\(2001\)021<0855:BEOSSA>2.0.CO;2](https://doi.org/10.1577/1548-8675(2001)021<0855:BEOSSA>2.0.CO;2)
- Xu, H., Paerl, H. W., Qin, B., Zhu, G., & Gao, G. (2010). Nitrogen and phosphorus inputs control phytoplankton growth in eutrophic Lake Taihu, China. *Limnology and Oceanography*, 55(1), 420–432. <https://doi.org/10.4319/lo.2010.55.1.0420>
- Yamada, M., Ishibashi, R., Toyoda, K., Kawamura, K., & Komaru, A. (2014). Phylogeography of the brackish water clam *Corbicula japonica* around the Japanese archipelago inferred from mitochondrial COII gene sequences. *Zoological Science*, 31(3), 168–179. <https://doi.org/10.2108/zsj.31.168>
- Zhan, J. F. (2020). *Genetic structures and sex compositions of lake-inhabited Corbicula clams along the mid-lower Yangtze River*. (Master Thesis, Hunan Agricultural University, China). <https://doi.org/10.27136/d.cnki.ghunu.2020.000418>

Zhao, L., Schöne, B. R., & Mertz-Kraus, R. (2017). Delineating the role of calcium in shell formation and elemental composition of *Corbicula fluminea* (Bivalvia). *Hydrobiologia*, 790(1), 259–272. <https://doi.org/10.1007/s10750-016-3037-7>

#### BIOSKETCH

The research team is made up of researchers focusing on the evolutionary biology and ecology of invasive *Corbicula* complex. We aim to increase understanding of the relationships between environmental factors and *Corbicula*'s adaptation, genetic diversity and reproductive modes.

Author contributions: DLL and CZ conceived the idea. CZ and YXT performed data analysis and draft the manuscript. MV conducted data quality control, verified *COI* data analysis and revised the manuscript. NEC revised and improved the manuscript. KVD verified *COI* data analysis and revised the manuscript. TZ took part in data analysis and manuscript drafting. YJC measured and analysed environmental data. All authors contributed critically to the drafts and gave final approval for publication.

#### SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

**How to cite this article:** Zeng, C., Tang, Y., Vastrade, M., Coughlan, N. E., Zhang, T., Cai, Y., Van Doninck, K., & Li, D. (2023). Salinity appears to be the main factor shaping spatial *COI* diversity of *Corbicula* lineages within the Chinese Yangtze River Basin. *Diversity and Distributions*, 29, 364–378. <https://doi.org/10.1111/ddi.13666>