

Mapping a brain parasite: occurrence and spatial distribution in fish encephalon

Ana Born-Torrijos^{a,b,*}, Gabrielle S. van Beest^{a,c}, Paolo Merella^d, Giovanni Garippa^d, Juan Antonio Raga^c, Francisco E. Montero^c

^a Institute of Parasitology, Biology Centre of the Czech Academy of Sciences, Branišovská 31, 370 05, České Budějovice, Czech Republic

^b Department of Coastal Systems, NIOZ Royal Netherlands Institute for Sea Research, Den Burg, Texel, Netherlands

^c Cavanilles Institute for Biodiversity and Evolutionary Biology, Science Park, University of Valencia, PO Box 22 085, 46071, Valencia, Spain

^d Dipartimento di Medicina Veterinaria, Università di Sassari, Sassari, Italy

ARTICLE INFO

Keywords:

Microhabitat selection
Trematoda
Histology
Brain-encysting
Cardiocephaloides longicollis

ABSTRACT

Parasites, especially brain-encysting trematodes, can have an impact on host behaviour, facilitating the transmission to next host and completion of the life cycle, but insufficient research has been done on whether specific brain regions are targeted. Using *Cardiocephaloides longicollis* as a laboratory model, the precise distribution of metacercariae in experimentally-infected, wild and farmed fish was mapped. The brain regions targeted by this parasite were explored, also from a histologic perspective, and potential pathogenic effects were evaluated. Experimental infections allowed to reproduce the natural infection intensity of *C. longicollis*, with four times higher infection intensity at the higher dose (150 vs 50 cercariae). The observed metacercarial distribution, similar among all fish groups, may reflect a trematode species-specific pattern: metacercariae occur with highest density in the optic lobe area (primarily infecting the periventricular gray zone of optic tectum) and the medulla oblongata, whereas other areas such as the olfactory lobes and cerebellar lobes may be occupied when the more frequently invaded parts of the brain were crowded. Mono- and multicysts (i.e. formed either with a single metacercaria, or with 2–25 metacercariae encapsulated together) may be formed depending on the aggregation and timing of metacercariae arrival, with minor host inflammatory response. Larvae of *C. longicollis* colonizing specific brain areas may have an effect on the functions associated with these areas, which are generally related to sensory and motor functions, but are also related to other host fitness traits such as school maintenance or recognition of predators. The detailed information on the extent and distribution of *C. longicollis* in fish encephalon sets the ground to understand the effects of brain parasites on fish, but further investigation to establish if *C. longicollis*, through purely mechanical damage (e.g., occupation, pressure and displacement), has an actual impact on host behaviour remains to be tested under controlled experimental conditions.

1. Introduction

Parasites can directly impact host physiology by influencing its growth, reproduction, metabolic or immune functions (e.g. Lochmiller and Deerenberg, 2000; Sánchez et al., 2018) affecting sometimes their behaviour and survival (Poulin, 2013). Some parasites display a number of strategies that enhance their transmission to the next hosts, facilitating the completion of the parasite's life cycle (Poulin, 2010). Typical direct and indirect phenotypic alterations that parasites have developed to increase their chances of being preyed upon along with their hosts include altered activity, reduced antipredator responses, or changes in

host morphology and microhabitat selection (reviewed in Moore, 2002; Lafferty and Shaw, 2013). In fish, brain-encysting parasites have been shown to cause changes in neurotransmitter levels or to damage neural tissue (Shaw and Øverli, 2012), impair shoaling and antipredator behaviour (e.g., Shirakashi and Goater, 2001; Helland-Riise et al., 2020), and to also increase the frequency of conspicuous behaviours that attract bird final hosts, such as flashing, surfacing and contorting (Lafferty and Morris, 1996).

As noted by Helland-Riise et al. (2020), there is insufficient research on whether specific brain regions are targeted by brain-infecting parasites, but this information is necessary to understand specific

* Corresponding author. Department of Coastal Systems, NIOZ Royal Netherlands Institute for Sea Research, Den Burg, Texel, Netherlands.

E-mail address: borntorrijos.ana@gmail.com (A. Born-Torrijos).

<https://doi.org/10.1016/j.ijppaw.2023.03.004>

Received 11 January 2023; Received in revised form 9 March 2023; Accepted 10 March 2023

Available online 11 March 2023

2213-2244/© 2023 The Authors. Published by Elsevier Ltd on behalf of Australian Society for Parasitology. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

mechanisms and the neurological processes manipulated by parasites. Just as the patterns of migration to the brain may be species-specific, the site selection or distribution of metacercariae within the brain may also be (Erasmus, 1972). For example, within the Diplostomidae, the family to which many of the trematodes inducing behaviour changes belong, the species *Posthodiplostomum Ptychocheilus* (Faust, 1917) Achatz et al., 2021 (formerly *Ornithodiplostomum ptychocheilus* (Faust, 1917) Dubois, 1936; Achatz et al., 2021) usually occupies the optic lobes and cerebellum of fish (Conn et al., 2008; Hendrickson, 1979), whereas *Diplostomum phoxini* can occupy the optic lobes, but the superior lobe of the cerebellum and medulla oblongata are more severely infected (Dezfuli et al., 2007a). Once trematode larvae, including diplostomoideans, reach their target organ, they undergo an increase in body size and complexity from the cercarial stage to the infective metacercaria (Matisz et al., 2010a; Conn et al., 2008). The time required for metacercariae to become infective is species-specific, e.g., *Diplostomum spathaceum* (Diplostomidae) requires 21 days, while *P. ptychocheilus* (Diplostomidae) needs ten weeks, both to successfully infect their definitive bird host (Seppälä et al., 2005; Shirakashi and Goater, 2005). Once metacercariae encyst, they are surrounded by one or two cyst walls that separate the parasite from the host tissue (Hoffman, 1958), but information on the organization of the cyst walls in metacercariae that are encysted individually or closely together with other growing larvae is scarce.

Therefore, to improve the knowledge on brain-encysting trematodes, a species with unknown specific distribution and damage to the brain was selected, the trematode *Cardiocephaloides longicollis* (Rudolphi, 1819) Dubois, 1982 (Strigeidae). This parasite infects the brain of more than 31 fish species from nine families, including numerous sparid species and the farmed *Sparus aurata* L. (Born-Torrijos et al., 2016), and is trophically transmitted to the definitive host, mainly gulls, when

infected fish are preyed upon by seabirds. Metacercariae of this parasite have been described to infect the optic lobes of the fish midbrain and, as commonly occurs in brain-encysting trematodes, have been implicated in potential changes to the host's behaviour including impaired vision and motor control of swimming that would make fish more prone to be preyed upon by final hosts (Prévot, 1974, Recherches sur les cycles biologiques et l'écologie de quelques trématodes nouveaux parasites de *Larus argentatus michaelis* Naumann dans le midi de la France. Université d'Aix-Marseille, France, PhD thesis). Although this hypothesis has been maintained over time (Prévot and Bartoli, 1980; Osset et al., 2005; Bartoli and Boudouresque, 2007; Born-Torrijos et al., 2016), it has never been experimentally tested. Nevertheless, before performing time-consuming behavioural experiments, it is important to characterise the precise distribution of the metacercariae in the different fish brain areas that could justify potential behavioural changes. To this end, the microhabitat selection, encystment and development of *C. longicollis* in the brain of its fish host was monitored. The artificial infection process was first optimized in a laboratory model, using different infection doses. To investigate whether there is a common pattern in the colonization of the different brain areas, the precise distribution of metacercariae was mapped in experimentally-infected as well as in wild and farmed fish, using a histological perspective also to identify the specific targeted regions and their potential pathological effects.

2. Material and methods

2.1. Host collection and experimental fish infections

Snails *Tritia reticulata* (L.), collected manually in spring at the littoral of an area with a high prevalence of infection (Born-Torrijos et al., 2016;

Table 1

Summary of fish samples used in the present study to investigate various aspects of *Cardiocephaloides longicollis* metacercarial infection, distribution and encystment in the fish brain (see Materials and methods for details). Standard length and weight of fish are given as mean $\pm$ SD (min–max). ‘Mean intens.’ indicates mean intensity, ‘Mtc distrib.’ contains the number of samples used for *C. longicollis* metacercariae distribution per brain regions, and ‘Prev’ *C. longicollis* prevalence. Spaces marked with a dash indicate that no data were produced for that category.

