

Two new abyssal Harpiniinae Barnard & Drummond, 1978 (Amphipoda, Phoxocephalidae) from the Clarion-Clipperton Zone

Luiz F. Andrade¹, Anna M. Jażdżewska¹¹ Department of Invertebrate Zoology and Hydrobiology, University of Lodz, Banacha 12/16, 90-237, Lodz, PolandCorresponding author: Luiz F. Andrade (luiz.felipe.de.andrade@biol.uni.lodz.pl)

Abstract

Two new species of the phoxocephalid subfamily Harpiniinae are described from the easternmost sector of the Clarion-Clipperton Zone. The material examined was collected with epibenthic sledges during the projects ABYSSLINE-2, MANGAN 2016 and MANGAN 2018 from depths ranging from 4133 to 4359 metres. *Harpinia lobata* **sp. nov.** is mainly characterized by: head anteroventral corner with a small projection; gnathopod 1 coxa anteroventrally produced; pereopod 6 basis with posterior proximal expanded lobe; and epimeron 3 posteroventral corner rounded. *Harpiniopsis pedro* **sp. nov.** can be diagnosed by: head anteroventral corner with a large projection; mandible palp article 2 shorter than article 3; gnathopods 1–2 palm defined by a subacute hump; pereopod 6 basis with a posterior distal lobe reaching the apex of ischium; and epimeron 3 posteroventral corner produced as a slightly curved spine. Here, we provide the deepest record of the known *Harpinia* species and the first *Harpiniopsis* record from the Pacific abyssal plain.

Key words: Amphipods, barcoding, COI, deep sea, *Harpinia*, *Harpiniopsis*, new record, new species, Pacific Ocean, systematics



Academic editor: Tammy Horton

Received: 31 October 2024

Accepted: 24 January 2025

Published: 24 March 2026

ZooBank: <https://zoobank.org/94C0C5E1-C6E4-4579-BEEB-9F97E4B61CEF>

Citation: Andrade LF, Jażdżewska AM (2026) Two new abyssal Harpiniinae Barnard & Drummond, 1978 (Amphipoda, Phoxocephalidae) from the Clarion-Clipperton Zone. In: Jażdżewska AM (Ed.) New deep-sea Amphipoda from Clarion-Clipperton Zone. ZooKeys 1274: 103–118. <https://doi.org/10.3897/zookeys.1274.140727>

Copyright: ©

Luiz F. Andrade & Anna M. Jażdżewska.
This is an open access article distributed under
terms of the Creative Commons Attribution
License (Attribution 4.0 International – CC BY 4.0).

Introduction

The family Phoxocephalidae G.O. Sars, 1891 comprises primarily benthic species that constitute abundant and diverse components of both shallow-water and deep-sea marine assemblages (Barnard and Drummond 1978; Knox et al. 2012; Jażdżewska 2015; Brix et al. 2018). Study of the benthic fauna of the Clarion-Clipperton Zone (CCZ), a region in the central east Pacific characterized by the presence of manganese nodules, revealed Phoxocephalidae to be highly abundant and consisting of species new to science (Jażdżewska et al. 2025).

Phoxocephalids are divided into two subfamilies, Phoxocephalinae G.O. Sars, 1891 and Harpiniinae Barnard & Drummond, 1978, mainly differentiated by the pereopod 5 basis which is broad in the former and slender in the latter (De Broyer et al. 2007; Alonso de Pina et al. 2008; Andrade and Senna 2019). Harpiniinae comprises 11 genera and is widely distributed geographically and bathymetrically (Barnard and Drummond 1978). Among the harpiniins, *Harpinia* Boeck, 1876 and *Harpiniopsis* Stephensen, 1925 are the most speciose, consisting of 26 and 30 species, respectively (Horton et al. 2024).

Historically, *Harpinia* species have primarily been documented predominantly in the Northern Hemisphere. However, recent studies have extended the range of this genus to the Southern Hemisphere (Andrade et al. 2023). In contrast, *Harpiniopsis* has a more cosmopolitan distribution, with species reported from both hemispheres. Both genera are very similar morphologically, leading to a complicated taxonomic distinction. This has resulted in many taxonomic revisions in the past since male specimens, which are rarely sampled, are crucial for taxonomic differentiation between these genera (Karaman 1993; King et al. 2004; Andrade et al. 2023).

During the examination of phoxocephalid specimens collected in the Clarion-Clipperton Zone, in the North Pacific during the ABYSSLINE-2, MANGAN 2016, and MANGAN 2018 expeditions, two new species of Harpiniinae were identified. One of them is assigned to the genus *Harpinia* and the other to *Harpiniopsis*. The classification of the *Harpinia* species was more precise due to the presence of a male individual showing a short flagellum of antenna 2 and well-developed calynophores forming dense tufts on the peduncle articles of antenna 1. Although males were not available for *Harpiniopsis*, based on Barnard and Karaman (1991), subtle morphological characters of antenna 2 and uropods 1–3 combined make the species pertinent to this genus.

Material and methods

The material for the present study was sampled in the central east Pacific, specifically in the easternmost sector of the Clarion-Clipperton Zone. The material was collected with an epibenthic sledge (EBS) during three scientific deep-sea cruises: the ABYSSLINE-2 (ABYSSal baseLINE project) in 2015, MANGAN 2016, and MANGAN 2018. For details of gear deployment and sample processing see Jazdzewska and Horton (2026) and Jazdzewska et al. (2025).

Individuals were initially examined using a Nikon SMZ800 stereomicroscope. Appendage photographs were taken using a Nikon Eclipse Ci-L equipped with a Delta Optical DLT-Cam PRO 5MP. The habitus of the holotypes are presented as photographs obtained with a confocal laser scanning microscope (CLSM). The holotypes were stained in Congo red and acid fuchsin, temporarily mounted onto slides with glycerin and examined with a Leica TCS SPV equipped with a Leica DM5000 B upright microscope and three visible-light lasers (DPSS 10 mW 561 nm; HeNe 10 mW 633 nm; Ar 100 mW 458, 476, 488 and 514 nm), combined with the software LAS AF v. 2.2.1 (Leica Application Suite, Advanced Fluorescence). A series of photographic stacks were obtained, collecting overlapping optical sections throughout the whole preparation (Michels and Büntzow 2010; Kamanli et al. 2017).

Chosen specimens were dissected and mounted on permanent slides using polyvinyl-lactophenol containing lignin pink. Photographs from the microscope were used as the basis for line drawings, which were inked with CorelDRAW 2018.

In the descriptions and figures the following abbreviations were used: Hb = habitus; A1, 2 = antenna 1, 2; Ep1–3 = epimera 1–3; Hd = head; L/R Md = left/right mandible; Mx1, 2 = maxilla 1, 2; Mxp = maxilliped; c4 = coxa 4; G1, 2 = gnathopod 1, 2; P3–7 = pereopod 3–7; U1–3 = uropod 1–3; T = telson.

The registered type material is deposited in the Senckenberg Museum (SMF; Frankfurt, Germany).

All individuals were subjected to cytochrome c oxidase subunit I gene (COI) barcoding prior to the identification of the species. The molecular procedures are described in Jazdzewska et al. (2025). All sequences were deposited in GenBank with the accession numbers: [PQ734223](#), [PQ734298](#), [PQ734351](#), [PQ734390](#), [PQ734439](#), [PQ734492](#) and [PQ734519](#). The relevant voucher information, taxonomic classifications and sequences are deposited in the data set “DS-AMPHICCZ” in the Barcode of Life Data System (BOLD) (<https://doi.org/10.5883/DS-AMPHICCZ>) (www.boldsystems.org) (Ratnasingham and Hebert 2007).

