



RESEARCH ARTICLE

WILEY

Northern Europe's suitability for offshore European flat oyster (*Ostrea edulis*) habitat restoration: A mechanistic niche modelling approach

Brecht Stechele¹  | Anna Hughes² | Steven Degraer³ | Peter Bossier¹ | Nancy Nevejan¹ 

¹Laboratory of Aquaculture & Artemia Reference Center, Ghent University, Ghent, Belgium

²School of GeoSciences, University of Edinburgh, Edinburgh, UK

³Marine Ecology and Management, Operational Directorate Natural Environment, Royal Belgian Institute of Natural Sciences, Brussel, Belgium

Correspondence

Brecht Stechele, Laboratory of Aquaculture & Artemia Reference Center, Ghent University, Coupure Links 653, B-9000, Ghent, Belgium.
Email: brecht.stechele@hotmail.com

Funding information

Fonds Wetenschappelijk Onderzoek, Grant/Award Number: 1S84619N; Horizon 2020 Framework Programme, Grant/Award Number: 862915

Abstract

1. After centuries of overexploitation, European flat oyster (*Ostrea edulis*) habitats are increasingly being protected and large-scale oyster habitat restoration projects have been initiated in the North Sea, the English Channel, the Irish Sea and the north-eastern Atlantic.
2. Understanding the habitat requirements of flat oysters is key in the selection process of restoration sites.
3. In this study, a theoretical niche for all life stages of the flat oyster is determined using a dynamic energy budget model forced with temperature, chlorophyll *a* and salinity in combination with sediment composition. The suitability of Northern Europe's marine environment for flat oysters is evaluated by applying the proposed niche model on a spatial scale and evaluating spatio-temporal variability in life history traits and metabolic characteristics, such as fitness, reproductive output, settlement success and growth.
4. Results show that habitat suitability is limited by low maximum summer temperature and low food availability at higher latitudes and in areas under strong influence of the Atlantic Ocean. Restoration efforts in Ireland and Scotland should be confined to protected and shallow coastal environments. Suitability in the southern North Sea is patchy and mainly depends on seabed substrate. The nearshore areas of the English Channel are highly suitable.
5. The georeferenced dataset shared in this study will be useful for restoration practitioners.

KEYWORDS

dynamic energy budget, European flat oyster, habitat restoration, habitat suitability, niche model, northern Europe, *Ostrea edulis*, species distribution

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. *Aquatic Conservation: Marine and Freshwater Ecosystems* published by John Wiley & Sons Ltd.

1 | INTRODUCTION

Oyster reefs are recognized biodiversity hotspots that provide many ecosystem services (Coen et al., 2007; Smyth & Roberts, 2010; Kamermans et al., 2018; Crawford et al., 2019; Hancock & Ermgassen, 2019). Large scale oyster reef protection, restoration and creation projects have been initiated in many locations around the globe (Schulte, Burke & Lipcius, 2009; Gillies et al., 2015; Carranza & Ermgassen, 2020; Pogoda et al., 2020a) to reinstate or support degraded oyster habitats (Beck et al., 2011). Inshore and coastal European flat oyster habitat restoration is well established, but offshore restoration is generally overlooked due to high operational costs and logistical challenges. The rapid expansion of offshore wind in the North Sea and the deployment of hard substrate as scour protection material offer opportunities for large scale flat oyster reef restoration (Smaal et al., 2015; Kamermans et al., 2018; Bennema, Engelhard & Lindeboom, 2020). Understanding the environmental requirements for offshore flat oyster habitat restoration is key in the site selection process, and has been pinpointed as a significant knowledge gap (Pogoda et al., 2017; Pogoda et al., 2019; Pogoda et al., 2020a).

Two types of model are generally used to develop a spatio-temporal representation of species niches, both of which have their advantages and disadvantages. (i) Correlative niche models link species occurrence data to spatial datasets using statistical tools. Correlative niche models define the habitat, i.e. a description of the physical place where an organism actually or potentially lives (Kearney, 2006). These models do not include causal interactions between the environment and the species distribution, and therefore have poor predictive power when applied to new environments (for example, in the case of invasive species) or changing environments (for example, in case of climate change scenarios) (Davis et al., 1998; Dormann, 2007). Correlative niche models can be inconsistent when presence/absence data are scarce. (ii) Mechanistic niche models capture population processes explicitly as functional responses to environmental factors such as resource availability, temperature, salinity or oxygen availability (Kearney et al., 2008; Kearney et al., 2010; Teal et al., 2018). They aim to predict fitness components based on behavioural, morphological and physiological traits (Kearney & Porter, 2009), which again are based on the environmental space deemed suitable. Individual-based models such as the Dynamic Energy Budget (DEB) model (Kooijman, 2010) can be used in mechanistic niche models to enable the prediction of survival, reproduction, development and growth, and can provide information on population dynamics (Kearney, 2006; Holt, 2009; Kearney & Porter, 2009). Integrating DEB theory into biogeochemical models provides a framework that enables the evaluation of site suitability for organisms as well as the dynamics of life history events and metabolic traits (Kearney et al., 2008; Thomas et al., 2011; Teal et al., 2018; Palmer et al., 2020). This publication updates the theoretical niche for Northern Europe's population of flat oysters using a mechanical individual-based DEB model and evaluates the suitability of Northern Europe's marine environment for flat oysters.

2 | METHODS

2.1 | Geographical range and spatial data

The research domain for this study encompassed the North Sea, the English Channel, the Irish Sea and part of the north-east Atlantic Ocean, and stretches from 48° to 57°N and -9° to 9°E. This area includes the largest historical offshore flat oyster reefs in northern Europe (Figure 1a), and largely corresponds to the geographical range of the North Atlantic offshore populations (Launey et al., 2002). In the eastern Atlantic, the English Channel and the Irish Sea, coarse sand and muddy sea bed prevails offshore, while rocks and boulders are found nearshore. Apart from the sea bed off eastern England and north-west Denmark, North Sea sediments mainly consist of fine sand and mud, although rocks and boulders do occur sporadically (Figure 1b).

Drivers of the spatial model used in this study included bottom temperature, chlorophyll *a* and salinity and were extracted from the EU Copernicus Marine Service Information database (product id: NWSHELF_MULTIYEAR_PHY_004_009 and NWSHELF_MULTIYEAR_BGC_004_011). Bottom temperature was extracted using the 'sea_water_potential_temperature_at_sea_floor (bottomT)' variable (latitude × longitude × variable). Seawater salinity and chlorophyll *a* were extracted as depth profiles (latitude × longitude × depth × variable). Bottom salinities and bottom chlorophyll *a* concentrations were calculated by extracting the lowest numerical value of the vertical depth profile (personal communication Copernicus Marine Services).

Products were daily 25-hour, de-tided, averages. Extractions were made over a spatial scale of -9° to 9°E and 48° to 57°N from 1 January 2010 to 1 January 2020 at a timestep of 1 day. Horizontal resolution of the datasets is 7 km. European Marine Observation and Data Network (EMODnet) multiscale substrate maps (Folk 5) represent substrate suitability (Emodnet Geology, Kaskela et al., 2019).

The coarse horizontal resolution implies poor data representation for inshore and coastal environments. Inshore areas are defined as areas that are located in fjords, estuaries or coastal lagoons. Inshore waters are typically calm, shallow and are under high influence of the land. Coastal waters directly border the shoreline but are, in contrast to inshore waters, exposed to waves and wind (0–2 km offshore). Nearshore (2–20 km from the shore) and offshore areas (>20 km from the shore) are highly exposed to marine conditions. Cut-off values are set arbitrarily and only apply to this study.