Group of fish	No. fish	Standard length (cm)	Weight (g)	Prev (%)	Mean intens.	Mtc. distrib.	Infection rate
Experm. infections <i>S. aurata</i>							
Lethal dose	18	11.76 $\pm$ 1.63 (9.50–14.50)	13.33 $\pm$ 1.54 (10.85–15.09)	100	115.8	–	38.59 $\pm$ 11.83 (25.0–70.0)
Low dose ^e	33	9.20 $\pm$ 1.64 (5.50–13.70)	23.81 $\pm$ 6.10 (15.24–41.2) ^a	100	7.5	–	15.03 $\pm$ 13.61 (0–62)
High dose ^{e,f}	36	13.11 $\pm$ 2.03 (10.50–18.20)	72.68 $\pm$ 55.93 (25.8–265.1)	100	31.3	n = 14	20.89 $\pm$ 15.55 (4–68.67)
Both doses	69	11.24 $\pm$ 2.70 (5.5–18.2)	53.63 $\pm$ 49.80 (15.24–265.1) ^b	100	19.9	–	18.09 $\pm$ 14.85 (0–68.67)
Wild fish							
<i>S. aurata</i> ^f	4	29.38 $\pm$ 0.95 (28.0–30.0)	755.45 $\pm$ 118.99 (691.6–933.8)	100	6.5	n = 4	–
<i>P. erythrinus</i>	10	21.75 $\pm$ 1.62 (20.0–25.0)	270.92 $\pm$ 72.54 (188.0–423.0)	20	3.0	n = 2	–
<i>L. mormyrus</i> ^f	10	21.1 $\pm$ 1.05 (20.0–23.0)	285.68 $\pm$ 17.88 (228.0–285.78)	70	5.1	n = 7	–
Farmed <i>S. aurata</i>							
Golfo Aranci (Italy) ^f	120	25.1 $\pm$ 1.32 (21.0–27.0) ^c	329.01 $\pm$ 140.61 (115.6–582.5)	40	1.3	n = 25	–
Stintino (Italy)	30	24.12 $\pm$ 1.24 (22.0–7.05) ^d	418.71 $\pm$ 16.41 (399.0–464.0) ^d	0	0	–	–
Marine pond (Spain)	39	24.86 $\pm$ 3.1 (21.0–33.0)	410.82 $\pm$ 61.70 (311.0–628.86)	25.6	13.1	half brain n = 10	–
Levantine farm (Spain)	211	21.90 $\pm$ 2.97 (15.5–36.3)	311.54 $\pm$ 143.23 (90.79–750.4)	0	0	–	–
Histology <i>S. aurata</i>							
	9	12.29 $\pm$ 4.74 (8.36–22.1)	95.20 $\pm$ 121.00 (13.01–297.20)	100	–	–	–

^a Based on n = 23.

^b Based on n = 59.

^c Based on n = 25. Total length is 26.18 $\pm$ 4.02 (19.2–32.7) (n = 115).

^d Based on n = 25.

^e Used for statistical analyses testing the effect of infection dose on metacercarial infection rate.

^f Used for statistical analyses testing the effect of brain region on No. metacercariae.

“Beach of the Eucalyptus” (40°37′35.0″N, 0°44′31.0″E) in Els Alfacs Lagoon (Ebro Delta, Spain)), were acclimated and screened for *C. longicollis* infection, by placing them in individual wells for 24 h at 25 °C, 12:12 h light:dark cycle and 35 psu salinity and checking for the emergence of cercariae. The experimental fish were pathogen-free gilt-head seabream *Sparus aurata* L. (SL min–max, 5.5–8.2 cm, Table 1) supplied by a hatchery in Burriana (Castellón, Spain), which were kept in 3000 l tanks at the SCSIE facilities (Central Support Service for Experimental Research, University of Valencia) until used for the

experimental infections. Exposure of fish to *C. longicollis* cercariae followed the procedure described in van Beest et al. (2019). In brief, freshly emerged cercariae (<2 h old) pooled from different infected snails were placed in a limited volume of water together with fish for 1 h and 30 min. At the end of the experiment, fish were euthanised with an overdose of MS222 (tricaine methanesulfonate, 0.03% solution buffered in seawater; Sigma-Aldrich).

Data on the infection rate of *C. longicollis* metacercariae were obtained from experimental infections using different doses of cercariae

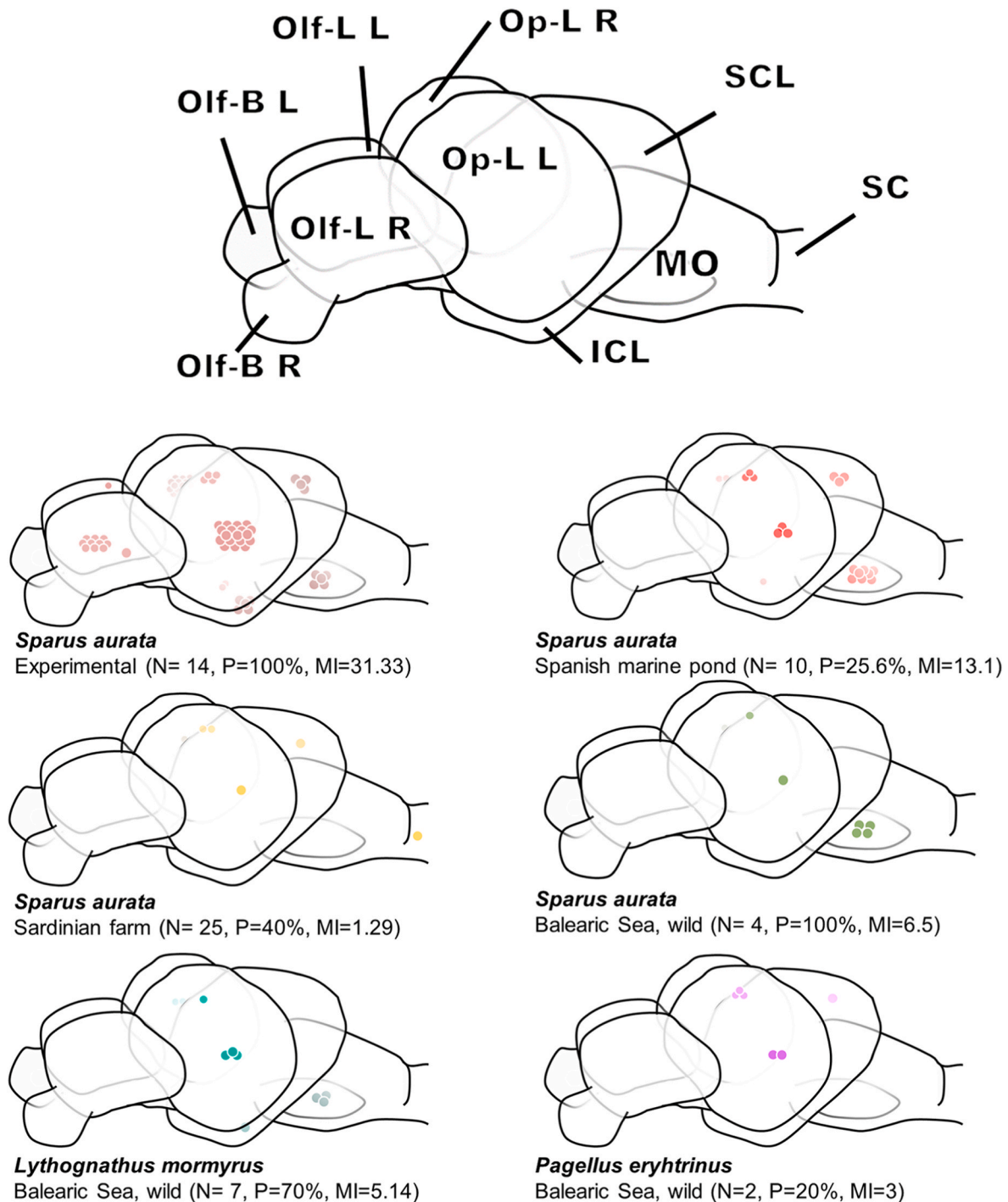


Fig. 1. Distribution of metacercariae of *Cardiocephaloides longicollis* in the different fish brain regions, i.e., olfactory bulbs (Olf-B), olfactory lobes (Olf-L), optic lobe region (Op-L), inferior and superior cerebellar lobes (ICL, SCL), medulla oblongata (Mo), and spinal cord (SC). Metacercarial distribution in different fish species sampling locations are provided. N, number of infected brains used for metacercarial distribution; P, prevalence (based on total number of fish, see Table 1); MI, mean intensity. Note that the number of metacercariae in the brain of fish from the marine pond is based only of half brain (see Materials and methods).

(total $n = 87$, see Table 1). To establish a standardized infection protocol and investigate the potential role of infection dose on infection success, experimental fish were infected with either 50 or 150 cercariae (low vs high infection dose, respectively), and the number of metacercariae was quantified after one month post-infection (mpi). In an attempt to evaluate the number of cercariae that an infected fish could survive and to maximize the infection of fish, a dose of 300 cercariae was used in a single infection, which caused the death of all fish 21 days post infection, whereas control fish that followed the same procedure (with absence of cercariae) and were kept in same conditions suffered no mortality.