Results

Taxonomy

Family Phoxocephalidae G.O. Sars, 1891

Subfamily Harpiniinae Barnard & Drummond, 1978

Genus *Harpinia* Boeck, 1876

***Harpinia lobata* Andrade & Jazdzewska, sp. nov.**

<https://zoobank.org/3C74FA4D-A905-47F7-8491-C269FD3D6352>

Figs 1–4

Material examined. Holotype: PACIFIC OCEAN • 1 ♀, 6.5 mm; Clarion-Clipperton Zone, 12°21.6'N, 116°42'W; depth 4170 m; 10/03/2015; UKSR-1 exploration contract area, R/V *Thompson*, ABYSSLINE-2, Station AB2-EB09, epibenthic sledge; dissected and drawn; SMF 63349, COI ([PQ734223](#)). **Paratype:** PACIFIC OCEAN • 1 ♂; Clarion-Clipperton Zone, 12°21.6'N, 116°42'W; depth 4170 m depth; 10/03/2015; UKSR-1 exploration contract area; R/V *Thompson*, ABYSSLINE-2, Station AB2-EB09, epibenthic sledge; dissected and drawn; SMF 63348, COI ([PQ734351](#)).

Type locality. Abyssal Pacific Ocean, Clarion-Clipperton Zone, 12°21.6'N, 116°42'W, depth 4170 m.

Diagnosis. **Head** anteroventral corner with a small subacute projection. **Antenna 2** peduncle article 4 with two distal facial rows of long setae. **Mandible** right *lacinia mobilis* with four spines, left *lacinia mobilis* apically multicuspidate, palp article 2 stout. **Gnathopod 1** coxa anteroventrally produced. **Pereopod 6** basis posterior margin with a proximal expanded lobe. **Pereopod 7** basis posteroventral margin expanded and weakly serrate. **Epimeron 3** posteroventral corner rounded, without any spine or projection. **Telson** deeply cleft, with one mid-dorsal plumose seta present on each side.

Description. Based on the female holotype SMF 63349.

Habitus as in Fig. 1.

Head (Fig. 2): **Head** anteroventral corner with a small subacute projection. **Antenna 1** peduncle article 1 slightly longer than wide, ventral margin with few distal setae, article 2 with a facial row of long setae, article 3 short and subquadrate; primary flagellum eight-articulate; accessory flagellum seven-articulate. **Antenna 2** peduncle article 4 slightly longer than wide, with two distal facial rows of long setae, being one extending dorsally and the other ventrally, article 5 with a lower facial row of long setae; flagellum damaged, at least three-articulate. **Mandible** incisor with a main spine followed by a cutting surface (left) – with two spines (right), *lacinia mobilis* apically multicuspidate (left) – with



Figure 1. *Harpinia lobata* sp. nov., female holotype SMF 63349. Scale bars: 0.5 mm (Hb).

four spines (right), molar as a hump with three robust setae, accessory setal row with four robust setae; palp three-articulate, article 2 stout, article 3 apex oblique and setose, as long as article 2. **Maxilla 1** inner plate with three apical plumose setae; outer plate with 11 apical robust setae; palp two-articulate, article 2 apex with six setae. **Maxilla 2** inner plate subequal to outer, with apical plumose setae; outer plate with apical simple setae. **Maxilliped** inner plate with plumose setae and one apical robust seta; outer plate with mesial robust setae increasing in length towards the apex; palp articles 2–3 weakly setose, article 4 about half the length of article 3, bearing a long, embedded nail.

Pereon (Fig. 3): Gnathopod 1 coxa anteroventrally produced, with sparse ventral pappose/simples setae; basis to carpus ordinary, weakly setose; propodus 1.9× longer than wide, posterior margin with a robust seta near palmar corner, palm acute, defined by an excavation forming a medium palmar hump; dactylus as long as palm. **Gnathopod 2** coxa with ventral pappose setae; carpus shorter than in gnathopod 1; propodus very similar to gnathopod 1 but slightly broader. **Pereopod 3** coxa with ventral pappose setae; merus 1.8× longer than wide, ventral margin with three sets of simple/pappose setae; propodus distally setose, 3.8× longer than wide; dactylus about 75% the length of propodus. **Pereopod 4** coxa posteroventral lobe strongly expanded, posterodorsal margin excavate, ventral margin with pappose setae; other articles very similar to pereopod 3. **Pereopod 5** coxa bilobate, posterior lobe with pappose setae; basis 3.1× longer than wide; merus weakly setose; carpus and propodus both margins setose with plumose setae; dactylus as long as