2.2 | Dynamic Energy Budget model

2.2.1 | General

The individual-based model used in this study is underpinned by the DEB theory established by Kooijman (2010). Species-specific parameters describe the important life history traits of the individual (Kooijman, 2010; Lika et al., 2011). Metabolism is defined

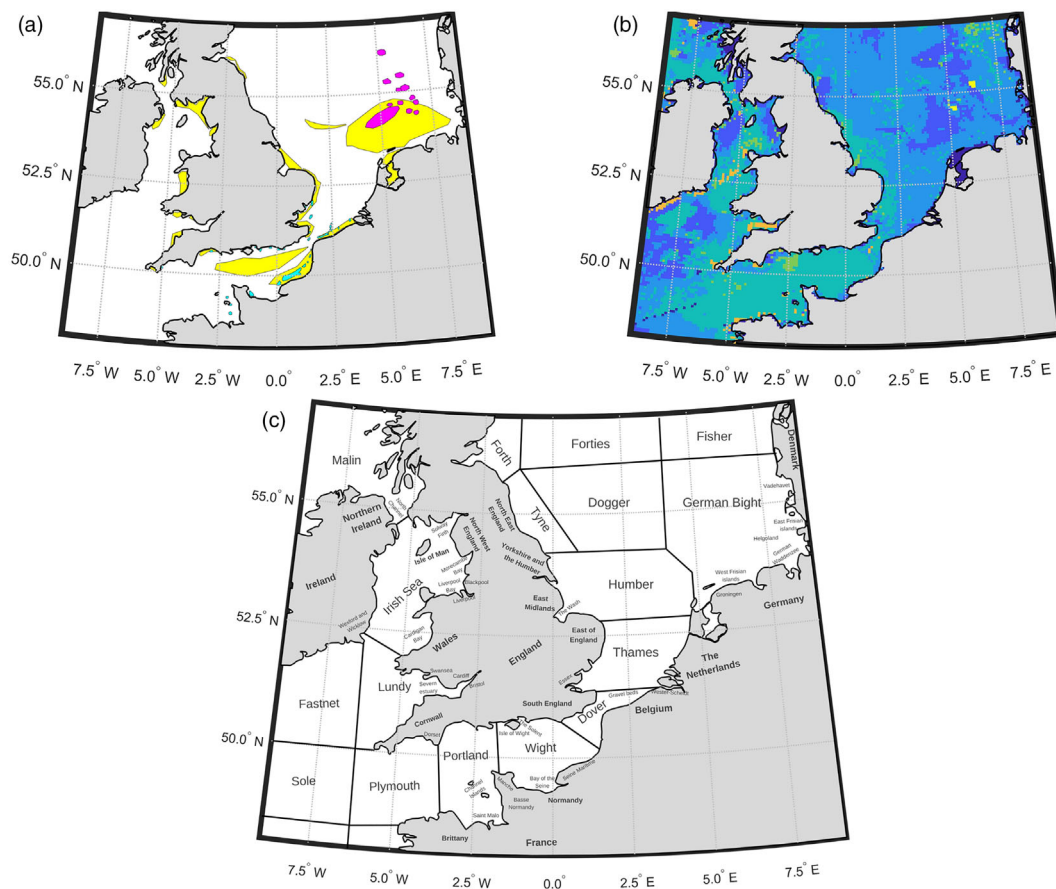


FIGURE 1 (a) Historical distribution of oyster habitats in northern Europe; representation adapted from Olsen's 'Piscatorial Atlas of the North Sea' (1883) (yellow), locations visited by oysterman M. H. Polley (Smaal et al., 2015) (blue) and oyster beds mapped by Gercken & Schmidt (2014) (red) (b) Seabed substrate type; mud (blue), fine sand (light blue), coarse sand (cyan), rocks and boulders (yellow), mixed sediment (green), absence of data (dark blue) (EMODnet Geology, Kaskela et al., 2019).

through four state variables; energy reserve, E (J); body structural volume, V (cm^3); the reproduction buffer, E_R (J); and the cumulative investment into development, called maturity, E_H (J) in DEB terminology.

According to Kooijman (2010), an individual's metabolism and energy flows (in joules) are a function of food availability and body temperature. Food availability relates to ingestion (\dot{p}_x) and assimilation of energy (\dot{p}_A) into the general reserve. Energy is mobilized (\dot{p}_C) from the reserve and is used for both somatic (κ) and reproductive ($1 - \kappa$) purposes. Energy allocated to somatic growth (\dot{p}_G) is used for the construction of functional tissue and overhead costs include the transformation of energy to structure. The functional maintenance (\dot{p}_S) flux maintains vital functions and has priority over growth. The remaining fraction $(1 - \kappa)\dot{p}_C$ of the mobilized energy is allocated to development (\dot{p}_R) (e.g. of the reproductive system, the immune system) if individuals are young and have not reached full maturity ($E_H < E_H^P$). When individuals are fully mature ($E_H = E_H^P$), the flux towards development is diverted toward the reproduction buffer. The reproduction buffer will be transformed into gametes, which will be released during reproduction. There is an additional energy cost for maintaining the state of maturation (\dot{p}_j).

The DEB model used in this study was parameterized and validated for the northern European flat oyster population (Stechele et al., 2022), and the DEB formulae and parameters are included as Supporting Information (S1, S2).

2.2.2 | DEB link to environment, validation and simulations

The DEB model used in this study was forced with chlorophyll a , temperature and salinity. Chlorophyll levels were linked to food availability using a Holling type II functional response (Kooijman, 2010), with X ($\mu\text{g L}^{-1}$) the environmental chlorophyll a concentration and K_X ($\mu\text{g L}^{-1}$) the food half saturation constant which indicates the chlorophyll a concentration that corresponds to $f_X = 0.5$, with f_X the food availability.

$$f_X = X(X + K_X)^{-1}$$

The effect of salinity on filtration rate is calculated according to Lavaud et al. (2017) and Lavaud et al. (2021), where c_s stands for the

salinity correction factor, S_H is the higher salinity threshold (above which salinity has no observed effect) and S_L is the lower salinity threshold at which filtration stops. Salinity does not influence the maintenance costs of oysters (Hutchinson & Hawkins, 1992; Lavaud et al., 2017).

$$C_S = \begin{cases} 1, & \text{at } S \geq S_H \\ \frac{S - S_L}{S_H - S_L}, & \text{at } S_L < S < S_H \\ 0, & \text{at } S \leq S_L \end{cases}$$

The response of ingestion rates to the environment then becomes:

$$\dot{P}_X = f_X C_s \{p_{Xm}\} V^{\frac{2}{3}}$$

with $\{p_{Xm}\} (J\text{cm}^{-2}\text{d}^{-1})$ representing the surface specific ingestion rate.

The link between the DEB model and its forcings (temperature, chlorophyll *a* and salinity) was calibrated using the Add-my-Pet procedure (Lika et al., 2011). The parameters z , K_X , S_L and S_H were optimized by minimizing the loss function in the four-dimensional parameter space using an iterative optimization routine based on the Nelder–Mead simplex algorithm. Optimization is based on the minimization of the weighted squared residuals (mean relative error, MRE) between model and observations (Lika et al., 2011).

During validation, initial state variables of the individual were fixed. V was recalculated from the initial dry weights or shell length. The reproduction buffer was assumed empty ($E_R = 0$). The initial maturity level was assumed to be maximum ($E_H = E_H^p$) and the energy reserve was set at 0.8 of its maximal capacity (this corresponds to a scaled energy density that represents good fitness; $e = 0.8$). See Stechele et al. (2022) for a detailed description of the calibration methodology.

2.3 | Suitability and life history trait simulations

Both the dynamics of life history traits and the metabolism are closely linked to site suitability. Locations determined as suitable should provide an environment that produces fit and reproductive individuals. DEB life traits such as minimum adult fitness, the occurrence of spawning events, the average yearly reproductive output and the age at which individuals mature are indicators of a suitable niche.

The adult fitness and reproductive output were simulated starting from large ($L = 7.5\text{ cm}$) and fit ($e = 0.8$) adults (with initial state variables: $[E = 2.31 \cdot 10^4\text{ J}, V = 9\text{ cm}^3, E_H = E_H^p, E_R = 200\text{ J}]$). Fitness in DEB theory is expressed as the scaled energy reserve ($e = E \cdot V^{-1} \cdot E_m^{-1}$), which is calculated using DEB state variables E and V and the DEB compound parameter E_m ($J\text{ cm}^{-3}$). Fitness is expressed in a value between 0 (no energy budget to meet maintenance demands) and 1 (energy reserves are complete) and is an indicator comparable to the condition index. The average reproductive output of one individual,

represented by the number of larvae released during a spawning event ($\kappa_R E_R E_0^{-1}$), with κ_R the reproduction efficiency and E_0 the energy content of an unfertilized egg. Reproduction was triggered by spring temperatures reaching $15\text{ }^\circ\text{C}$ (Orton, 1937; Korringa, 1957; Laing, Walker & Areal, 2005; Bromley et al., 2016; McGonigle, Jordan & Geddis, 2016). Second spawning peaks or continuous spawning events were not considered within the scope of this model. The age at which maturity is reached is an indicator of growth rate but also for the timing of first reproduction. Individuals that are mature have a fully developed immune system (Kooijman, 2010) and background mortalities after full maturation are generally low (Walne, 1961). The time required for individuals to reach full maturity (starting from the moment of release) was simulated starting from released larvae ($e = 0.8, L = 200\text{ }\mu\text{m}$; with initial state variables: $[E = 0.0019\text{ J}, V = 2.8 \cdot 10^{-6}\text{ cm}^3, E_H = E_H^r, E_R = 0\text{ J}]$).