2.2. Fish dissection and quantification of metacercariae

The number of metacercariae in the brain of different sparid species, including wild and farmed gilthead seabream *S. aurata*, was recorded in a total of 424 fish (Table 1). These included fish with different origins to test whether *C. longicollis* follows a similar pattern infecting them, i.e. wild-caught specimens of three species of sparids ($n = 24$, *Lithognathus mormyrus* (L.), *Pagellus erythrinus* (L.), *S. aurata*, see Table 1) from commercial fishermen and fish markets and farmed gilthead seabreams from a marine pond (Southern Atlantic, Spain, $n = 39$) and from fish farms in the Eastern and Western Spanish Mediterranean, i.e., from the Levantine coast in Spain ($n = 211$), and two fish farms in Italy (Sardinia, Golfo Aranci, $n = 120$, Stintino, $n = 30$) (Table 1). Of these, a total of 48 naturally-infected fish (including wild and farmed fish, Table 1) were used to investigate the exact distribution of metacercariae in the brain by carefully dissecting different subregions of the brain of infected fish. In addition, the brain of 14 fish infected with a high dose of cercariae were also dissected by subdividing the brain regions.

To investigate whether the parasite has a non-random distribution targeting specific areas of the brain, besides following the descriptions by Wullimann et al. (1996), the different brain regions were subdivided according to the natural division and distinct boundaries of the brain. This was also done to facilitate fine dissection and identification of areas during extensive samplings, when the brain is usually ignored and therefore not analysed. The areas were identified into seven regions (see Fig. 1): olfactory bulbs (Olf-B), olfactory lobes (Olf-L), optic lobe regions (Op-L), inferior and superior cerebellar lobes (ICL, SCL), medulla oblongata (Mo), and spinal cord (SC). When possible, the areas were divided by the sagittal plane separating the left and right sides of the brain, but differences in the number of metacercariae between sides were discarded (linear model: $\log(\text{No. metacercariae} + 1) \sim \text{Brain side} + \text{No. total metacercariae}$, estimate = -0.0076 , SE = 0.0197 , $p = 0.699$), and data from both sides were pooled. Eyes were also analysed to exclude infections, as described for other *Cardiocephaloides* spp. (Timi et al., 1999; Vermaak et al., 2021). All fish were measured and weighed prior to dissection, and experimental fish were also measured before experimental infection to assess whether the infection rate was related to size. All brain region dissections and quantification of metacercariae were performed under a stereomicroscope (Leica MZ APO, Germany, equipped with camera Leica DFC295).

2.3. Statistical analyses: infection dose and distribution in the brain

To examine whether there was an effect of infection dose on the number of encysted metacercariae in the fish brain, i.e., the infection rate, a linear model was fitted, using ‘number of metacercariae’ (log transformed) as the response variable, and ‘infection dose’ (low vs high, i.e., 50 vs 150 cercariae) and ‘fish size’ (standard length, mm) as covariables.

To investigate possible differences in the number of metacercariae per brain region between different fish groups, two linear models (LMs) were fitted using ‘number of cysts/region’ as the response variable (log transformed), and ‘brain regions’ (Olf-B, Olf-L, Op-L, ICL, SCL, Mo, SC) and ‘fish group’ as well as their interaction as fixed variables, with the number of total cysts in the brain as a covariable to control for different

total cyst numbers. The fish groups tested were (i) experimentally-infected vs wild gilthead seabream (*S. aurata*) or (ii) wild gilthead seabream vs striped seabream (*L. mormyrus*). Other comparisons could not be statistically done as the number of fish was too small or only partial data were available (i.e. only half of the brains of gilthead seabream from the marine pond were dissected because the other half were used for histology, but the samples were not properly fixed and therefore could not be used, Table 1), but were included in the figures to visually represent the distribution of metacercariae. Data were statistically analysed using R (R Core Team, version 4.1.2), with the significance level set at 0.05, and the R^2 values of the models performed were reported (MuMIn package, Barton, 2022).

2.4. Ultrastructure and histology of metacercariae in fish brain

The formation of the cyst from metacercariae after reaching the target tissue was studied in nine experimentally-infected fish (Table 1), infected with 100 or 150 cercariae and dissected for histological analyses at different post-infection times (6, 21 days post infections (dpi), and 8 and 15 months post infection (mpi)). The brain and eyes were fixed in 10% buffered formalin for two days, dehydrated in graded alcohol and embedded in paraffin. Paraffin blocks were sectioned (5–7 μm , 20–40 sections per block) using a microtome (Leica RM 2125RT, Germany). Histological sections were stained with hematoxylin and eosin, mounted in DPX, and examined at $100\times$ to $400\times$ magnification (Leica DMR, Germany, equipped with a Leica DMC5400 camera).

To investigate the cysts formed by *C. longicollis* metacercariae in the brain, their ultrastructure was examined in isolated cysts collected from experimentally-infected fish (8 mpi) and wild fish (*S. aurata* and *L. mormyrus*). In addition, metacercariae encysted in brain tissue were collected from wild fish (*S. aurata*). All samples were fixed in 2.5% glutaraldehyde in phosphate-buffered saline (PBS, 0.1 mol L^{-1} , pH 7.4) and preserved at 4°C for 48 h. Samples were then washed three times in PBS (15 min each), post-fixed in 2% osmium tetroxide (2 h), washed three times in phosphate buffer (PB), and dehydrated in a graded series of acetone. Samples were then infiltrated and embedded in Epon resin. First, semithin sections of 400 nm were cut using a glass knife, stained with toluidine blue, and observed under a light microscope. Then, ultrathin sections (60–90 nm) were cut through selected regions using a diamond knife on a Leica Ultracut UCT ultramicrotome. These were placed on copper grids and stained with uranyl acetate and lead citrate according to Reynolds (1963). For observation a JEOL JEM-1010 transmission electron microscope was used (JEOL, Tokyo, Japan, accelerating voltage 80 kV, equipped with a CCD digital camera Mega View III at the Laboratory of Electron Microscopy, Institute of Parasitology, Biology Centre of the Czech Academy of Sciences (Czech Republic)).

3. Results

3.1. Quantification of *C. longicollis* metacercariae, experimental infection dose and infection rate

Metacercariae of *C. longicollis* were found in all groups of fish, with variable infection levels among experimentally-infected, wild and farmed fish (Table 1). In the farmed fish group, *C. longicollis* was found only in one of the Italian fish farms with a prevalence of 40% and a mean intensity of infection of 1.3, while it was absent from the Spanish Levantine coast. Nevertheless, in *S. aurata* from the marine ponds, the prevalence was 25.6%, with a mean intensity of 13.1. In wild fish, the prevalence of *C. longicollis* ranges from 20 to 100% (mean intensity 3.0 to 6.5). The low sample size in wild fish is due to these specimens being obtained for the analysis of metacercarial distribution in the brain (see below) rather than to study the prevalence of *C. longicollis* in these species, which is reported in Born-Torrijos et al. (2016).

All fish used in the infection experiments were successfully infected

with a prevalence of 100% and varying infection intensity depending on the fish group, with the experimental gilthead seabream being the most heavily infected. The mean intensity of infection was higher in the experimentally-infected fish (7.5–115.8) than in the wild (3–6.5) and farmed ones (1.3–13.1). No *C. longicollis* were found in the eyes of any of the fish. The extremely high intensity observed in the fish infected with 300 cercariae in a single dose (115.8 mean metacercariae/fish, 75–210 min–max metacercariae) was likely the reason why the fish did not survive longer than 21 dpi, since all 18 fish died within two days (while 25 uninfected fish from the same fish lot kept in the same conditions suffered no mortality). Therefore, a dose of 300 cercariae was considered lethal in fish from 10 to 15 cm. In the rest of experimentally-infected fish, which had a 100% survival, the infection dose showed a positive effect on the number of metacercariae that successfully arrived in the fish brain, meaning that using a higher dose significantly increased the infection rate of *C. longicollis* metacercariae (LM, estimate = 0.0107, SE = 0.0028, $p < 0.001$), with no effect of fish size (LM, estimate = 0.1024, SE = 0.0522, $p = 0.054$) (Suppl. Table S2, Fig. S1).