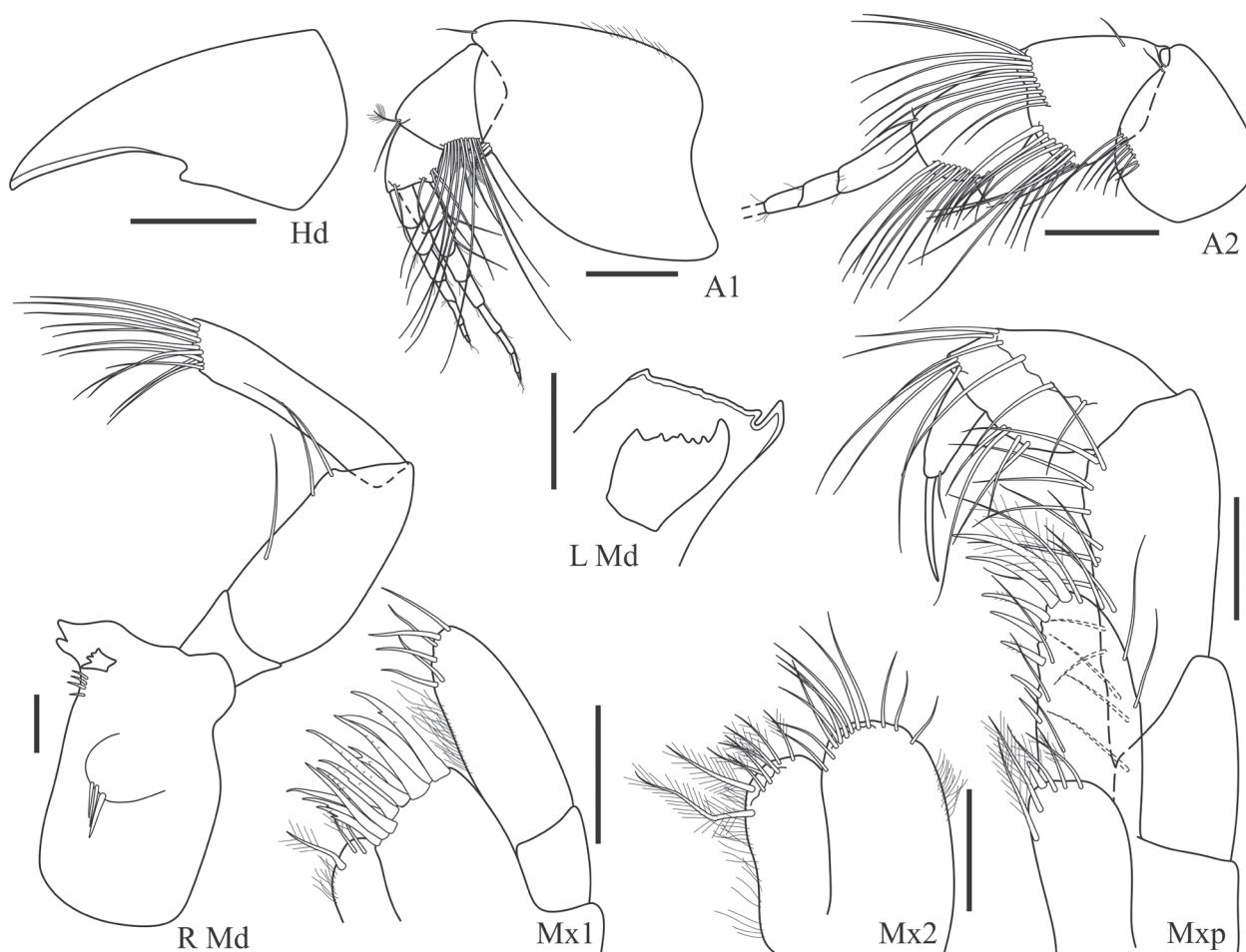


Figure 2. *Harpinia lobata* sp. nov., female holotype SMF 63349. Scale bars: 0.5 mm (Hd); 0.3 mm (A1–2); 0.1 mm (L Md, R Md, Mxp, Mx1–2).

propodus. **Pereopod 6** coxa posterior margin weakly expanded, with one seta; basis posterior margin with a proximal expanded lobe; carpus anterior margin with sets of robust setae; dactylus long, slightly shorter than propodus. **Pereopod 7** coxa subtriangular, posterior margin with four setae; basis slightly longer than wide, posteroventral margin expanded and weakly serrate; dactylus as long as carpus and propodus combined.

Pleon (Fig. 4): Epimeron 1 without setae. **Epimera 2–3** ventral margin with simple setae, posteroventral corner rounded, without any spine or projection. **Uropod 1** peduncle 2.6× longer than wide, with dorsal setae; rami subequal in length, without apical nail, bearing dorsal robust setae. **Telson** deeply cleft, as long as wide, with one mid-dorsal and two apical plumose setae.

Sexually dimorphic characters (Fig. 4). Based on the male paratype SMF 63348.

Antenna 1 with well-developed callynophores forming dense tufts on peduncle articles 2–3.

Pereopod 6 basis posterior margin with a proximal expanded lobe, posterior margin with a dense row of setules. **Pereopod 7** merus broad.

Uropod 1 peduncle 4.4× longer than wide, bearing simple dorsal setae and robust setae distally; rami subequal in length, with dorsal setae; outer ramus with an apical embedded nail. **Uropod 2** peduncle 3.3× longer than wide,

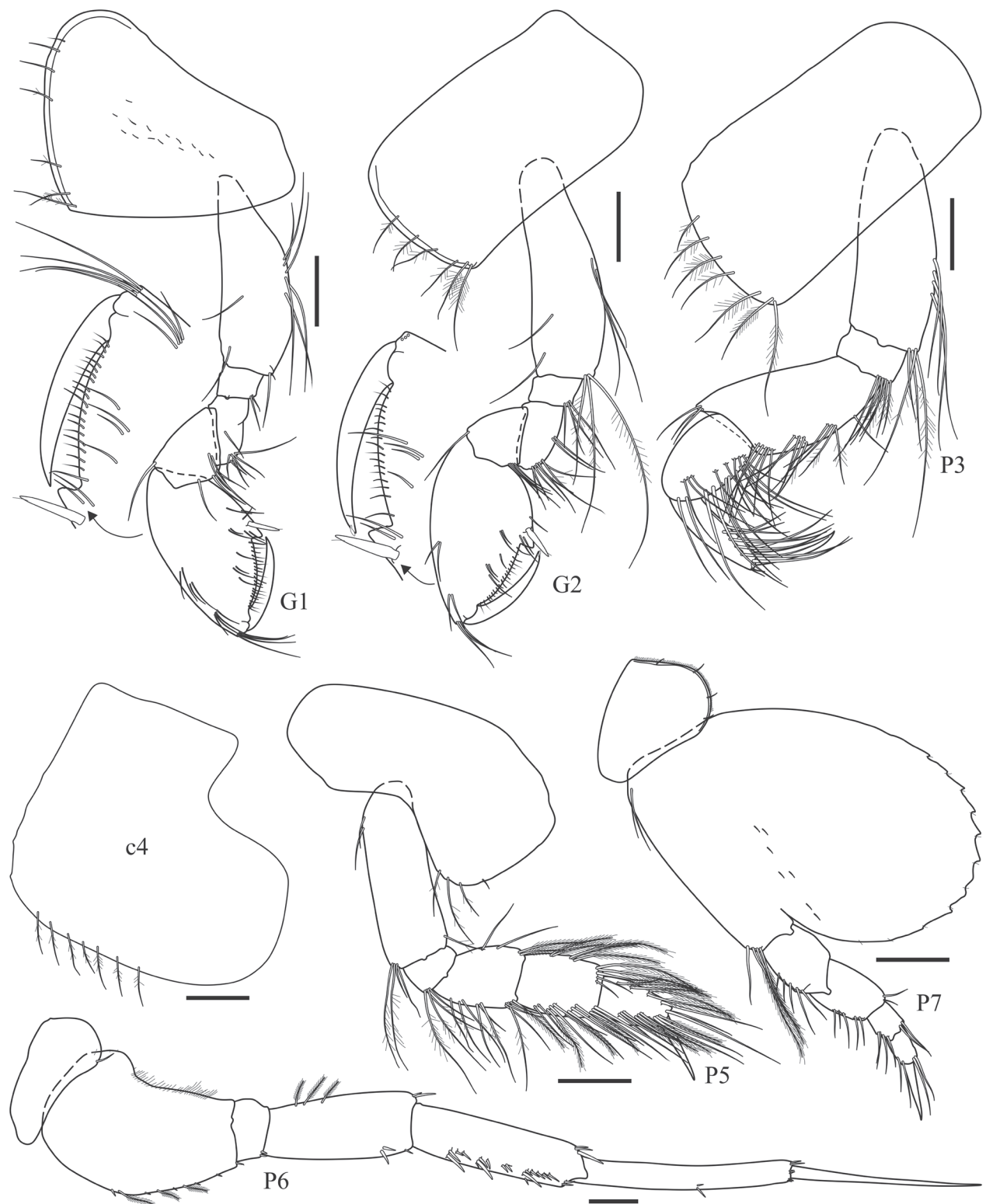


Figure 3. *Harpinia lobata* sp. nov., female holotype SMF 63349. Scale bars: 0.3 mm (G1–2, P3–7).

bearing dorsal setae; rami subequal in length, with dorsal setae. **Uropod 3** peduncle with five distal robust setae; rami subequal in length; outer ramus article 1 with two distal plumose setae, article 2 with one apical plumose seta; inner ramus slightly longer than article 1 of outer ramus, with two distal plumose setae.