A Habitat Suitability Index (HSI) was constructed based on life history and metabolic performance. Pixels with minimum adult fitness levels of $e < 0.3$ were considered unsuitable and were given a value of 0. Reproductive output and age at which maturity was reached, were scaled to values between 0 and 1. Fine sand sediments are unsuitable for settlement and were therefore given a value of 0. All other sediment types were considered suitable.

$$HSI = \sqrt[4]{\text{Fitness} \cdot \text{Reproduction} \cdot \text{Growth} \cdot \text{Sediment suitability}}$$

In the present study, scripts were run using MATLAB version R2019b. All rendered maps are georeferenced and raw output grids are included as Supporting Information (S3).

3 | RESULTS

3.1 | DEB calibration and validation

DEB parameters z , K_X , S_L and S_H were optimized, based on growth curves from the German Bight (dry tissues weight), the Belgian part of the North Sea (shell length) and filtration rates at different salinities collected in the lab (Hutchinson & Hawkins, 1992). These growth curves span three different growth periods (2004, 2007 and 2017–2019), originate from cultivation in inshore, nearshore and offshore conditions, for several size classes (1.1–7.0 cm). The calibration included parameterization of the metabolic responses to food and salinity. The metabolic response to temperature has been already calibrated by Stechele et al. (2022).

The calibration resulted in a good model-to-data fit, expressed in a total MRE of 0.12 (values below 0.2 are acceptable) which indicates a good coupling between the DEB and the ‘Atlantic-European Northwest Shelf- Ocean Physics and Biogeochemical’ CMEMS datasets (Figure 2).

Estimates of growth at the German Bight (Figure 2a,b) were acceptable, and although spring growth was underestimated, estimations of the final dry weights were good. R^2 values are close to one and indicate sufficient precision of the model.

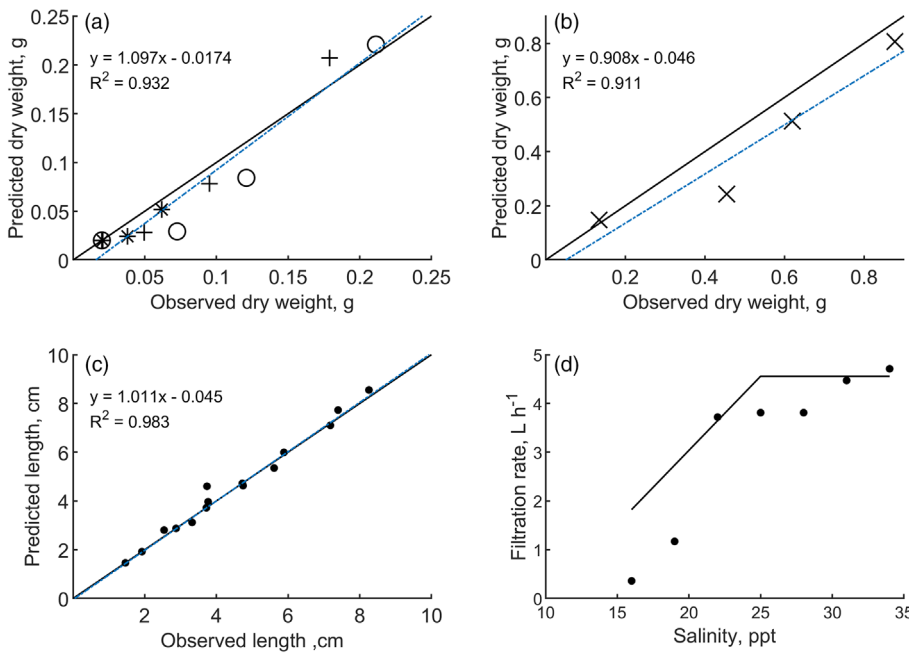


FIGURE 2 Model to data fitting of: (a) Dry weight growth data of juvenile oyster in Butendiek (o), Helgoland (+) and Wursterarm (*), collected in the German Bight (Pogoda, Buck & Hagen, 2011); (b) Dry weight growth data of juvenile oysters in the second year of aquaculture growth at Nordergründe, Germany (Pogoda, Buck & Hagen, 2011). (c) Shell lengths collected from aquaculture growth experiments at Westdiep, Belgium (Delbare, 2019). (d) Filtration rates at different salinities observed during laboratory experiments (Hutchinson & Hawkins, 1992).

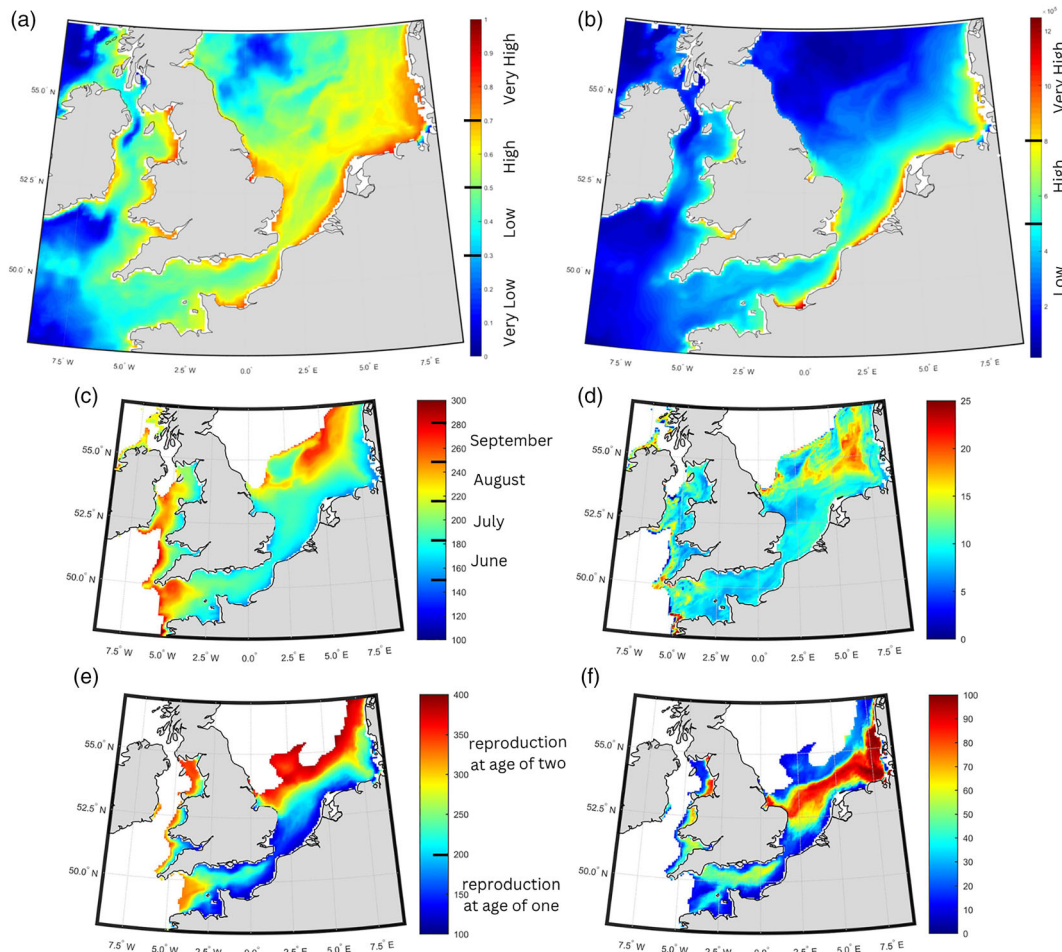


FIGURE 3 (a) Minimum fitness of a European flat oyster adult (7.5 cm). (b) Average yearly reproductive output of an adult oyster (expressed in number of larvae released per adult). Average (c) and standard deviation (d) estimates of the reproduction time based on 15°C temperature threshold (expressed as Julian day of the year). Average (e) and standard deviation (f) of the age at which European flat oyster juveniles reach full maturity (expressed in days since fertilization). All results cover the 10-year period from 2010–2020.

Growth curves collected from different size classes (T8 – adults) in the Belgian part of the North Sea (Figure 2c) were estimated very well. T8, T12 and T20 oysters doubled in size during the first year of growth. T25 oysters grew from 3.72 cm to 5.88 cm and adults from 7.18 cm to 8.25 cm.