3.2. Metacercariae distribution in the fish brain

Metacercariae were found in all the seven regions of the fish brain, except for the olfactory bulb (the most anterior region, Fig. 1) and the spinal cord (the most posterior; except for one metacercaria in *S. aurata* from a Sardinian farm). Considering each fish group separately, the areas with higher numbers of metacercariae were the optic lobe (Op-L) region and the medulla oblongata (Mo) (Figs. 1 and 2), therefore the metacercarial distribution being similar in fish of all origins and species and showing only minor differences. While wild *P. erythrinus* and experimentally-infected *S. aurata* showed a clear increase in metacercariae in the Op-L region, wild and farmed *S. aurata* from the marine pond showed a higher number of metacercariae encysted in the Mo, as did wild *L. mormyrus*. The “free” metacercariae found during dissections were likely in a fluid-filled cavity within the optic lobes (see later). Comparisons between experimentally-infected and wild gilthead seabreams showed certain differences in the number of metacercariae in different brain regions. While in experimentally-infected fish, the Op-L

showed the highest number of metacercariae compared to the Olf-B (LM, estimate = 3.444, SE = 0.2663, $p < 0.001$), followed by the Olf-L, ICL and Mo (Suppl. Table S1i), the number of metacercariae in wild *S. aurata* was comparable between regions, with a more pronounced difference in the Op-L region in experimentally-infected fish. The number of metacercariae in the brain regions of both wild gilthead and striped seabream was similarly distributed, being significantly higher in the Op-L and Mo regions than in the Olf-B in the striped seabream (estimate = 0.940, SE = 0.2481, $p < 0.001$; estimate = 0.5919, SE = 0.2481, $p = 0.020$, Suppl. Table S1ii).

To describe the distribution of metacercariae, the Op-L region was included as an integrated area as certain structures such as the periventricular gray zone of optic tectum (PGZ) represent a continuum that cannot be to cleanly separated during dissection. In freshly dissected brains, the presence of metacercariae, including multicysts (see later; Figs. 3 and 4), was observed within the PGZ and below the medulla oblongata (Fig. 3 A, B), but to identify the specific location, histology was required to distinguish the different regions according to cell types. When the cercariae arrived in the brain, they were found at 6 dpi occupying only the tectal ventricle (Fig. 3 C) with no occupation within the surrounding tissues. When metacercariae grew (21 dpi, 8 and 15 mpi) they occupied a larger part of the ventricle and also migrated to the PGZ, but not to the tectum opticum (TeO, part of the Op-L, see Fig. 3 D). There were no remarkable pathologies associated with the metacercariae presence revealing that even though some of the *C. longicollis* metacercariae accumulate in the tectal ventricle, most did not occupy the optic lobe itself but were located in the PGZ and on the cerebellum (Fig. 3 E, F). In any case, the tectum opticum (TeO) was not occupied in any of the slides, with metacercariae being found in the fluid-filled cavity PGZ when the optic lobe was infected.

3.3. Mono- and multicysts of *Cardiocephaloides longicollis*

Cercariae of *C. longicollis* were found in the brain by 6 dpi with a similar morphology to that of free-swimming cercariae (Fig. 3). However, by 21 dpi the parasite increased considerably in size and the tegumental structures changed to resemble metacercariae rather than free-

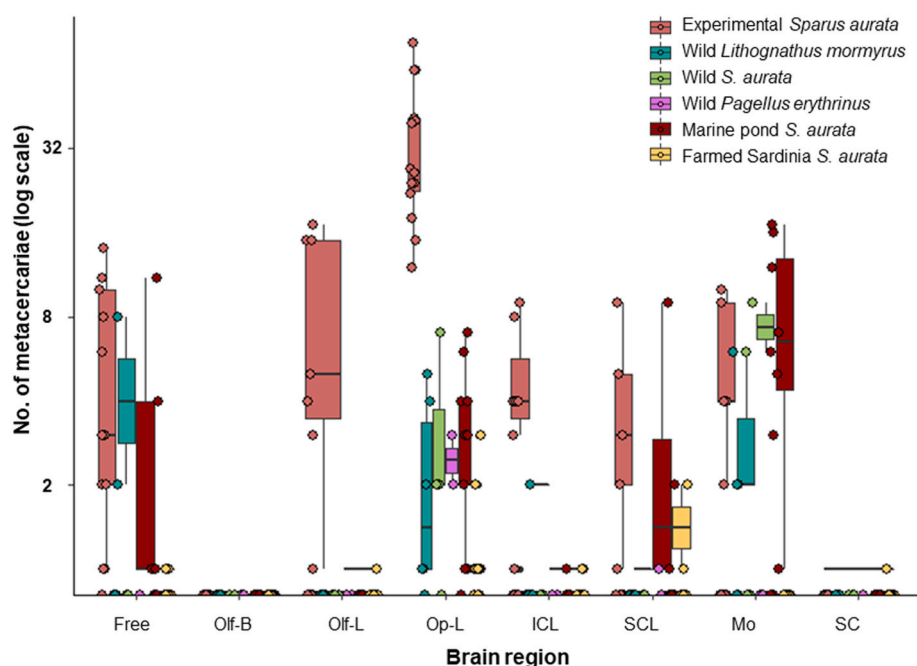


Fig. 2. Variation in the number of metacercariae of *Cardiocephaloides longicollis* encysted in different fish groups. Box plots represent the median number of metacercariae per brain region, upper and lower quartile (box) with maximum and minimum ranges (whiskers). Olfactory bulbs (Olf-B), olfactory lobes (Olf-L), optic lobe region (Op-L), inferior and superior cerebellar lobes (ICL, SCL), medulla oblongata (Mo), and spinal cord (SC). Y-axis is represented in logarithmic scale, and dots represent jittered raw data.