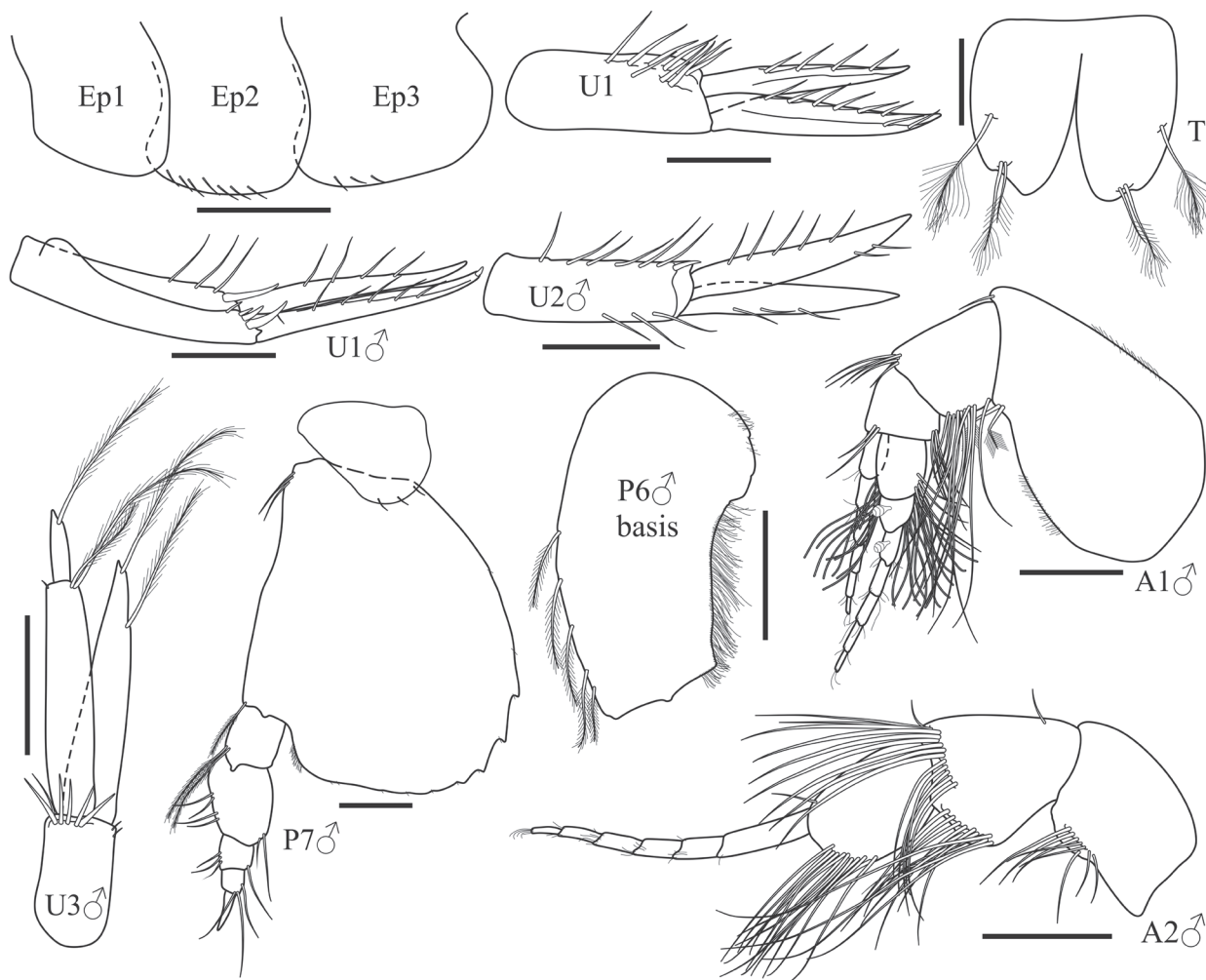


Figure 4. *Harpinia lobata* sp. nov., female holotype SMF 63349, male paratype SMF 63348. Scale bars: 0.5 mm (Ep1–3); 0.3 mm (A1–2, U1–3, P6–7); 0.1 mm (T).

Molecular identification. Following the definition given by Pleijel et al. (2008), the sequence of the female holotype of *Harpinia lobata* sp. nov. (SMF 63349, GenBank accession number [PQ734223](https://doi.org/10.26434/chemrxiv-2023-pq734)) is designated as a hologenophore of all obtained sequences. The sequence of the paratype of the species is deposited in GenBank with the following accession number: [PQ734351](https://doi.org/10.26434/chemrxiv-2023-pq734). The species has also received a Barcode Index Number from Barcode of Life Data Systems: [BOLD:AEA9473](https://doi.org/10.26434/chemrxiv-2023-pq734) (<https://doi.org/10.5883/BOLD:AEA9473>).

Distribution. Only known from the type locality.

Etymology. The specific epithet *lobata*, from the Latin “lobatus” (lobed), is given due to the prominent posterior lobe on the pereopod 6 basis.

Remarks. *Harpinia lobata* sp. nov. is morphologically similar to *Harpinia curti-pes* Stephensen, 1925 and *Harpinia truncata* G.O. Sars, 1891 as they all present the following: epimeron 3 posteriorly rounded, without any projection or spine; pereopod 6 dactylus shorter than propodus; and pereopod 7 basis anteroventral corner not produced, posteroventral margin crenulate/serrate, without large spines. However, the new species can be mainly distinguished from them due to the following: gnathopod 1 coxa anteroventrally produced (vs. not produced in both species); and pereopod 6 basis posterior margin with a proximal expanded lobe (vs. without proximal lobe in both species).

Harpiniopsis pedro Andrade & Jazdzewska, sp. nov.

<https://zoobank.org/F117A725-7D8F-4510-B955-E2E0B272F269>

Figs 5–8

Material examined. Holotype: PACIFIC OCEAN • 1 ♀, 5.7 mm; Clarion-Clipperton Zone; 11°47.862'N, 117°30.639'W – 11°47.152'N, 117°29.490'W; depth 4359–4356 m; 9/05/2016; BGR exploration contract area, R/V *Kilo Moana*, MANGAN 2016, Station Ma 16-95, epibenthic sledge; dissected and drawn; SMF 63354, COI (PQ734298). **Paratype:** PACIFIC OCEAN • 2 ♀; Clarion-Clipperton Zone, BGR exploration contract area; 11°47.862'N, 117°30.639'W – 11°47.152'N, 117°29.490'W; depth 4359–4356 m; 9/05/2016; BGR exploration contract area, R/V *Kilo Moana*, MANGAN 2016; Station Ma 16-95, epibenthic sledge; SMF 63352, COI (PQ734439), SMF 63353, COI (PQ734519). • 1 ♀; Clarion-Clipperton Zone; 11°47.436'N, 117°32.213'W – 11°47.677'N, 117°30.910'W; depth 4351–4352 m; 9/05/2018; BGR exploration contract area, R/V *Sonne*; Station SO 262-155, epibenthic sledge; SMF 63350, COI (PQ734390). • 1 ♀; Clarion-Clipperton Zone; 11°49.654'N, 117°00.299'W – 11°49.902'N, 116°59.174'W; depth 4133–4143 m; 1/05/2016; BGR exploration contract area, R/V *Kilo Moana*, MANGAN 2016; Station Ma 16-28, epibenthic sledge; SMF 63351, COI (PQ734492).

Type locality. Abyssal Pacific Ocean, Clarion-Clipperton Zone, 11°47.862'N, 117°30.639'W – 11°47.152'N, 117°29.490'W, depth 4359–4356 m.