The influence of salinity on physiological response was calibrated based on the data of Hutchinson & Hawkins (1992) and the growth data at the Wursterarm location in the German Bight, which is located in the mouth of the River Weser at Bremerhaven and therefore often experiences low salinities (Pogoda, Buck & Hagen, 2011; Stechele et al., 2022). The effect of salinity on the feeding mechanism is captured well. Growth at low salinity (Wursterarm, Germany) was estimated well, but mortalities could not be reproduced with a salinity response defined by $S_L = 10$ ppt and $S_H = 25$ ppt (Pogoda, Buck & Hagen, 2011). The observations of Hutchinson & Hawkins (1992) were generally estimated well (Figure 2d), although the filtration at low salinities was overestimated. Filtration rates at 15 ppt were observed to be close to zero, while our model predicted filtration rates of 2 L h^{-1} .

3.2 | Northern Europe's suitability based on DEB

Distinct environments can be linked to oyster performance and life history traits (Figure 3) leading to a heterogeneous distribution of suitable areas (Figure 4) over the research domain.

3.2.1 | Atlantic

Areas that are heavily influenced by Atlantic waters, including the northern and southern offshore areas of Ireland (Malin, Fastnet and Sole), the offshore areas off Cornwall (UK), Brittany (France) and the northern North Sea regions (Forties, Forth and Tyne), are generally unsuitable ($HSI < 0.3$) for flat oysters. Atlantic waters in these areas never reach the minimum threshold for spawning and periods of very

low fitness (<0.3) occur due to low food availability. In these marine areas, historical reefs and habitat restoration projects can be present, but are generally located in warmer, nutrient rich, inshore, estuarine, coastal or nearshore areas.

3.2.2 | Irish Sea

The environment in the northern offshore areas of the Irish Sea is unsuitable for flat oyster restoration. The North Channel and offshore areas between Isle of Man and Northern Ireland, are under Atlantic influence and seabed temperatures are too cold for spawning. Northern offshore areas in the Irish Sea are limited in food and fitness in these areas is very low to low (0.3–0.5). Some areas with high (0.5–0.7) to very high HSI (>0.7) do occur in the eastern parts of the Irish Sea, for example off the coast of north-western England and Wales (Solway Firth, Morecambe Bay, the coastal plain of Mersey and Cardigan Bay). The oyster fitness in these nearshore areas is high (0.5–0.7). The reproductive output of fully grown adults is expected to be high (0.5–0.8 million larvae). Reproduction occurs early in summer (during June and July). Flat oyster growth is slow and in the best case, maturity is reached after 250 days making it unlikely that the oysters will spawn during their first year of life. Similar conditions are present in the nearshore areas off the counties of Wicklow and Wexford in the south-western parts of the Irish Sea.

3.2.3 | The English Channel

The offshore western parts of the English Channel (Plymouth and Portland) are heavily influenced by the Atlantic waters and very high HSI for oysters occurs nearshore, around the coast of Dorset (UK), the Channel Islands (UK) and in the Bay of St Malo (France). The eastern parts of the English Channel are indicated by a high HSI offshore and very high HSI nearshore. The minimum fitness of oysters is high to very high throughout the eastern English Channel indicating

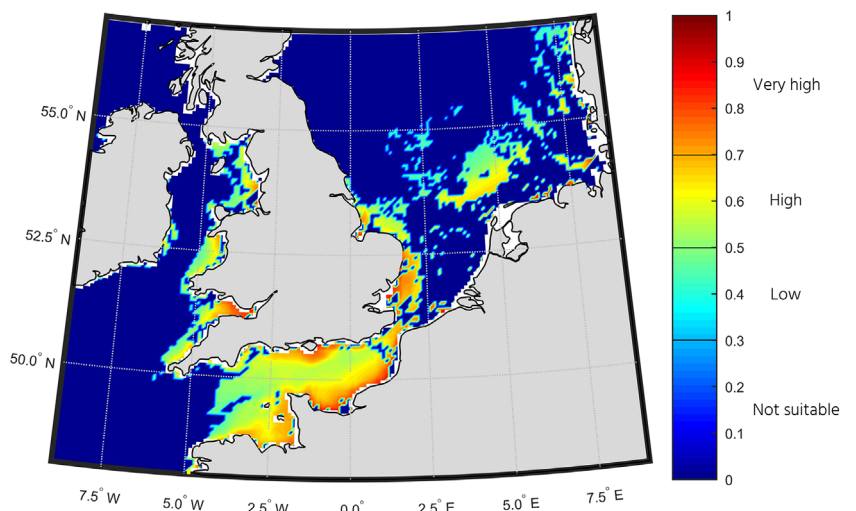


FIGURE 4 Habitat Suitability Index for European flat oyster (*Ostrea edulis*); Suitable locations support locations that support high fitness, growth rate and reproductive output and enable life history events such as spawning and settlement (on suitable substrate).

the absence of starvation periods. Reproductive output is high offshore and very high nearshore (>0.8 million larvae) around the Solent (UK), Manche and Seine Maritime (France). In the nearshore areas of Normandy and South England oysters mature quickly (before day 200) and first reproduction can be expected at the age of 1 year. These areas are likely to support high growth rates.

3.2.4 | North Sea

In the northern offshore parts of the North Sea (Forties) as well as the areas off the coast of north-eastern England and the Yorkshire and the Humber region (UK), suitable niches for flat oysters do not occur offshore or nearshore and restoration efforts as well as historical reefs are located inshore or in coastal areas (Firth of Forth, Dornoch Firth and The Humber). In the Southern Bight of the North Sea (Thames, Dover and Humber), covering the nearshore and offshore areas off the East Midlands, East of England (UK) and the Belgian and Dutch offshore territories, the flat oyster niche is present, and HSI is high to very high. In these areas, oysters have high to very high (>0.7) average fitness levels and reproduction events occur generally in June (Thames and Dover) and July (Humber). Reproductive output is high offshore, and very high nearshore (>0.8 million larvae per spawning event). Low reproduction rates (<0.5 million larvae per spawning event) occur in the north Humber offshore areas. In Thames and Dover, maturity is reached quickly (<200 days) and oysters are expected to reproduce at the age of one. Growth in the Southern Bight is expected to be fast. HSI in the German Bight and the Dogger Bank is high but decreases towards the north (low HSI). Compared to the Southern Bight, spawning events in the Dogger Bank occur later in the summer (July–August) and reproductive output is generally low (<0.5 million larvae per spawning event). Although in general, coastal areas of the German Bight have similar HSI to coastal areas in the Southern Bight (Thames and Dover), the outflow of the Elbe and the Weser rivers creates unsuitable conditions for flat oysters in and around the German Waddenzee.

3.3 | Restoration hotspots

The spatial distribution of suitable niches for flat oysters depends on the spatial variability in metabolic performance (survival, growth, reproduction) and life-history events such as successful spawning and settlement which are linked to substrate suitability. Combining life-history process indicators into one habitat suitability indicator enables the identification of areas suitable for flat oysters, including the presence of hotspots (HSI > 0.7), with very high suitability for flat oysters (Figure 4).

Hotspots occur in coastal or nearshore areas. In and around Ireland, hotspots for flat oysters only occur in the south east on the coasts of Wexford and Wicklow. In England and Wales, some coastal areas can specifically be highlighted as hotspots, such as

the coastal areas close to Liverpool and Blackpool, the outer Severn estuary around Bristol, Cardiff and Swansea, the Solent, the Blackwater estuary in Essex and the Wash. On the coast of northern France, hotspots exist in the wider Bay of the Seine and off the coast of Basse Normandie. In Belgium, hotspots exist in the southern Wester-Scheldt estuary, and on the offshore gravel beds. In the Netherlands, hotspots occur off the coast of Groningen. In Germany, hotspots occur around the East Frisian islands, and in and around the area between Helgoland and the West Frisian Islands. In Denmark, hotspots occur off the National Park Vadehavet.

4 | DISCUSSION

4.1 | General

Site suitability studies for flat oyster beds have been performed at national and regional levels and are generally correlative niche models that statistically link environmental conditions to absence/presence data of reefs (Robinson et al., 2017; Kamermans et al., 2018; Bennema, Engelhard & Lindeboom, 2020; Pogoda et al., 2020b; Bergström et al., 2021). Although correlative niche models are useful tools for identifying niches, they are often inconsistent when establishing the niche for endangered species for which presence data are scarce; or unreliable when using historical datasets. The use of historical datasets for correlative niche modelling is potentially problematic for two reasons: (i) historical data are often incorrect, as shown by Bennema, Engelhard & Lindeboom (2020) in the case of flat oysters; and (ii) climate change has altered species distribution in the North Sea (Kröncke et al., 2011), so relating historical presence data to environmental data from contemporary climate conditions will result in the identification of the wrong environmental space deemed suitable for the species (Sillero et al., 2021).