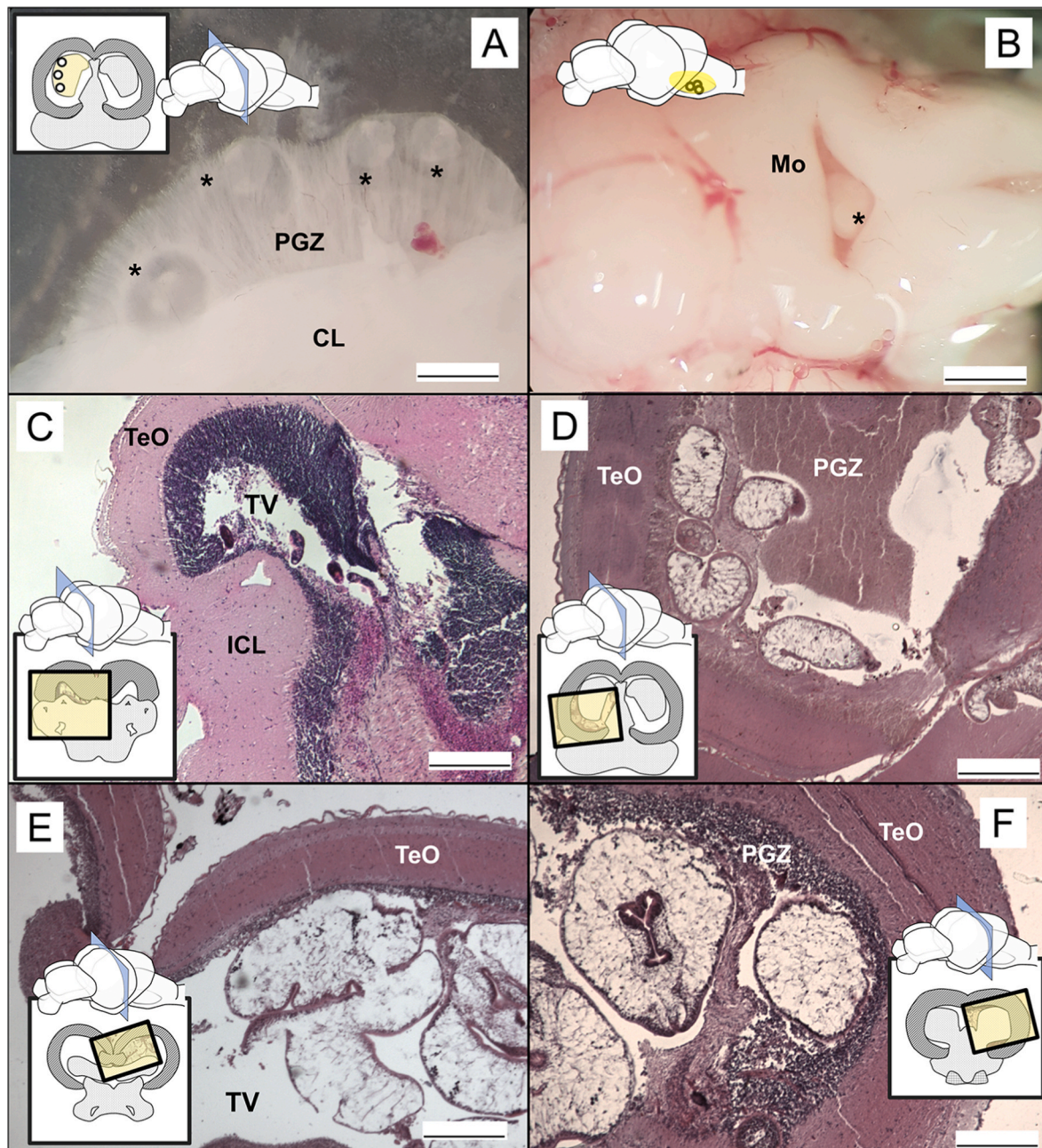


Fig. 3. Occupation of the fish brain by *Cardiocephaloides longicollis* in fresh (A, B) and histological samples (C–F). *Cardiocephaloides longicollis* metacercariae within (A) the PGZ and (B) the medulla oblongata in experimentally-infected fish one month after infection. Asterisks indicate the position of metacercariae. *Cardiocephaloides longicollis* metacercariae are found at 6 dpi in the tectal ventricle (C), and as they grow (D, 21 dpi; E, 8 mpi; F, 15 mpi) they occupy larger part of the tectal ventricle, and also the PGZ. The representations of brains indicate the sections and positions (yellow square) where metacercariae have been found. Legend: TeO striped, cerebellum in dots and Mo squared. ICL, inferior cerebellar lobe; Mo, medulla oblongata; PGZ, periventricular gray zone of optic tectum; TeO, tectum opticum; TV, tectal ventricle. Scale bars: A = 300 μ m; B = 450 μ m; C–F = 200 μ m. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

living cercariae, but lacking a visible cyst. Larvae found at 8 and 15 mpi had the normal morphology of encysted metacercariae that had developed a cyst in the brain. These were surrounded by a wall, and filled with a viscous glycocalyx-like material that maintained the position of the parasite within the cyst. Nevertheless, the host surface showed little disruption in response to the parasite.

Two types of cyst of *C. longicollis* metacercariae were observed in the brain of captured and experimentally-infected fish: (i) monocyst in which a single metacercaria was wrapped by a capsule (Fig. 4 B), or (ii) multicyst in which 2 to 25 metacercariae were encapsulated together (Fig. 4 I). Whereas the first type had a single wall in direct contact with

the host, the multicyst may have a common capsule surrounding all metacercariae, or each metacercariae may produce its own capsule as a single parasite, likely resulting in metacercariae with more than one capsule. The composition of the capsule walls, both inner and outer, was identified by TEM and was composed mostly of host cells in necrosis and filamentous material possibly originated from the glycocalyx between the tegument of the parasite and the capsule (Fig. 4 C, G). The capsule was produced by the parasite so that the material accumulated in the inner layers of the capsule wall (Fig. 4 C–D), where the necrotic cells were probably produced by the host tissue when it came into contact with the parasite. Thus, the necrotic cells appeared to accumulate on the

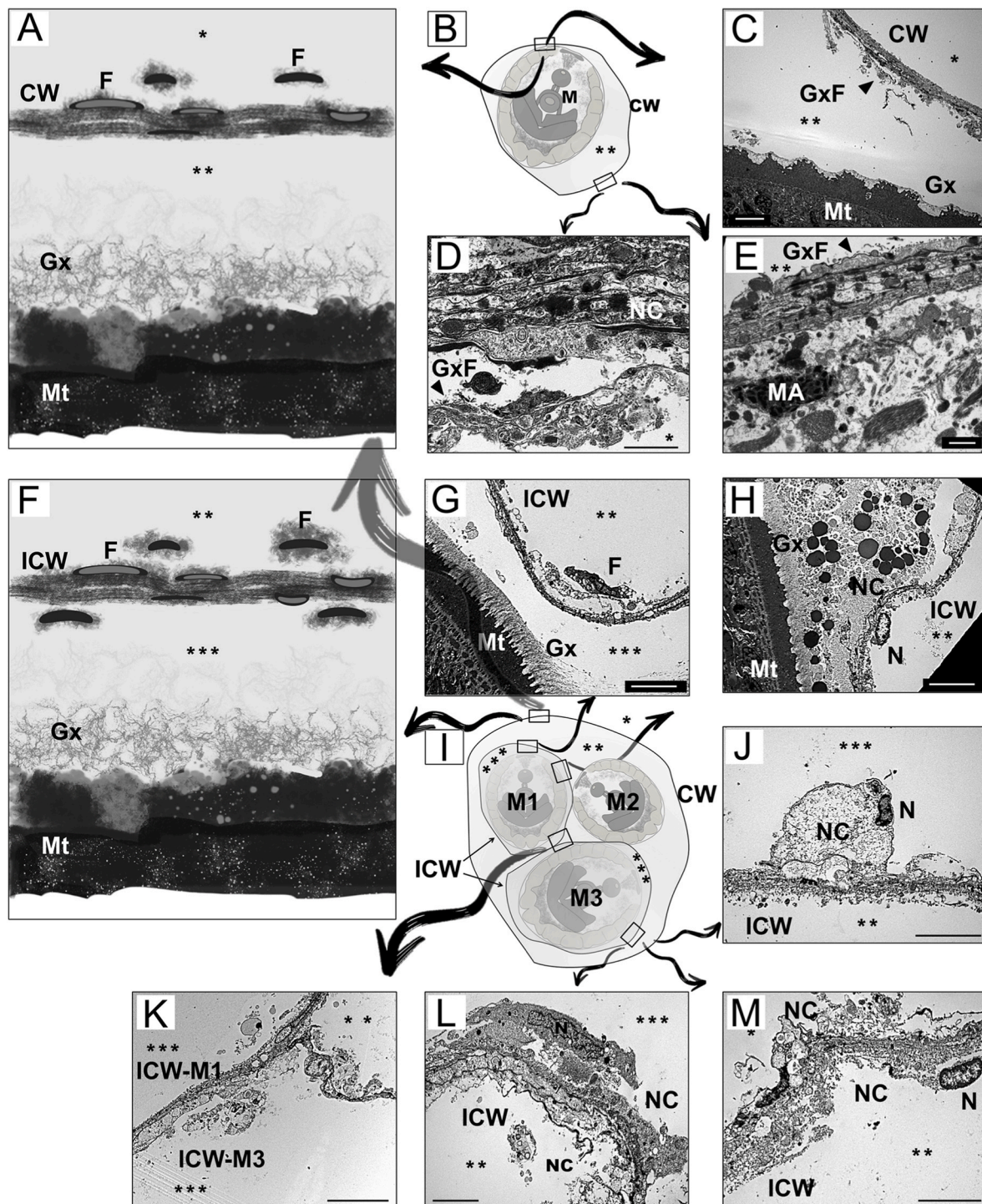


Fig. 4. Transmission electron micrographs showing the tegument and capsule walls of metacercariae of *Cardiocephaloides longicollis*. A and F illustrate the capsule wall of monocyst and multicyst metacercariae; B and I represent diagrams of monocyst and multicyst metacercariae showing the location of the following TEM micrographs. C – E Longitudinal section through the capsule wall and tegument of a monocyst. G, H, J–M Longitudinal section through the inner capsule wall and tegument of a multicyst metacercaria. D, E, J–M Detail of necrotic material accumulated on the capsule wall surrounding the metacercaria. K, L, M Detail of inner capsule walls merging together within a multicyst. CW, capsule wall; F, fibrocyte; Gx, glycocalyx; GxF, glycocalyx filaments; ICW, inner capsule wall; M, metacercaria; M1–M3 number of metacercaria in a multicyst; MA, macrophage; Mt, metacercarial tegument; N, nucleus; NC, necrotic cells. Head arrows indicate glycocalyx filaments, asterisks (*) outside of the cyst, (**) inside of the cyst, (***) inside of the cyst when encysted with more than one capsule wall. Scale bars: D, E = 1 µm; C, G, H, J, L, M = 5 µm; K = 10 µm.

outer layers of the capsule of monocysts and also in the outermost capsule of a multicyst (Fig. 4 A–E). However, the composition of the wall in each individual capsule within a multicyst was different, showing cells in necrosis on both sides of the wall (Fig. 4 F–M). In this case, the

degraded material was not only in the outer, but also in the inner layers of the capsule wall, resulting in a thick but loose capsule wall (Fig. 4 L, M) that likely merged with other inner wall capsules close to it (Fig. 4 K). Although the immune response was likely to be very low, TEM revealed

that the capsule wall was characterised by the presence of fibrocytes that were flattened in the inner layers of the wall. In addition, the host tissue in contact with the cyst showed the occasional presence of macrophages (Fig. 4 E).