Diagnosis. **Head** anteroventral corner with a large acute projection. **Mandible** right lacinia mobilis bifid, left lacinia mobilis with four teeth, palp article 2 longer than article 3, with an inner medial long pappose setae. **Gnathopods 1–2** palm defined by a subacute hump. **Pereopod 6** basis posterior margin with a distal lobe reaching the distal apex of ischium. **Pereopod 7** posteroventral margin expanded and serrate. **Epimeron 3** posteroventral corner produced as a slightly curved spine. **Uropod 2** rami naked. **Telson** deeply cleft, with two mid-dorsal plumose setae present on each side.

Description. Based on the female holotype (SMF 63354).

Habitus as in Fig. 5.

Head (Fig. 6): Head anteroventral corner with a large acute projection. **Antenna 1** peduncle article 1 about 1.3× longer than wide, ventral margin with three distal brush setae, article 2 ventral margin with row of long pappose setae; article 3 short and subquadrate; primary flagellum 6-articulate; accessory flagellum 3-articulate. **Antenna 2** peduncle article 1 not ensiform, article 4 setose, with two facial rows of setae dorso-distally, article 5 ventral margin with a row of long setae; flagellum 4-articulate. **Mandible** incisor with six spines (left) – with three spines (right), lacinia mobilis with four teeth (left) – bifid (right), molar as a hump with one plumose and four robust setae; palp 3-articulate, article 2 with an inner medial long pappose setae, article 3 apex oblique and setose, 0.8× the length of article 2. **Maxilla 1** inner plate with three apical setae; outer plate with 10 apical robust setae; palp 2-articulate, article 2 apex setose. **Maxilla 2** inner plate subequal to outer, with apical plumose setae; outer plate with lateral and apical simple setae. **Maxilliped** inner plate with plumose setae and one apical robust seta; outer plate with mesial robust setae increasing in length towards the apex; palp articles 2–3 weakly setose, article 4 about 0.7× the length of article 3, bearing a medium embedded nail.



Figure 5. *Harpiniopsis pedro* sp. nov., female holotype SMF 63354. Scale bars: 0.5 mm (Hb).

Pereon (Fig. 7): **Gnathopod 1** coxa with ventral pappose/simple setae; basis to carpus ordinary, weakly setose; propodus 2× longer than wide, posterior margin with a robust seta near palmar corner, palm acute, defined by a subacute hump; dactylus as long as palm. **Gnathopod 2** coxa with ventral pappose/simple setae; carpus shorter than in gnathopod 1; propodus very similar to gnathopod 1 but with palm defined by an acute hump. **Pereopod 3** coxa with ventral pappose setae; merus 2.2× longer than wide, ventral margin moderately setose with simple and plumose setae; propodus with setae, 3.3× longer than wide; dactylus about 85% the length of propodus. **Pereopod 4** coxa posteroventral lobe strongly expanded, posterodorsal margin excavate, ventral margin with pappose setae; other articles very similar to pereopod 3. **Pereopod 5** coxa bilobate, posterior lobe with pappose setae; merus to propodus anterior margin with long pappose setae, posterior margin with plumose setae; dactylus 80% the length of propodus. **Pereopod 6** coxa subtriangular, without setae; basis posterior margin with a distal lobe reaching the distal apex of ischium, anterior margin with sets of plumose setae; carpus anterior margin with sets of robust setae; dactylus long, 1.2× longer than propodus. **Pereopod 7** coxa subtriangular; basis 1.2× longer than wide, posteroventral margin expanded and serrate; dactylus as long as carpus and propodus combined.

Pleon (Fig. 8): **Epimeron 1** without setae. **Epimeron 2** ventral margin with simple setae. **Epimeron 3** posteroventral corner produced as a slightly curved spine. **Uropod 1** peduncle 2.8× longer than wide; rami subequal in length, without apical nail,

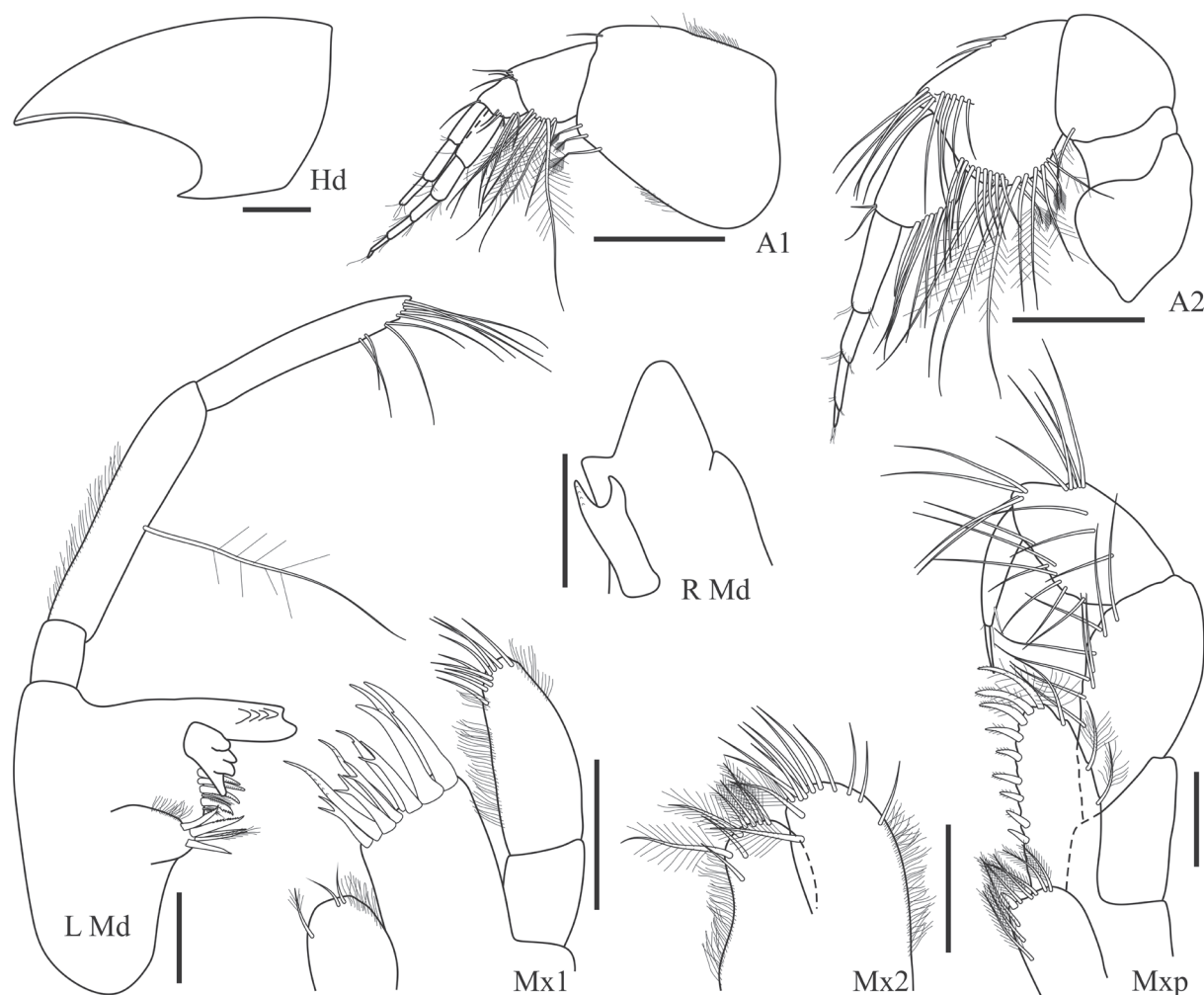


Figure 6. *Harpiniopsis pedro* sp. nov., female holotype SMF 63354. Scale bars: 0.1 mm (L Md, Mxp, Mx1–2); 0.3 mm (Hd, A1–2); 0.05 mm (R Md).

with spart dorsal setae. **Uropod 2** peduncle 2.3× longer than wide; rami subequal in length, naked, without apical nail. **Uropod 3** peduncle with five distal robust setae; rami equal in length, almost naked. **Telson** slightly wider than long, with two mid-dorsal and one apical plumose seta.