In addition, correlative niche models identify a set of environmental factors linked to species presence data. Which factors to include in the analysis, and the relative importance of these factors differs between studies and sites. van Duren, Kamermans & Kleissen (2022) tested a set of 10 environmental factors, Pogoda et al. (2020b) included six environmental factors while de Mesel et al. (2018) included three environmental factors in their habitat suitability studies.

A complete mechanistic niche model, by contrast, includes all known influences of the environment on the metabolism of the individual and generates traits that are easy to evaluate with regards to suitability. Suitability indicators such as fitness and reproductive output are calculated based on a combination of factors that change over time and space. Therefore, DEB-based mechanistic niche models enable spatio-temporal estimation of the variability of the condition of the individual and therefore can explain, for example, summer or winter mortalities, starvation, or failed reproduction events.

4.2 | Northern Europe's suitability for the flat oyster

The suitability of Northern Europe's marine environment for the flat oyster is limited by various factors. In the north and west of the research area, no flat oysters are expected to occur in offshore and nearshore waters because summer temperatures never reach the spawning threshold of 15°C. Presence data (observations of flat oyster habitats) in northern latitudes (Olsen, 1883; SS, 1889), are limited to shallow inshore and coastal habitats where seawater temperature does exceed the spawning threshold in summer (these environments are not covered by this study). Therefore, suitable locations for flat oysters are limited to coastal and inshore environments in Scotland, Ireland and the eastern coast of England (personal communications, Prof. William Sanderson and Oliver Tully). Where the current study used a threshold of 15°C, various spawning temperature thresholds have been recorded for flat oysters (Mesel et al., 2018). Oysters from the offshore deep water beds in the English Channel and the Helgoland Bank were historically assumed to release spawn at colder temperatures (Korringa, 1957). Unfortunately, these populations that historically thrived in colder waters no longer exist (Thurstan et al., 2013; Bennema, Engelhard & Lindeboom, 2020). Thermo-tolerance and temperature spawning thresholds should be examined when considering translocating flat oysters for restoration. Besides a temperature threshold, several studies recommend the use of temperature-sum (i.e. the accumulated temperature when higher than a threshold temperature, measured in 'degree-days') as a predictor for reproduction timing (Maathuis et al., 2020; Chapman et al., 2021). Nevertheless, temperature-sum is location dependent (Chapman et al., 2021) and non-mechanistic, so can therefore not be used in spatial applications. For example, Maathuis et al. (2020) state that in the Oosterschelde, The Netherlands, a temperature sum of 493–661 degree-days indicates the first larval peak, while in Lake Grevelingen, the first larval peak is expected at a temperature sum of between 313 and 1,100 degree days. Bernard, de Kermoisan & Pouvreau (2011) modified the DEB model to incorporate gonad development in spring and predicted that spawning time is based on a gonado-somatic index as well as a temperature threshold. Including this adaptation to the flat oyster DEB model would optimize estimates of the timing of spawning events.

Unsuitable feeding conditions occur in areas bordering the Atlantic Ocean due to food deprivation. Bottom layers in deeper waters where thermal stratification is strong receive less phytoplankton. Coastal areas that are highly influenced by river runoff are unsuitable for flat oysters as low salinities reduce filtration and ingestion rates. Aside from these factors, suitability is strongly limited by substrate. Flat oyster larvae are able to settle on shell, bedrock, cobble, gravel, boulders, stones or artificial hard substrate (Barry, 1981; Héral & Deslous-Paoli, 1991; Perry & Jackson, 2017). Historically reefs were often present in habitats where hard, muddy or clay sediments prevail (Laing, Walker & Areal, 2005; Perry & Jackson, 2017; Christianen et al., 2018). There are various hypotheses as to why sandy sediments are unsuitable for example:

(ii) increased near-bed flow over smooth, flat sandy bottoms decreases settlement success (Rodriguez-Perez, 2020); or (ii) high bottom shear stress in combination with sandy sediment results in a mobile sea bed and resuspension of sand, which can cause burial of reefs, or inhibition of feeding (Laing, Walker & Areal, 2005; Kamermans et al., 2018).

4.3 | Completing the theoretical niche

The current study focuses on establishing the theoretical niche of the European flat oyster based on the DEB model. The DEB model successfully links environmental variables to the metabolic performance of individuals. The DEB forcings included in this study were chlorophyll *a*, seawater temperature and salinity. Several other environmental variables such as oxygen, suspended particulate matter, current speed, shear stress and food quality, are known to influence oyster metabolism and including these variables in future iterations will complete the theoretical niche of flat oysters (Bennema, Engelhard & Lindeboom, 2020). However, these variables are not included in this analysis at present because: (i) no scientific observations are currently available to mechanistically link additional variables to the metabolic performance of flat oysters; (ii) management strategies can influence variables such as current speed and shear stress; and (iii) these variables are not expected to influence the metabolism in the geographical range researched in this publication.

The flat oyster's metabolic response to hypoxia has not been studied, but marine bivalves in general can tolerate low oxygen concentrations (Vaquer-Sunyer & Duarte, 2008) (lethal concentration-50 of 1.42 mg O₂ L⁻¹ and a lethal time-50 of 412 h). The study area is typically well-oxygenated, and therefore hypoxia is not expected. Limited information is available regarding the tolerance of flat oysters to suspended solids. Presence-absence data (unpublished dataset Historical Ecology working group NORA) suggest that flat oysters are resilient to high concentrations of suspended material. Duchêne, Bernard & Pouvreau (2015) gave an upper limit of 50 mg L⁻¹ while Hutchinson & Hawkins (1992) gave a negative correlation between filtration rate and particulate organic matter (algae) concentrations of 5 to 10 mg L⁻¹. Oysters grown in the Dutch part of the North Sea are often exposed to conditions well exceeding 50 mg L⁻¹ (Kamermans et al., 2018) but data are too scarce to calibrate and include a particle selection algorithm (Kooijman, 2010) in the proposed DEB model. Current speed can affect filtration rates (Walne, 1961), but more importantly, current speed and related shear stress are factors that can affect settlement success. These factors are variable on a small scale and restoration practices (e.g. deployment of cultch, artificial reefs, scour protection material) and local seabed characteristics (e.g. presence of boulders, rocks, banks) can greatly influence these factors. The resolution of this study is not compatible with the scales at which current speed or shear stress influence the oyster's life cycle and therefore these factors have not been included.

4.4 | Suitability for flat oyster reef formation

The current study evaluates the suitability of northern European waters for flat oysters, indicating which locations have a set of environmental conditions that are potentially suitable for oysters to survive, grow and reproduce. Nevertheless, suitability for an individual is not the same as suitability for populations; and for population numbers to increase, additional environmental conditions will have to be present (e.g. low predation pressure, swarming patterns that support self-recruitment). Since populations exist of individuals, populations will always occur at locations that are suitable for individuals, and reefs should therefore be present at locations indicated as suitable in this study.

Historical and current flat oyster reefs (Olsen, 1883; Lübbert, 1906; Gercken & Schmidt, 2014; Mesel et al., 2018) generally correspond to the hotspots identified in this study (Figure 1a). This can be exemplified using Olsen's historical map (Olsen, 1883), which shows large areas where oyster reefs were historically present. However, according to Olsen, oyster beds were found on sandy substrate (e.g. parts of the Oyster Grounds in Germany and The Netherlands, the Doggerbank, and Belgian coastal areas), which does not correspond with the assumptions related to settlement proposed in this study. Olsen also reports the presence of nearshore oyster reefs off the north east of England and the east of Ireland, which were areas deemed unsuitable ($HIS < 0.3$) in this study, and Bennema, Engelhard & Lindeboom (2020), have shown that Olsen's historical distribution map cannot be regarded as an unambiguous source and locations of some oyster beds are questionable. A more detailed map of deep-sea reefs in the English Channel frequently visited by oysterman M. H. Polley (Smaal et al., 2015) corresponds with the results of this study, and all historical reefs overlap with hotspots indicated in this study. The oyster beds mapped by Gercken & Schmidt (2014) in the German Bight, largely overlap with the presented hotspots (Figure 4), although some reefs are located on sandy substrate. Also, the current distribution of reefs in the English Channel (Mesel et al., 2018) corresponds fully with the results of this study.

4.5 | Towards a realistic niche model for habitat restoration

To establish models that predict the suitability for flat oyster populations, there is a need to extend the current theoretical niche model towards a realistic niche model by integrating population, dispersal and ecosystem models. Where a theoretical niche model only includes the physical and biological environment, and only estimates suitability on an individual level, realistic niche models also include species interactions (often in the form of competition and predation) and connectivity.