4. Discussion

The optimized experimental infection protocol allowed for the reproduction of the natural infection intensity of *C. longicollis* in the fish's brain and showed that metacercariae occur with the highest intensity in the optic lobe area. Higher resolution examination of the brain revealed that the cercariae occupy the PGZ area as early as six days after infection and remain without occupation in the surrounding tissue, growing into larger encysted metacercariae that may be single or grouped in multiple cysts. The higher intensity in the optic lobe area and the medulla oblongata may point to a targeted site selection occupying the brain areas encoding motor, visual and basal physiological functions. However, directly connecting the occupation of these areas to effective behavioural changes remains to be confirmed with behaviour experiments.

As expected, the different doses used resulted in different infection rates, with more than four times higher intensity of infection at the higher dose. Although an infection intensity of ~116 metacercariae was achieved with the highest dose used (300 cercariae), the concentration of larvae was so high that it became lethal to ~12 cm long fish. Prévot and Bartoli (1980) reported hyperinfection with *C. longicollis* and subsequent death of two other sparid species, but did not provide information on infection dose and intensity. The highest intensity recorded in wild fish hosts is ~67 metacercariae in another sparid species (*Diplodus annularis*, 150–162 mm, Born-Torrijos et al., 2016, Suppl. Table S7), so it is not surprising that this dose was lethal. The lethality observed a few days after infection could also be related to the simultaneous arrival and disruption of brain tissue when cercariae begin to grow, which could interfere with the normal development, and likely contribute to brain tissue destruction and reduced fish survival (Sandland and Goater, 2001).

The presence of *C. longicollis* is reported in farmed gilthead seabream from a new aquaculture facility in Italy, with relatively low prevalence compared to that found in a Spanish fish farm by Born-Torrijos et al. (2016). However, the current negative data from Spanish farms on the Levantine coast suggest that the occurrence of the parasite in aquaculture net pens may have a patchy distribution, likely related to several factors such as seafloor topography, ocean currents, the presence of suitable hosts in the surrounding, and even the ability of larvae to reach their next host (van Beest et al., 2022). Fish from marine ponds showed a higher infection intensity than other farmed fish, likely because these ponds are in natural spaces and have an influx of parasite infective stages (i.e. cercariae) from the wild surrounding fauna through the external water supply they typically require (Bouwmeester et al., 2021). The transmission of *C. longicollis* to fish could thus be enhanced if intermediate hosts are concentrated in these ponds with slower water renewal, but further snail sampling would be needed to confirm this. The absence of *C. longicollis* in the eyes of any fish (see also Born-Torrijos et al., 2016) is noteworthy because other *Cardiocephaloides* spp. and various brain-encysting larvae have also been found in the eyes (Hendrickson, 1979; Timi et al., 1999; Muzzall and Kilroy, 2007; Vermaak et al., 2021).

The symmetrical distribution of *C. longicollis* metacercariae in both cerebral hemispheres seems to be a common feature (Siegmund et al., 1997). Nevertheless, the metacercariae followed a non-random distribution, possibly reflecting a trematode species-specific distribution, as their pattern in the brain of the experimentally-infected, wild and farmed hosts was overall similar, with the optic lobe region and the medulla oblongata as the most heavily infected (Fig. 1).

Experimentally-infected gilthead seabream showed higher concentration in the optic lobe than the wild individuals, however, this could be

related to their higher overall intensity, as other areas also had higher infection rates (19.94 vs 6.5, Figs. 1 and 2).

The olfactory bulb (Olf-B) was the only area completely free of metacercariae of *C. longicollis*, and only a single metacercaria was found in the spinal cord of a farmed fish from Italy. Dezfili et al., 2007a showed in another diplostomoidean, *D. phoxini*, the absence of metacercariae in the olfactory bulb, as well as the olfactory lobes, the inferior lobe of the cerebellum and the pituitary. However, Barber and Crompton (1997) suggested that this parasite species, infecting in this case a different fish species, could occupy the olfactory lobes and the spinal cord when the more frequently invaded parts of the brain were crowded. This might not be the case with *C. longicollis* as the spinal cord was free of infection even in experimentally heavily infected fish. However, the medulla oblongata, the region closest to the spinal cord (Fig. 1), had a high presence of *C. longicollis* metacercariae in experimentally-infected, wild and farmed fish except in the specimens from Golfo Aranci and wild *P. erythrinus*, which also had lower overall intensities. It cannot be thus ruled out that cercariae occupy areas near their entry point, as suggested in species aggregating in brain regions after likely following the optic and cranial nerves (Hendrickson, 1979; Helland-Riise et al., 2020). Although cercariae of *C. longicollis* have been observed penetrating areas likely connected to migration routes to neuronal canals and also detected in the connective tissue of the oral cavity of the fish 4 h post infection (van Beest et al., 2019, 2022), the exact route of migrating cercariae remains unknown. They could also use the muscle or circulatory system to reach the target organ, as do other diplostomoidean (Erasmus, 1959; Ratanarat-Brockelman, 1974; Hendrickson, 1979; Haas et al., 2007; Matisz et al., 2010b; Matisz and Goater, 2010).

With all this together, it can be suggested that the primary site of arrival for *C. longicollis* might be the optic lobe area, more specifically the PGZ, and the medulla oblongata, whereas other areas such as Olf-L and ICL/SCL are occupied depending on the intensity of infection. This would also explain the higher occupation of the latter areas in the experimentally-infected fish, as metacercariae would accumulate there when the optic lobe and medulla oblongata are occupied by the sudden arrival of a high number of metacercariae. The “preference” of *C. longicollis* for the optic lobe area and medulla oblongata seems to be consistent with other brain-encysting parasites including diplostomids (Hendrickson, 1979; Radabaugh, 1980a; Barber and Crompton, 1997; Dezfili et al., 2007a; Helland-Riise et al., 2020). In general, metacercariae colonizing these areas are capable of altering the sensory and motor functions in fish (Barber and Crompton, 1997; Dezfili et al., 2007a). The function of the medulla oblongata has also been linked with involuntary responses such as breathing and digestion (Bernstein, 1970; Wullimann et al., 1996). Moreover, the optic tectum is recognized as the primary vision-processing centre in teleosts, responsible for receiving and integrating visual stimuli, including movement, color and shape, and also playing a role in integrating information received from the lateral line system (Springer et al., 1977; Weale, 1982; Guthrie, 1986; Wullimann et al., 1996). The midbrain, which includes the optic tectum, also regulates reproductive physiology (Wullimann et al., 1996). This broadens the traits that could be affected, as damage to the optic tectum is clearly associated with several host fitness traits such as school maintenance, prey detection or recognition of predators, as has been described in fish infected with other diplostomoideans (Hendrickson, 1979; Radabaugh, 1980b; So and Wittrock, 1982; Barber and Crompton, 1997; Sandland and Goater, 2001).

Metacercariae are usually considered immature, non-feeding and resting stages (Bush et al., 2001), but prior to cyst formation and this resting phase, diplostomoidean larvae that infect the brain normally undergo a variable and extended period of development before encystment (Erasmus, 1972; Dönges, 1969; Conn et al., 2008). Larvae of *C. longicollis* are described, for the first time, occupying the tectal ventricle and specifically the PGZ cavity as early as 6 dpi, increasing their size considerably, from ~100 to ~400 µm within three weeks, with no cyst formation during this period (Fig. 3), a similar growth to that

described by Prévot and Bartoli (1980) (this refers to size increase; note that the present measurements refer to fixed and histological material and the publication referred to fresh larvae). Then, *C. longicollis* remains occupying a larger section of the PGZ area until at least 15 months post infection (Fig. 3 F), not moving to other regions, as observed for some diplostomid metacercariae, which initially disperse in deeper areas of the brain and after a few days of infection concentrate in the outermost layers of the optic lobes and cerebellum (Conn et al., 2008; Matisz et al., 2010a; Matisz and Goater, 2010). Therefore, rather than “sitting” on the optic lobes as is the case in some species (see Plate 2 in Hoffman and Hoyme, 1958, and Figure 6 in Matisz et al., 2010b), *C. longicollis* appears to only exert mechanical pressure on the tectum opticum without invading it while specifically occupying the PGZ area, a fluid-filled cavity below the lobes. This would also explain the presence of “free” metacercariae during brain dissection (Fig. 2), probably suggesting these were also infecting the PGZ in the optic lobes.