Molecular identification. Following the definition given by Pleijel et al. (2008), the sequence of the female holotype of *Harpiniopsis pedro* sp. nov. (SMF 63354, GenBank accession number [PQ734298](#)) is designed as a hologenophore of all obtained sequences. The sequences of the paratypes of the species are deposited in GenBank with the following accession numbers: [PQ734390](#), [PQ734439](#), [PQ734492](#) and [PQ734519](#). The species has also received a Barcode Index Number from Barcode of Life Data Systems: [BOLD:AEB4266](#) (<https://doi.org/10.5883/BOLD:AEB4266>).

Distribution. Abyssal Pacific Ocean, Clarion-Clipperton Zone, 4133–4359 m.

Etymology. The specific epithet *pedro*, used as a noun in apposition, is given in honor of Prof. Dr Pedro Martínez Arbizu from the German Center for Marine Biodiversity Research (DZMB), Senckenberg am Meer, renowned marine biologist and a good friend of the junior author who kindly offered the CCZ amphipod collection to study and was first who suggested organizing taxonomic description workshop that resulted in multiple amphipod species descriptions.



Figure 7. *Harpiniopsis pedro* sp. nov., female holotype SMF 63354. Scale bars: 0.5 mm (G1–2, P3, P5–7); 0.3 mm (c4).

Remarks. *Harpiniopsis pedro* sp. nov. is morphologically similar to *Harpiniopsis australis* (J.L. Barnard, 1961) and *Harpiniopsis wandichia* (J.L. Barnard, 1962) as they present the following: head anteroventral corner with an acute projection; pereopod 7 basis with a similar posteroventral serrate arrangement, reaching the distal apex of merus; epimeron 3 posteroventral corner produced into a spine.

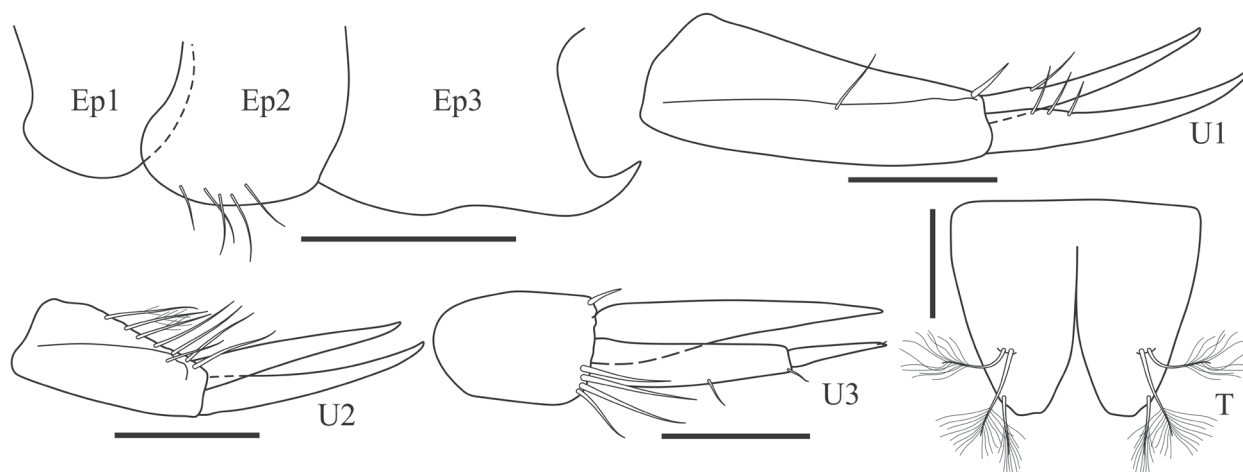


Figure 8. *Harpiniopsis pedro* sp. nov., female holotype SMF 63354. Scale bars: 0.5 mm (Ep1–3); 0.3 mm (U1–3); 0.1 mm (T).

However, the new species can be mainly distinguished from *Harpiniopsis australis* by the following (characters of *Harpiniopsis australis* between parentheses): pereopod 6 basis distal lobe reaching the distal apex of ischium (vs. reaching half of ischium), dactylus longer than propodus (vs. as long as propodus); pereopod 7 basis posteroventral projections broader about 1.2× as broad as long and bearing both subacute and blunt projections (vs. narrower about 0.8× as broad as long and bearing acute projections); epimeron 3 posteroventral spine slightly curved and of moderate size (vs. strongly curved and large); and telson with mid-dorsal setae (vs. without), apex truncate (vs. acute).

Also, *Harpiniopsis pedro* sp. nov. differs from *Harpiniopsis wandichia* by the following (characters of *Harpiniopsis wandichia* in parentheses): head anteroventral corner with large acute projection (vs. small); pereopod 5 merus to propodus anterior margin setose (vs. weakly setose); and telson with mid-dorsal setae (vs. without), apex truncate (vs. subacute).

Discussion

Until present, eight of the 26 known *Harpinia* species were reported from depths below 2000 m and they are: *Harpinia curtipes* Stephensen, 1925, *Harpinia dellavallei* Chevreux, 1910, *Harpinia mucronata* G.O. Sars, 1879, *Harpinia pico* Bellan-Santini, 2007, *Harpinia plumosa* (Krøyer, 1842), *Harpinia propinqua* G.O. Sars 1891, *Harpinia serrata* G.O. Sars 1879 and *Harpinia truncata*. All these species, excluding *Harpinia mucronata*, were recorded from the Atlantic Ocean or the Mediterranean Sea (Barnard 1962; Bellan-Santini 1985; Brandt 1997; Bellan-Santini 2007; Budaeva et al. 2008; Bergmann et al. 2011). *Harpinia mucronata* was reported by Blake et al. (2009) from the San Francisco Deep Ocean Disposal Site, California from a depth range of 2160–3142 m. However, considering this species being described and commonly recorded from the North Atlantic the identification shall be treated with caution. Moreover, none of the above-cited species was found at abyssal (below 4000 m) depths, so the present finding of *Harpinia lobata* sp. nov. at the depth of 4170 m provides the deepest record of the known *Harpinia* species.