Predation is challenging to quantify in the field (Smaal et al., 2015). Oysters are preyed upon by a variety of species, and often experience high levels of predation from starfish (such as *Asterias rubens*), shore and edible crabs (*Carcinus maenas* and *Cancer*

pagurus), and predatory snails such as the sting winkle (*Ocenebra erinacea*) and the common whelk (*Buccinum undatum*) (Smaal et al., 2015; Perry & Jackson, 2017; Mesel et al., 2018). The invasive American oyster drill (*Urosalpinx cinerea*) predates almost exclusively on oyster spat, leading to low levels of survival in young oysters where *U. cinerea* are prevalent (Fey-Hofstede et al., 2010; Perry & Jackson, 2017; Smaal et al., 2017). Young oysters (<3 cm) have thin, weak shells, which are easily penetrated by predators (Gercken & Schmidt, 2014). More work is required to understand and quantify the effect of predation of population dynamics.

Complex interactions leading to interspecific competition are again difficult to quantify but can influence both survival and growth of native oysters (Smaal et al., 2015). Two major types of competition act upon oysters: (i) competition for food; and (ii) competition for settlement area. Other filter feeding organisms compete with oysters for food suspended in the water column. Competitors for settlement space include but are not limited to tube-building polychaetes (*Sabellaria* sp. and *Spirobranchus triqueter*) barnacles (*Balanus crenatus*), anthozoans, ascidians and bryozoans (Mesel et al., 2018). Perhaps the most impactful form of competition is through the introduction of the invasive American slipper limpet (*Crepidula fornicata*) (Helmer et al., 2019; Preston et al., 2020). This species was brought to Europe from North America in the late 1800s through the expansion of commercial aquaculture of the Pacific oyster (*Magallana gigas/Crassostrea gigas*) and Eastern oysters (*Crassostrea virginica*). Once introduced, *C. fornicata* populations expanded rapidly, and competed with the native oyster for both food and settlement area (Perry & Jackson, 2017). The Pacific oyster itself is also thought to compete with native oysters for food (Zwerschke et al., 2018; Ezgeta-Balić et al., 2020). Some studies, however, suggest that this competition is outweighed by the benefits Pacific oysters bring in increasing stability of the biogenic reef system and providing settlement area, which benefits the native oyster (Christianen et al., 2018).

Connectivity with broodstock is also important to include when determining suitable locations for reef restoration. Broodstock may be naturally occurring reefs or beds, broodstock can also be installed to support restoration projects. In addition, aquaculture sites can also provide larvae to support restoration efforts. Larval connectivity and hence connectivity between adult populations is determined primarily by abiotic factors such as tides, current velocity and trajectory.

When population models that include inter-individual and ecosystem interactions are established, it will become possible to quantify and predict population dynamics. Evaluating location-specific population dynamics will allow accurate site selection and will also enable the evaluation of restoration management strategies in the long term.

5 | CONCLUSION

With increasing interest for flat oyster habitat restoration in northern Europe, the development of site selection tools becomes essential. This coupling of an individual based DEB model with biogeographical datasets provides information on flat oyster fitness, growth potential,

reproductive output and timing of reproduction events on a temporal and spatial scale. Applying this theoretical niche model to northern Europe's offshore and nearshore waters enables the identification of locations that are unsuitable for flat oysters and therefore for flat oyster reef restoration. This model also identifies hotspots where environmental conditions are highly suitable for flat oysters. These hotspots for individuals generally correspond to locations where historical populations were present. The theoretical niche model used in this study can be effectively applied to identify suitable locations outside of the research areas (inshore and coastal areas or areas outside of the geographical range). Further research is required to identify suitable locations for flat oyster populations or flat oyster reef restoration using realistic niche models, which are extensions of theoretical niche models.

AUTHOR CONTRIBUTIONS

Brecht Stechele: Conceptualization; formal analysis; methodology; software; validation; visualization; writing—original draft; writing—review and editing. **Anna Hughes:** Conceptualization; methodology; writing—original draft; writing—review and editing. **Steven Degraer:** Conceptualization; funding acquisition; methodology; project administration; resources; writing—review and editing. **Peter Bossier:** Conceptualization; funding acquisition; project administration; writing—original draft; writing—review and editing. **Nancy Nevejan:** Conceptualization; funding acquisition; project administration; writing—review and editing.

ACKNOWLEDGEMENTS

This work was supported by the Research Foundation—Flanders (FWO) through an SB PhD fellowship granted to Brecht Stechele [project number 1S84619N]. The work was also funded by the European Commission HORIZON 2020 Grant Agreement nr 862915 (Views and opinions are those of authors only and do not necessarily reflect those of the European Union. Neither the European Union nor the Granting Authority can be held responsible for them). This study has been conducted using E.U. Copernicus Marine Service Information; <https://doi.org/10.48670/moi-00054> and <https://doi.org/10.48670/moi-00058>. I would like to thank Romain Lavaud for his input on DEB applications to shellfish.

CONFLICT OF INTEREST STATEMENT

The authors declare that there is no conflict of interest.

DATA AVAILABILITY STATEMENT

All data has been submitted as supplementary information.

ORCID

Brecht Stechele  <https://orcid.org/0000-0001-8450-1133>

Nancy Nevejan  <https://orcid.org/0000-0001-6566-4607>

REFERENCES

Barry, M.D. (1981). *Distribution and ecology of oysters, Ostrea edulis (L.) in Killieran and Bertraghboy bays*. Connemara, Co. Galway.

- Beck, M.W., Brumbaugh, R.D., Airoidi, L., Carranza, A., Coen, L.D., Crawford, C. et al. (2011). Oyster reefs at risk and recommendations for conservation, restoration, and management. *Bioscience*, 61(2), 107–116. <https://doi.org/10.1525/bio.2011.61.2.5>
- Bennema, F.P., Engelhard, G.H. & Lindeboom, H. (2020). *Ostrea edulis* beds in the Central North Sea: delineation, ecology, and restoration. *ICES Journal of Marine Science*, 77(7–8), 2694–2705. <https://doi.org/10.1093/icesjms/fsaa134>
- Bergström, P., Thorngren, L., Strand, Å. & Lindegarth, M. (2021). Identifying high-density areas of oysters using species distribution modeling: lessons for conservation of the native *Ostrea edulis* and management of the invasive *Magallana (Crassostrea) gigas* in Sweden. *Ecology and Evolution*, 11(10), 5522–5532. <https://doi.org/10.1002/ece3.7451>
- Bernard, I., de Kermoisan, G. & Pouvreau, S. (2011). Effect of phytoplankton and temperature on the reproduction of the Pacific oyster *Crassostrea gigas*: investigation through DEB theory. *Journal of Sea Research*, 66(4), 349–360. <https://doi.org/10.1016/j.seares.2011.07.009>
- Bromley, C., McGonigle, C., Ashton, E.C. & Roberts, D. (2016). Bad moves: pros and cons of moving oysters – a case study of global translocations of *Ostrea edulis* Linnaeus, 1758 (Mollusca: Bivalvia). *Ocean and Coastal Management*, 122, 103–115. <https://doi.org/10.1016/j.ocecoaman.2015.12.012>
- Carranza, A. & Ergassan, P.S. (2020). A global overview of restorative shellfish Mariculture. *Frontiers in Marine Science*, 7, 722. <https://doi.org/10.3389/fmars.2020.00722>
- Chapman, E.C.N., Rodriguez-Perez, A., Hugh-Jones, T., Bromley, C., James, M.A., Diele, K. et al. (2021). Optimising recruitment in habitat creation for the native European oyster (*Ostrea edulis*): implications of temporal and spatial variability in larval abundance. *Marine Pollution Bulletin*, 170, 112579. <https://doi.org/10.1016/j.marpolbul.2021.112579>
- Christianen, M., Lengkeek, W., Bergsma, J., Coolen, J., Didderen, K., Dorenbosch, M. et al. (2018). Return of the native facilitated by the invasive? Population composition, substrate preferences and epibenthic species richness of a recently discovered shellfish reef with native European flat oysters (*Ostrea edulis*) in the North Sea. *Marine Biology Research*, 14(6), 590–597. <https://doi.org/10.1080/17451000.2018.1498520>
- Coen, L.D., Brumbaugh, R.D., Bushek, D., Grizzle, R., Luckenbach, M.W., Posey, M.H. et al. (2007). Ecosystem services related to oyster restoration. *Marine Ecology Progress Series*, 341, 303–307. <https://doi.org/10.3354/meps341303>
- Crawford, C., Edgar, G., Gillies, C.L., Heller-Wagner, G., Crawford, C., Edgar, G. et al. (2019). Relationship of biological communities to habitat structure on the largest remnant flat oyster reef (*Ostrea angasi*) in Australia. *Marine and Freshwater Research*, 71(8), 972–983. <https://doi.org/10.1071/MF19171>
- Davis, A.J., Jenkinson, L.S., Lawton, J.H., Shorrocks, B. & Wood, S. (1998). Making mistakes when predicting shifts in species range in response to global warming. *Nature*, 391(6669), 783–786. <https://doi.org/10.1038/35842>
- Delbare, D. (2019). Eindrapport Value@Sea - Geïntegreerde teelt van extractieve aquacultuursoorten (EFMZV project. (16/UP2/04/Aqua/368)), 89.
- Dormann, C.F. (2007). Promising the future? Global change projections of species distributions. *Basic and Applied Ecology*, 8(5), 387–397. <https://doi.org/10.1016/j.baae.2006.11.001>
- Duchêne, J., Bernard, I. & Pouvreau, S. (2015). Programme d'Expérimentation et de recherche sur L'huître plate *Ostrea edulis* Vers un retour de l'huître indigène en rade de Brest. Version complète de l'article paru dans la revue Espèces, (16).
- Ezgeta-Balić, D., Šantić, D., Šegvić-Bubić, T., Bojanić, N., Bužančić, M., Vidjak, O. et al. (2020). Competitive feeding interactions between