Microlamellar and microvillar teguments have been described in developing metacercariae to allow their growth, expanding and forming a net around the larva (Goater et al., 2005). The tegumental filaments observed in *C. longicollis* could be the remnants of microvilli as these disappear at early stages (ca. 28 dpi, Goater et al., 2005). The filaments were surrounded by a glycocalyx-like material, as described for *P. pychocheilus* (Goater et al., 2005), occupying the space between the parasite and the cyst wall. In addition, the presence of two cyst walls has been described, with inner and outer cysts, similar to other trematodes including diplostomids (Hoffman, 1958; Halton and Johnston, 1982; So and Wittrock, 1982). The second or further cysts are likely to be formed when a new metacercaria arrives to an area already occupied by another larva and the two are then encapsulated together forming a double cyst. Because cyst walls in the brain are usually thin compared with cysts in other tissues (Halton and Johnston, 1982), the inner and outer layers are sometimes not easily distinguished. Monocysts of *C. longicollis* had a very thin inner layer composed largely of loose filaments of the parasite tegument surrounded by a thicker layer composed mostly of host cells, e. g., fibrocytes, with the degree of degradation varying depending on when these cells were incorporated into the outer layer. A similar interface of necrotic material/cell degeneration has been described between the inner and outer cyst layers (Mitchell, 1974; Halton and Johnston, 1982). Similarly, when *C. longicollis* is surrounded by two cysts, the outer cyst had host cells in the outer part and a thin inner layer in contact with the inner cyst had a thicker and lax cyst wall with disorganised structure. This phenomenon is probably due to host cells surrounding a metacercarial cyst that became entrapped in a new cyst, probably caused by the presence of a metacercaria nearby. Fibrocytes and macrophages are observed in the cyst areas where the host tissue and parasite cyst are in direct contact (Fig. 4 E), concentrated near the PGZ. Fibroblasts in the vicinity of the cyst have also been described (So and Wittrock, 1982; Matisz et al., 2010b) as a classic host inflammatory response to the parasite cyst wall or its excretory products (Halton and Johnston, 1982). Some studies have described massive inflammation of the meninges caused by encysted metacercariae (Matisz et al., 2010a) and even large cleared areas with a low proportion of the nervous tissue remaining, surrounded by cellular debris in heavily-infected fish (Dezfuli et al., 2007a). In this study, no severe pathological damage to brain tissue was observed. According to previous studies, the presence of rodlet cells is related to the host inflammatory defence response and have been described in brain-encysting trematodes (Dezfuli et al., 2007b; Matisz et al., 2010b). As Dezfuli et al., 2007b noted for *Diplostomum* sp., the host cellular reaction has been largely overlooked in histopathological studies with the rodlet cells mostly absent from the literature until recently. Further TEM studies of metacercariae in the brain may help to identify the presence of these cells. However, brain-infecting parasites generally do not appear to cause severe inflammation or pathology in brain tissue (this study; Halton and Johnston, 1982; Siegmund et al., 1997; Goater et al., 2005), with the most common effects being changes in host behaviour. Alterations in

host visual acuity have been associated with developing stages rather than with the encysted stages (Shirakashi and Goater, 2005), possibly related to direct penetration of parasite microvillar filaments into the host intercellular matrix or due to the feeding habits of developing metacercariae, which has only been demonstrated in two diplostomids, *D. phoxini* and *P. pychocheilus* (Bibby and Rees, 1971; Goater et al., 2005). Further studies on *C. longicollis* are necessary to investigate whether this is an extended process.

The similar distribution of metacercariae of *C. longicollis* in fish with different origins, i.e. experimentally-infected, wild and farmed fish, suggests a trematode species-specific targeted site selection to colonize the optic lobe area, specifically the PGZ, and the medulla oblongata of fish, with no severe tissue destruction. Using *C. longicollis* as a laboratory model has allowed to gain detailed knowledge on its infection and microhabitat selection, setting the ground to understand the effects of brain parasites on fish. Further investigations to establish whether *C. longicollis*, through mere mechanical damage (i.e., occupation, pressure, traction or displacement of tissue) has an actual impact on host behaviour, remains to be tested under experimental conditions.

Author contributions

A.B.-T., F.E.M. and G.S.B. conceived the ideas and designed the methodology; A.B.-T. and G.S.B. performed the experimental work, with help of P.M. for fish dissection and of F.E.M. for analysis of histological material; A.B.-T. and F.E.M. obtained the ethics approval; A.B.-T. supervised the project and acquired the funding, did the formal data analysis and wrote the first draft of the manuscript. All the authors contributed critically to the draft and gave final approval for the publication.

Funding

This research was financially supported by Czech Science Foundation [20-14903Y] to G.S.B., A.B.-T.; the Spanish Government[MINECO/FEDER PID2019-110730RB-I00]; the European Regional Development Fund [MCIN/AEI/10.13039/501100011033]; and the Valencian Regional Government [AICO/2021/279, GVA-THINKINAZUL/2021/029] (F.E.M., J.A.R.).

Data availability statement

Data available from Zenodo Digital Repository, doi: [10.5281/zenodo.7728536](https://doi.org/10.5281/zenodo.7728536).

Ethics statements

This study was approved by the Ethics Committee of the University of Valencia (Spain), authorisation procedure A1390575271192 (2014/037/UVEG/001, Dirección General de Agricultura, Ganadería y Pesca, Valencian Local Government).

Declaration of competing interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Acknowledgments

We acknowledge the core facility Laboratory of Electron Microscopy (LEM), Biology Centre of ASCR institution, supported by the MEYS CR (LM2018129 Czech-Biolmaging) and Central Service for Experimental Research of the University of Valencia (Spain), specifically the microscopy service and the experimental aquarium plant. We thank the Andromeda Group (Spain) for kindly providing the fish used in this

study, and Clemente Graziano (Compagnie Ittiche Riunite and Emanuele Depalmas (Palma d'Oro) (Italy) for their collaboration in fish sampling. The authors are grateful to Phillip Riekenberg for improving the English text.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ijppaw.2023.03.004>.