In the case of *Harpiniopsis* as many as 12 species were reported from depths below 2000 m. Of them, *Harpiniopsis capensis* (J.L. Barnard, 1962) and *Harpiniopsis wandichia* were found in the Atlantic Ocean, with the first collected at abyssal

depths (4961–4983 m) (Barnard 1962). The group of the Pacific deep-sea *Harpiniopsis* species consists of: *Harpiniopsis emeryi* J.L. Barnard, 1960, *Harpiniopsis fulgens* J.L. Barnard, 1960, *Harpiniopsis galera* J.L. Barnard, 1960, *Harpiniopsis na-iadis* J.L. Barnard, 1960, *Harpiniopsis orientalis* (Bulycheva, 1936), *Harpiniopsis pacifica* (Bulycheva, 1936), *Harpiniopsis percellaris* J.L. Barnard, 1971, *Harpiniopsis profundis* J.L. Barnard, 1960, *Harpiniopsis spaercki* (Dahl, 1959), and *Harpiniopsis triplex* J.L. Barnard, 1971. Among these taxa, *Harpiniopsis spaercki* was recorded from the hadal zone (7335–7340 m in the Banda Sea Trench) (Kamenskaya 1981) whereas all remaining were not recorded below 3000 m (Barnard 1967, 1971; Blake et al. 2009; Golovan et al. 2013). The description of *Harpiniopsis pedro* sp. nov. acts as the first record of the genus from the Pacific abyssal plain.

Acknowledgements

The authors wish to thank the chief scientists, scientific teams and crews of the ABYSSLINE-2, MANGAN 2016 and MANGAN 2018 expeditions for their assistance in collecting the material.

We would like to extend a special thank you to Dr Terue Cristina Kihara (Ines Solutions, Germany) for her help during CLSM photograph preparations.

Thanks are also due to MSc Karolina Biniek (University of Lodz) and Jarosław Brodecki (University of Lodz) for their help in CLSM pictures' post-processing.

This is publication number 81 that uses data from the Senckenberg am Meer Confocal Laserscanning Microscope Facility (SGN-SaM-cLSM).

Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

Funding

This research was supported by the International Seabed Authority's Sustainable Seabed Knowledge Initiative: One Thousand Reasons Campaign (co-financed by the European Maritime and Fisheries Fund of the European Union, Project 101071214 – SSKI-I – EMFAF-2021-ISA-SSKI-IBA). The funding was given to A.M.J. to organize a taxonomic workshop devoted to describing new species from the CCZ (at the Department of Invertebrate Zoology and Hydrobiology, University of Lodz), as well as to ensure publication of newly described species.

The ABYSSLINE-2 cruise was funded by UK Seabed Resources Ltd. The MANGAN 2016 and MANGAN 2018 cruises were funded by the BGR (Bundesanstalt für Geowissenschaften und Rohstoffe – German Federal Institute for Geosciences and Natural Resources). Both institutions are thanked for providing the research material published here.

A.M.J. received financial support to visit the “Deutsches Zentrum für Marine Biodiversitätsforschung” (DZMB) in Wilhelmshaven from the German Academic Exchange Service (Deutscher Akademischer Austauschdienst - [DAAD]) within the program “Research Stays for University Academics and Scientists, 2019” (57440915) to study their collection of Pacific amphipods.

Author contributions

Conceptualization: AMJ. Funding acquisition: AMJ. Investigation: LFA, AMJ. Methodology: LFA, AMJ. Resources: AMJ. Writing – original draft: LFA. Writing – review and editing: AMJ.

Author ORCIDs

Luiz F. Andrade  <https://orcid.org/0000-0002-4868-737X>

Anna M. Jażdżewska  <https://orcid.org/0000-0003-2529-0641>

Data availability

All of the data that support the findings of this study are available in the main text.

References

- Alonso de Pina GM, Rauschert M, De Broyer C (2008) A catalogue of the Antarctic and sub-Antarctic Phoxocephalidae (Crustacea: Amphipoda: Gammaridea) with taxonomic, distribution and ecological data. *Zootaxa* 1752: 1–40. <https://doi.org/10.11646/zootaxa.1752.1.1>
- Andrade LF, Senna AR (2019) Two new species of *Cephalophoxoides* Gurjanova, 1977 (Crustacea: Amphipoda: Phoxocephalidae) from southeastern Brazil, with comments on the taxonomic status of the genus. *Zootaxa* 4712(4): 531–551. <https://doi.org/10.11646/zootaxa.4712.4.3>
- Andrade LF, Senna AR, Souza-Filho JF (2023) First record of the genus *Harpinia* Boeck, 1876 (Amphipoda: Phoxocephalidae: Harpiniinae) from the southern hemisphere, with description of three new species. *Marine Biology Research* 19(1): 59–80. <https://doi.org/10.1080/17451000.2023.2194059>
- Barnard JL (1960) The amphipod family Phoxocephalidae in the eastern Pacific Ocean with analyses of other species and notes for a revision of the family. *Allan Hancock Pacific Expeditions* 18(3): 175–368.
- Barnard JL (1962) South Atlantic abyssal amphipods collected by R.V. "Vema". *Abyssal Crustacea. Vema Research Series 1*. Columbia University Press, New York, 1–78.
- Barnard JL (1967) Bathyal and abyssal Gammaridean Amphipoda of Cedros Trench, Baja California. *United States National Museum Bulletin* 260: 1–205. <https://doi.org/10.5479/si.03629236.260.1>
- Barnard JL (1971) Gammaridean Amphipoda from a deep-sea transect off Oregon. *Smithsonian Contributions to Zoology* 61: 1–86. <https://doi.org/10.5479/si.00810282.61>
- Barnard JL, Drummond MM (1978) Gammaridean Amphipoda of Australia, part III: The Phoxocephalidae. *Smithsonian Contributions to Zoology* 245: 1–551. <https://doi.org/10.5479/si.00810282.103>
- Barnard JL, Karaman GS (1991) The families and genera of marine Amphipoda (except marine Gammaroids). *Records of the Australian Museum* 13(parts 1–2): 1–866. <https://doi.org/10.3853/j.0812-7387.13.1991.367>
- Bellan-Santini D (1985) Etude de la faune profonde de Méditerranée: les amphipodes des trois campagnes Polymède 1, Polymède 2 et Biomède 1. *Rapports et procès-verbaux des réunions de la Commission Internationale pour l'Exploitation Scientifique de la Mer Méditerranée* 29(5): 333–334.
- Bellan Santini D (2007) New amphipods of hydrothermal vent environments on the Mid Atlantic Ridge, Azores Triple junction zone. *Journal of Natural History* 41(9–12): 567–596. <https://doi.org/10.1080/00222930701262537>