- native *Ostrea Edulis* and non-native *Crassostrea gigas* with implications of introducing *C. gigas* into commercial aquaculture in the eastern Adriatic Sea. *Marine Environmental Research*, 160, 105051. <https://doi.org/10.1016/j.marenvres.2020.105051>
- Fey-Hofstede, F.E., van den Brink, A.M., Wijsman, J.W.M. & Bos, O.G. (2010). Risk assessment on the possible introduction of three predatory snails (*Ocenebrellus inornatus*, *Urosalpinx cinerea*, *Rapana venosa*) in the Dutch Wadden Sea. Report/IMARES Wageningen UR; No. no. C032/10. <https://edepot.wur.nl/143118>
- Gercken, J. & Schmidt, A. (2014). Current status of the European oyster (*Ostrea edulis*) and possibilities for restoration in the German North Sea. *Bfn-Skripten*, 379, 107.
- Gillies, C.L., Fitzsimons, J.A., Branigan, S., Hale, L., Hancock, B. & Creighton, C. (2015). Scaling-up marine restoration efforts in Australia. *Ecological Management and Restoration*, 16(2), 84–85. <https://doi.org/10.1111/emr.12159>
- Hancock, B. & Ermgassen, P. (2019). Enhanced production of finfish and large crustaceans by bivalve reefs. In: *Goods and Services of Marine Bivalves*. Cham: Springer, pp. 295–312.
- Helmer, L., Farrell, P., Hendy, I., Harding, S., Robertson, M. & Preston, J. (2019). Active management is required to turn the tide for depleted *Ostrea edulis* stocks from the effects of overfishing, disease and invasive species. *PeerJ*, 7, 6431. <https://doi.org/10.7717/peerj.6431>
- Héral, M. & Deslous-Paoli, J.M. (1991). Oyster culture in European countries. In: Menzel, W. (Ed.) *Estuarine and marine bivalve mollusk culture*. Baton Rouge: CRC Press, pp. 154–190.
- Holt, R.D. (2009). Bringing the Hutchinsonian niche into the 21st century: ecological and evolutionary perspectives. *Proceedings of the National Academy of Sciences of the United States of America*, 106(Supplement 2), 19659–19665. <https://doi.org/10.1073/pnas.0905137106>
- Hutchinson, S. & Hawkins, L.E. (1992). Quantification of the physiological responses of the European flat oyster *Ostrea edulis* L. to temperature and salinity. *Journal of Molluscan Studies*, 58(2), 215–226. <https://doi.org/10.1093/mollus/58.2.215>
- Kamermaans, P., Walles, B., Kraan, M., Duren, L.A., Kleissen, F. & Have, T. M. (2018). Offshore wind farms as potential locations for flat oyster (*Ostrea edulis*) restoration in the Dutch North Sea. *Sustainability*, 10, 3942. <https://doi.org/10.3390/su10113942>
- Kaskela, A.M., Kotilainen, A.T., Alanen, U., Cooper, R., Green, S., Guinan, J., et al. (2019). Picking up the pieces—harmonising and collating seabed substrate data for European maritime areas. *Geosciences*, 9(2), 84. <https://doi.org/10.3390/geosciences9020084>
- Kearney, M. (2006). Habitat, environment and niche: what are we modelling? *Oikos*, 115(1), 186–191. <https://doi.org/10.1111/j.2006.0030-1299.14908.x>
- Kearney, M., Phillips, B.L., Tracy, C.R., Christian, K.A., Betts, G. & Porter, W.P. (2008). Modelling species distributions without using species distributions: the cane toad in Australia under current and future climates. *Ecography*, 31(4), 423–434. <https://doi.org/10.1111/j.0906-7590.2008.05457.x>
- Kearney, M. & Porter, W. (2009). Mechanistic niche modelling: combining physiological and spatial data to predict species' ranges. *Ecology Letters*, 12(4), 334–350. <https://doi.org/10.1111/j.1461-0248.2008.01277.x>
- Kearney, M., Simpson, S.J., Raubenheimer, D. & Helmuth, B. (2010). Modelling the ecological niche from functional traits. *Philosophical Transactions of the Royal Society, B: Biological Sciences*, 365(1557), 3469–3483. <https://doi.org/10.1098/rstb.2010.0034>
- Kooijman, S.A.L.M. (2010). *Dynamic energy budget theory for metabolic organisation*. Cambridge University Press.
- Korringa, P. (1957). Water temperature and breeding throughout the geographical range of *Ostrea edulis*. *Colloque international de biologie marine station biologique de Roscoff. L'Année Biologique*, 33(5–6), 1–17. <https://doi.org/10.1111/emr.12159>
- Kröncke, I., Reiss, H., Eggleton, J.D., Aldridge, J., Bergman, M.J.N., Cochrane, S. et al. (2011). Changes in North Sea macrofauna communities and species distribution between 1986 and 2000. *Estuarine, Coastal and Shelf Science*, 94(1), 1–15. <https://doi.org/10.1016/j.ecss.2011.04.008>
- Laing, I., Walker, P. & Areal, F. (2005). A feasibility study of native oyster (*Ostrea edulis*) stock regeneration in the United Kingdom. *CARD Project FC1016*, CEFAS. p. 95.
- Launey, S., Ledu, C., Boudry, P., Bonhomme, F. & Naciri-Graven, Y. (2002). Geographic structure in the European flat oyster (*Ostrea edulis* L.) as revealed by microsatellite polymorphism. *Journal of Heredity*, 93(5), 331–351. <https://doi.org/10.1093/jhered/93.5.331>
- Lavaud, R., La Peyre, M.K., Casas, S.M., Bacher, C. & La Peyre, J.F. (2017). Integrating the effects of salinity on the physiology of the eastern oyster, *Crassostrea virginica*, in the northern Gulf of Mexico through a dynamic energy budget model. *Ecological Modelling*, 363, 221–233. <https://doi.org/10.1016/j.ecolmodel.2017.09.003>
- Lavaud, R., La Peyre, M.K., Justic, D. & La Peyre, J.F. (2021). Dynamic energy budget modelling to predict eastern oyster growth, reproduction, and mortality under river management and climate change scenarios. *Estuarine, Coastal and Shelf Science*, 251, 107188. <https://doi.org/10.1016/j.ecss.2021.107188>
- Lika, K., Kearney, M.R., Freitas, V., Veer, H.W., Meer, J. & Wijsman, J.W. (2011). The “covariation method” for estimating the parameters of the standard dynamic energy budget model I: philosophy and approach. *Journal of Sea Research*, 66(4), 270–277. <https://doi.org/10.1016/j.seares.2011.07.010>
- Lübbert, H.O. (1906). Die Einführung von Motor und Schernetz in die deutsche Segelfischerei. *Abhandlungen des Deutschen Seefischereivereins*, 8. <https://doi.org/10.1016/j.seares.2011.07.010>
- Maathuis, M.A.M., Coolen, J.W.P., van der Have, T. & Kamermaans, P. (2020). Factors determining the timing of swarming of European flat oyster (*Ostrea edulis* L.) larvae in the Dutch Delta area: implications for flat oyster restoration. *Journal of Sea Research*, 156, 101828. <https://doi.org/10.1016/j.seares.2019.101828>
- McGonigle, C., Jordan, C. & Geddis, K. (2016). *Native oyster spawning assessment lough Foyle summer 2016*. The Loughs Agency (Foyle Carlingford and Irish Lights Commission).
- Mesel, I., Kapasakali, D., Kerckhof, F., Vigin, L., Lacroix, G. & Barbut, L. (2018). *Ostrea edulis* restoration in the Belgian part of the North Sea: feasibility study. *Royal Belgian Institute of Natural Sciences, Operational Directorate Natural Environment, Marine Ecology and Management*, 89.
- Olsen, O.T. (1883). *The Piscatorial Atlas of the North Sea*, English and St. George's Channels, Illustrating the Fishing Ports, Boats, Gear, Species of Fish (How, Where, and When Caught), and Other Information Concerning Fish and Fisheries.
- Orton, J.H. (1937). *Oyster biology and oyster culture. The Buckland lectures for 1935*. London: Edward Arnold and Co.
- Palmer, S.C.J., Gernez, P.M., Thomas, Y., Simis, S., Miller, P.I., Glize, P. et al. (2020). Remote sensing-driven Pacific oyster (*Crassostrea gigas*) growth modeling to inform offshore aquaculture site selection. *Frontiers in Marine Science*, 6, 802. <https://doi.org/10.3389/fmars.2019.00802>
- Perry, F. & Jackson, A. (2017). *Ostrea edulis* Native oyster. In: Tyler-Walters, H. & Hiscock, K. (Eds.) *Marine life information network: biology and sensitivity key information reviews, [on-line]*. Plymouth: Marine Biological Association of the United Kingdom. <https://doi.org/10.17031/marlinsp.1146.2>
- Pogoda, B., Boudry, P., Bromley, C., Cameron, T.C., Colsoul, B., Donnan, D. et al. (2020a). NORA moving forward: developing an oyster restoration network in Europe to support the Berlin oyster recommendation. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 30(11), 2031–2037. <https://doi.org/10.1002/aqc.3447>
- Pogoda, B., Brown, J., Hancock, B. & Nordheim, H. (2017). Berlin Oyster Recommendation on the Future of Native Oyster Restoration in