- Bergmann M, Soltwedel T, Klages M (2011) The interannual variability of megafaunal assemblages in the Arctic deep sea: Preliminary results from the HAUSGARTEN observatory (79°N). *Deep Sea Research Part I* 58(6): 711–723. <https://doi.org/10.1016/j.dsr.2011.03.007>
- Blake JA, Maciolek NJ, Ota AY, Williams IP (2009) Long-term benthic infaunal monitoring at a deep-ocean dredged material disposal site off Northern California. *Deep Sea Research Part II* 56(19–20): 1775–1803. <https://doi.org/10.1016/j.dsr2.2009.05.021>
- Boeck A (1876) *De Skandinaviske og Arktiske Amphipoder*. A.W. Brogger, Christiania, 712 pp. <https://doi.org/10.5962/bhl.title.10131>
- Brandt A (1997) Biodiversity of peracarid crustaceans (Malacostraca) from the shelf down to the deep Arctic Ocean. *Biodiversity and Conservation* 6: 1533–1556. <https://doi.org/10.1023/A:1018318604032>
- Brix S, Lörz A-N, Jazdzewska A, Hughes L, Tandberg AH, Pabis K, Stransky B, Krapp-Schickel T, Sorbe J-C, Hendrycks E, Vader WJM, Frutos I, Horton T, Jazdzewski K, Peart R, Beermann J, Coleman CO, Buhl-Mortensen L, Corbari L, Havermans C, Tato R, Jimenez Campean A (2018) Amphipod family distributions around Iceland. *Zookeys* 731: 41–53 <https://doi.org/10.3897/zookeys.731.19854>
- Budaeva NE, Mokievsky VO, Soltwedel T, Gebruk AV (2008) Horizontal distribution patterns in Arctic deep-sea macrobenthic communities. *Deep Sea Research Part I*: 55(9): 1167–1178. <https://doi.org/10.1016/j.dsr.2008.05.002>
- Bulycheva A (1936) New species of Amphipoda from the Japan Sea. *Annals and Magazine of Natural History, Series 10*, 18: 242–256. <https://doi.org/10.1080/00222933608655190>
- Chevreaux E. (1910) Campagnes de la melita: les amphipodes d'Algérie et de Tunisie. *Zoologique de France* 23: 145–285. <https://doi.org/10.5962/bhl.title.15982>
- Dahl E (1959) Amphipoda from depths exceeding 6000 m. *Galathea Report* 1: 211–241.
- De Broyer C, Lowry JK, Jazdzewski K, Robert H (2007) Catalogue of the Gammaridean and Corophiidean Amphipoda (Crustacea) of the Southern Ocean, with distribution and ecological data. In: De Broyer C (Ed.) *Census of Antarctic Marine Life: Synopsis of the Amphipoda of the Southern Ocean*. Vol. 1. *Biologie* Vol 77 Supplement 1, *Bulletin de L'institut Royal Des Sciences naturelles de Belgique, Brussels*, 1–325. <https://doi.org/10.13140/rg.2.1.1388.6489>
- Golovan OA, Blazewicz-Paszkowycz M, Brandt A, Budnikova LL, Elsner NO, Ivin VV, Lavrenteva AV, Malyutina MV, Petryashov VV, Tzareva LA (2013) Diversity and distribution of peracarid crustaceans (Malacostraca) from the continental slope and the deep-sea basin of the Sea of Japan. *Deep-Sea Res. II* 86–87, 66–78. <https://doi.org/10.1016/j.dsr2.2012.08.002>
- Horton T, Lowry J, De Broyer C, Bellan-Santini D, Copilas-Ciocianu D, Corbari L, Costello MJ, Daneliya M, Dauvin J-C, Fišer C, Gasca R, Grabowski M, Guerra-García JM, Hendrycks E, Hughes L, Jaime D, Jazdzewski K, Kim Y-H, King R, Krapp-Schickel T, LeCroy S, Lörz A-N, Mamos T, Senna AR, Serejo C, Souza-Filho JF, Tandberg AH, Thomas JD, Thurston M, Vader W, Väinölä R, Valls Domedel G, Vonk R, White K, Zeidler W (2024) World Amphipoda Database. Harpiniinae Barnard & Drummond, 1978. <https://marinespecies.org/amphipoda/aphia.php?p=taxdetails&id=176844> [Accessed on 21.10.2024]
- Jazdzewska A (2015) Kuril-Kamchatka deep sea revisited insights into the amphipod abyssal fauna. *Deep-Sea Research II* 111: 294–300. <https://doi.org/10.1016/j.dsr2.2014.08.008>
- Jazdzewska AM, Biniek K, Martínez Arbizu P, Vink A (2025) Hidden behind the scene – high diversity, low connectivity of deep-sea Amphipoda in the polymetallic nodule

- fields in the central East Pacific. Biogeosciences, preprint. <https://doi.org/10.5194/egusphere-2025-1794>
- Jazdzewska AM, Horton T (2026) New deep-sea Amphipoda from the Clarion-Clipperton Zone: 24 new species described under the Sustainable Seabed Knowledge Initiative: One Thousand Reasons campaign. In: Jazdzewska AM (Ed.) New deep-sea Amphipoda from Clarion-Clipperton Zone. ZooKeys 1274: 1–16. <https://doi.org/10.3897/zookeys.1274.176711>
- Kamanli SA, Kihara TC, Ball AD, Morritt D, Clark PF (2017) A 3D imaging and visualization workflow, using confocal microscopy and advanced image processing for brachyuran crab larvae. Journal of Microscopy 266(3): 307–323. <https://doi.org/10.1111/jmi.12540>
- Kamenskaya OE (1981) Amphipody (Amphipoda, Crustacea) glubokovodnykh zhelobov zapadnoj chasti Tikhogo Okeana [The amphipods (Amphipoda, Crustacea) from the deep-sea trenches of the western part of the Pacific Ocean]. Trudy Instituta Okeanologii 115: 94–107.
- Karaman GS (1993) Family Phoxocephalidae: Harpinia. In: Ruffo S (Ed.) The Amphipoda of the Mediterranean, Part 3: Gammaridea, (Melpheippidae to Talitridae), Ingolfiellidea, Caprellidea. Memoires de l'Institut Oceanographique, Fondation Albert Ier, Prince de Monaco 13: 639–653.
- King RA, Myers AA, McGrath D (2004) A review of shallow-water Irish and British *Harpinia* Boeck (Crustacea: Amphipoda: Phoxocephalidae) species including the first detailed descriptions of the males of *Harpinia laevis* Sars and *Harpinia pectinata* Sars. Journal of Natural History 38: 1263–1286. <https://doi.org/10.1080/0022293031000079598>
- Knox MA, Hogg ID, Pilditch CA, Lörz AN, Hebert PDN, Steinke D (2012) Mitochondrial DNA (COI) analyses reveal that amphipod diversity is associated with environmental heterogeneity in deep-sea habitats. Molecular Ecology 21: 4885–4897. <https://doi.org/10.1111/j.1365-294X.2012.05729.x>
- Krøyer H (1842) Nye nordiske slægter og arter af amfipodernes orden, henhørende til familien Gammarina. (Forelobigt uddrag af et større arbejde). Naturhistorisk Tidsskrift Ser. I 4: 141–166.
- Michels J, Büntzow M (2010) Assessment of Congo red as a fluorescence marker for the exoskeleton of small crustaceans and the cuticle of polychaetes. Journal of Microscopy 238(2): 95–101. <https://doi.org/10.1111/j.1365-2818.2009.03360.x>
- Pleijel F, Jondelius U, Norlinder E, Nygren A, Oxelman B, Schander C, Sundberg P, Thollesson M (2008) Phylogenies without roots? A plea for the use of vouchers in molecular phylogenetic studies. Molecular Phylogenetics and Evolution 48(1): 369–371. <https://doi.org/10.1016/j.ympev.2008.03.024>
- Ratnasingham S, Hebert P (2007) BOLD: The Barcode of Life Data System. Molecular Ecology Notes 7(3): 355–364. <https://doi.org/10.1111/j.1471-8286.2007.01678.x>
- Sars GO (1879) Crustacea et Pycnogonida nova in itinere 2do et 3tio Expeditionis Norvegicae anno 1877 & 78 collecta (prodromus descriptionis). Archiv for Mathematik og Naturvidenskab 4: 427–476.
- Sars GO (1891) An account of the Crustacea of Norway with short descriptions and figures of all the species. Vol I. Amphipoda. Alb. Cammermeyers Forlag., Christiania and Copenhagen, 711 pp. <https://doi.org/10.5962/bhl.title.1164>
- Stephensen K (1925) Crustacea Malacostraca, VI: (Amphipoda, II). Danish Ingolf Expedition 3(9): 101–178.