- Europe-Part I, Preface and Recommendations. Native oyster restoration in Europe-current activities and future perspectives.
- Pogoda, B., Brown, J., Hancock, B., Preston, J., Pouvreau, S., Kamermans, P. et al. (2019). The native oyster restoration Alliance (NORA) and the Berlin oyster recommendation: bringing back a key ecosystem engineer by developing and supporting best practice in Europe. *Aquatic Living Resources*, 32, 13. <https://doi.org/10.1051/alr/2019012>
- Pogoda, B., Buck, B.H. & Hagen, W. (2011). Growth performance and condition of oysters (*Crassostrea gigas* and *Ostrea edulis*) farmed in an offshore environment (North Sea, Germany). *Aquaculture*, 319(3), 484–492. <https://doi.org/10.1016/j.aquaculture.2011.07.017>
- Pogoda, B., Merk, V., Colsoul, B., Hausen, T., Peter, C., Pesch, R. et al. (2020b). Site selection for biogenic reef restoration in offshore environments: the Natura 2000 area Borkum reef ground as a case study for native oyster restoration. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 30(11), 2163–2179. <https://doi.org/10.1002/aqc.3405>
- Preston, J., Fabra, M., Helmer, L., Johnson, E., Harris-Scott, E. & Hendy, I.W. (2020). Interactions of larval dynamics and substrate preference have ecological significance for benthic biodiversity and *Ostrea edulis* Linnaeus, 1758 in the presence of *Crepidula fornicata*. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 30(11), 2133–2149. <https://doi.org/10.1002/aqc.3446>
- Robinson, N.M., Nelson, W.A., Costello, M.J., Sutherland, J.E. & Lundquist, C.J. (2017). A systematic review of marine-based species distribution models (SDMs) with recommendations for best practice. *Frontiers in Marine Science*, 4, 421. <https://doi.org/10.3389/fmars.2017.00421>
- Rodriguez-Perez, A. (2020). Informing the conservation and restoration of a keystone species: the larval behaviour of the European oyster *Ostrea edulis*. (Doctoral dissertation, University of St Andrews).
- Schulte, D.M., Burke, R.P. & Lipcius, R.N. (2009). Unprecedented restoration of a native oyster metapopulation. *Science*, 325(5944), 1124–1128. <https://doi.org/10.1126/science.1176516>
- Sillero, N., Arenas-Castro, S., Enriquez-Urzelai, U., Vale, C.G., Sousa-Guedes, D., Martínez-Freiría, F. et al. (2021). Want to model a species niche? A step-by-step guideline on correlative ecological niche modelling. *Ecological Modelling*, 456, 109671. <https://doi.org/10.1016/j.ecolmodel.2021.109671>
- Smaal, A., Kamermans, P., Kleissen, F., Duren, L. & Have, T. (2017). Flat oysters on offshore wind farms: opportunities for the development of flat oyster populations on existing and planned wind farms in the Dutch section of the North Sea. (C052/17).
- Smaal, A.C., Kamermans, P., Have, T.M., Engelsma, M.Y. & Sas, H. (2015). Feasibility of Flat Oyster (*Ostrea edulis* L.) restoration in the Dutch part of the North Sea. Report/IMARES Wageningen UR No. no. C028/15.
- Smyth, D. & Roberts, D. (2010). The European oyster (*Ostrea edulis*) and its epibiotic succession. *Hydrobiologia*, 655(1), 25–36. <https://doi.org/10.1007/s10750-010-0401-x>
- SS, H. (1889). Sixth annual report of the fishery Board for Scotland. *Nature*, 39, 498–499. <https://doi.org/10.1038/039498a0>
- Stechele, B., Maar, M., Wijsman, J., Van der Zande, D., Degraer, S., Bossier, P. et al. (2022). Comparing life history traits and tolerance to changing environments of two oyster species (*Ostrea edulis* and *Crassostrea gigas*) through dynamic energy budget theory. *Conservation Physiology*, 10(1), coac034. <https://doi.org/10.1093/conphys/coac034>
- Teal, L.R., Marras, S., Peck, M.A. & Domenici, P. (2018). Physiology-based modelling approaches to characterize fish habitat suitability: their usefulness and limitations. *Estuarine, Coastal and Shelf Science*, 201, 56–63. <https://doi.org/10.1016/j.ecss.2015.11.014>
- Thomas, Y., Mazurié, J., Alunno-Bruscia, M., Bacher, C., Bouget, J.-F., Gohin, F. et al. (2011). Modelling spatio-temporal variability of *Mytilus edulis* (L.) growth by forcing a dynamic energy budget model with satellite-derived environmental data. *Journal of Sea Research*, 66(4), 308–317. <https://doi.org/10.1016/j.seares.2011.04.015>
- Thurstan, R.H., Hawkins, J.P., Raby, L. & Roberts, C.M. (2013). Oyster (*Ostrea edulis*) extirpation and ecosystem transformation in the Firth of Forth, Scotland. *Journal for Nature Conservation*, 21(5), 253–261. <https://doi.org/10.1016/j.jnc.2013.01.004>
- van Duren, L., Kamermans, P. & Kleissen, F. (2022). Suitability for the development of flat oyster populations in new offshore wind farm zones and two search areas for restoration projects in the Dutch section of the North Sea. Deltares, Wageningen Marine Research.
- Vaquer-Sunyer, R. & Duarte, C.M. (2008). Thresholds of hypoxia for marine biodiversity. *Proceedings of the National Academy of Sciences of the United States of America*, 105(40), 15452–15457. <https://doi.org/10.1073/pnas.0803833105>
- Walne, P.R. (1961). Observations on the mortality of *Ostrea Edulis*. *Journal of the Marine Biological Association of the United Kingdom*, 41(1), 113–122. <https://doi.org/10.1017/S0025315400001569>
- Zwerschke, N., Kochmann, J., Ashton, E.C., Crowe, T.P., Roberts, D. & O'Connor, N.E. (2018). Co-occurrence of native *Ostrea edulis* and non-native *Crassostrea gigas* revealed by monitoring of intertidal oyster populations. *Journal of the Marine Biological Association of the United Kingdom*, 98(8), 2029–2038. <https://doi.org/10.1017/S0025315417001448>

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Stechele, B., Hughes, A., Degraer, S., Bossier, P. & Nevejan, N. (2023). Northern Europe's suitability for offshore European flat oyster (*Ostrea edulis*) habitat restoration: A mechanistic niche modelling approach. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 33(7), 696–707. <https://doi.org/10.1002/aqc.3947>