References

- Achatz, T.J., Chermak, T.P., Martens, J.R., Pulis, E.E., Fecchio, A., Bell, J.A., Greiman, S. E., Cromwell, K.J., Brant, S.V., Kent, M.L., Tkach, V.V., 2021. Unravelling the diversity of the Crassiphallinae (Digenea: Diplostomidae) with molecular phylogeny and descriptions of five new species. *Curr. Res. Parasitol. Vector-Borne Dis.* 1, 100051 <https://doi.org/10.1016/j.crpvbd.2021.100051>.
- Barber, I., Crompton, D.W.T., 1997. The distribution of the metacercariae of *Diplostomum phoxini* in the brain of minnows, *Phoxinus phoxinus*. *Folia Parasitol.* 44, 19–25.
- Bartoli, P., Boudouresque, C.F., 2007. Effect of the digenean parasites of fish on the fauna of Mediterranean lagoons. *Parassitologia* 49, 111–117.
- Bartón, K., 2022. MuMIn: multi-model inference. Package 'MuMIn' 1.47.1. <https://CRAN.R-project.org/package=MuMIn>.
- Bernstein, J., 1970. Anatomy and physiology of the central nervous system. In: Hoar, W. S., Randall, D.J. (Eds.), *Fish Physiology*. Academic Press, New York, pp. 1–90.
- Bibby, M.C., Rees, G., 1971. The ultrastructure of the epidermis and associated structures in the metacercaria, cercaria and sporocyst of *Diplostomum phoxini* (Faust, 1918). *Z. Für Parasitenkd. Parasitol. Res.* 37, 169–186.
- Born-Torrijos, A., Poulin, R., Pérez-del-Olmo, A., Culurgioni, J., Raga, J.A., Holzer, A.S., 2016. An optimised multi-host trematode life cycle: fishery discards enhance trophic parasite transmission to scavenging birds. *Int. J. Parasitol.* 46, 745–753. <https://doi.org/10.1016/j.ijpara.2016.06.005>.
- Bouwmeester, M.M., Goedknecht, M.A., Poulin, R., Thielges, D.W., 2021. Collateral diseases: aquaculture impacts on wildlife infections. *J. Appl. Ecol.* 58, 453–464. <https://doi.org/10.1111/1365-2664.13775>.
- Bush, A.O., Fernández, J.C., Esch, G.W., Seed, J.R., 2001. *Parasitism: the Diversity and Ecology of Animal Parasites*. Cambridge University Press, U.K.
- Conn, D.B., Goater, C.P., Bray, D., 2008. Developmental and functional ultrastructure of *Ornithodiplostomum ptychocheilus* diplostomula (Trematoda: Strigeoidea) during invasion of the brain of the fish intermediate host, *Pimephales promelas*. *J. Parasitol.* 94, 635–642. <https://doi.org/10.1645/GE-1421.1>.
- Dezfuli, B.S., Capuano, S., Simoni, E., Giari, L., Shinn, A.P., 2007a. Histopathological and ultrastructural observations of metacercarial infections of *Diplostomum phoxini* (Digenea) in the brain of minnows *Phoxinus phoxinus*. *Dis. Aquat. Org.* 75, 51–59. <https://doi.org/10.3354/dao075051>.
- Dezfuli, B.S., Giari, L., Shinn, A.P., 2007b. The role of rodlet cells in the inflammatory response in *Phoxinus phoxinus* brains infected with *Diplostomum*. *Fish Shellfish Immunol.* 23, 300–304. <https://doi.org/10.1016/j.fsi.2006.11.003>.
- Dönges, J., 1969. Entwicklungs- und Lebensdauer von Metacercarien. *Z. Parasitenkd.* 31, 340–366. <https://doi.org/10.1007/BF00259732>.
- Erasmus, D.A., 1972. *The Biology of Trematodes*. Edward Arnold, London.
- Erasmus, D.A., 1959. The migration of *Cercaria X* Baylis (Strigeida) within the fish intermediate host. *Parasitology* 49, 173–190. <https://doi.org/10.1017/S0031182000026810>.
- Goater, C.P., Bray, D., Conn, D.B., 2005. Cellular aspects of early development of *Ornithodiplostomum ptychocheilus* metacercariae in the brain of fathead minnows, *Pimephales promelas*. *J. Parasitol.* 91, 814–821. <https://doi.org/10.1645/GE-3485.1>.
- Guthrie, D.M., 1986. Role of vision in fish behaviour. In: Pitcher, T.J. (Ed.), *The Behaviour of Teleost Fishes*. Croom Helm, London, pp. 111–121.
- Haas, W., Wulff, C., Grabe, K., Meyer, V., Haeberlein, S., 2007. Navigation within host tissues: cues for orientation of *Diplostomum spathaceum* (Trematoda) in fish towards veins, head and eye. *Parasitology* 134, 1013–1023. <https://doi.org/10.1017/S0031182007002430>.
- Halton, D.W., Johnston, B.R., 1982. Functional morphology of the metacercarial cyst of *Bucephaloides gracilescens* (Trematoda: Bucephalidae). *Parasitology* 85, 45–52. <https://doi.org/10.1017/S0031182000054135>.
- Helland-Riise, S.H., Nadler, L.E., Vindas, M.A., Bengtson, E., Turner, A.V., Johansen, I.B., Weinertsmith, K.L., Hechinger, R.F., Øverli, Ø., 2020. Regional distribution of a brain-encysting parasite provides insight on parasite-induced host behavioral manipulation. *J. Parasitol.* 106, 188. <https://doi.org/10.1645/19-86>.
- Hendrickson, G.L., 1979. *Ornithodiplostomum ptychocheilus*: migration to the brain of the fish intermediate host, *Pimephales promelas*. *Exp. Parasitol.* 48, 245–258. [https://doi.org/10.1016/0014-4894\(79\)90106-1](https://doi.org/10.1016/0014-4894(79)90106-1).
- Hoffman, G.L., 1958. Studies on the life-cycle of *Ornithodiplostomum ptychocheilus* (Faust) (Trematoda: Strigeoidea) and the “self cure” of infected fish. *J. Parasitol.* 44, 416–421. <https://doi.org/10.2307/3274326>.
- Hoffman, G.L., Hoyme, J.B., 1958. The experimental histopathology of the “tumor” on the brain of the stickleback caused by *Diplostomum baeri eucaliae* Hoffman and Hundley, 1957 (Trematoda: Strigeoidea). *J. Parasitol.* 44, 374. <https://doi.org/10.2307/3274318>.
- Lafferty, K.D., Morris, A.K., 1996. Altered behavior of parasitized killifish increases susceptibility to predation by bird final hosts. *Ecology* 77, 1390–1397. <https://doi.org/10.2307/2265536>.
- Lafferty, K.D., Shaw, J.C., 2013. Comparing mechanisms of host manipulation across host and parasite taxa. *J. Exp. Biol.* 216, 56–66. <https://doi.org/10.1242/jeb.073668>.
- Lochmiller, R.L., Deerenberg, C., 2000. Trade-offs in evolutionary immunology: just what is the cost of immunity? *Oikos* 88, 87–98. <https://doi.org/10.1034/j.1600-0706.2000.880110.x>.
- Matisz, C.E., Goater, C.P., 2010. Migration, site selection, and development of *Ornithodiplostomum* sp. metacercariae (Digenea: Strigeoidea) in fathead minnows (*Pimephales promelas*). *Int. J. Parasitol.* 40, 1489–1496. <https://doi.org/10.1016/j.ijpara.2010.04.018>.
- Matisz, C.E., Goater, C.P., Bray, D., 2010a. Migration and site selection of *Ornithodiplostomum ptychocheilus* (Trematoda: Digenea) metacercariae in the brain of fathead minnows (*Pimephales promelas*). *Parasitology* 137, 719–731. <https://doi.org/10.1017/S0031182009991545>.
- Matisz, C.E., Goater, C.P., Bray, D., 2010b. Density and maturation of rodlet cells in brain tissue of fathead minnows (*Pimephales promelas*) exposed to trematode cercariae. *Int. J. Parasitol.* 40, 307–312. <https://doi.org/10.1016/j.ijpara.2009.08.013>.
- Mitchell, C.W., 1974. Ultrastructure of the metacercarial cyst of *Posthodiplostomum minimum* (MacCullum, 1921). *J. Parasitol.* 60, 67–74.
- Moore, J., 2002. *Parasites and the Behaviour of Animals*. Oxford University Press, Oxford.
- Muzzall, P.M., Kilroy, A.L., 2007. *Tylodelphys scheuringi* (Diplostomidae) infecting the brain of the central mudminnow, *Umbra limi*, in silver creek, Michigan. *U.S.A. Comp. Parasitol.* 74, 164–166. <https://doi.org/10.1654/4278.1>.
- Osset, E.A., Fernández, M., Raga, J.A., Kostadinova, A., 2005. Mediterranean *Diplostomus annularis* (Teleostei: Sparidae) and its brain parasite: unforeseen outcome. *Parasitol. Int.* 54, 201–206. <https://doi.org/10.1016/j.parint.2005.05.002>.
- Poulin, R., 2013. Parasite manipulation of host personality and behavioural syndromes. *J. Exp. Biol.* 216, 18–26. <https://doi.org/10.1242/jeb.073353>.
- Poulin, R., 2010. Parasite manipulation of host behavior. In: *Advances in the Study of Behavior*. Elsevier, pp. 151–186. [https://doi.org/10.1016/S0065-3454\(10\)41005-0](https://doi.org/10.1016/S0065-3454(10)41005-0).
- Prévot, G., Bartoli, P., 1980. Démonstration de l'existence d'un cycle marin chez les Strigeides: *Cardiocephalus longicollis* Szidat, 1928 (Trematoda: Strigeidae). *Ann. Parasitol. Hum. Comp.* 55, 407–425. <https://doi.org/10.1051/parasite/1980554407>.
- Radabaugh, D.C., 1980a. Encystment site selection in the brain-inhabiting metacercariae of *Ornithodiplostomum ptychocheilus* (Trematoda: Strigeoidea). *J. Parasitol.* 66, 183. <https://doi.org/10.2307/3280624>.
- Radabaugh, D.C., 1980b. Changes in minnow, *Pimephales promelas* Rafinesque, schooling behaviour associated with infections of brainencysted larvae of the fluke, *Ornithodiplostomum ptychocheilus*. *J. Fish Biol.* 16, 621–628. <https://doi.org/10.1111/j.1095-8649.1980.tb03741.x>.
- Ratanarat-Brockelman, C., 1974. Migration of *Diplostomum spathaceum* (Trematoda) in the fish intermediate host. *Z. Parasitenkd.* 43, 123–134. <https://doi.org/10.1007/BF00329170>.
- Reynolds, E.S., 1963. The use of lead citrate at high pH as an electronopaque stain in electron microscopy. *J. Cell Biol.* 17, 208–212.
- Sánchez, C.A., Becker, D.J., Teitelbaum, C.S., Barriga, P., Brown, L.M., Majewska, A.A., Hall, R.J., Altizer, S., 2018. On the relationship between body condition and parasite infection in wildlife: a review and meta-analysis. *Ecol. Lett.* 21, 1869–1884. <https://doi.org/10.1111/ele.13160>.
- Sandland, G.J., Goater, C.P., 2001. Parasite-Induced variation in host morphology: brain-encysting trematodes in fathead minnows. *J. Parasitol.* 87, 267. <https://doi.org/10.2307/3285040>.
- Seppälä, O., Karvonen, A., Valtonen, E.T., 2005. Manipulation of fish host by eye flukes in relation to cataract formation and parasite infectivity. *Anim. Behav.* 70, 889–894. <https://doi.org/10.1016/j.anbehav.2005.01.020>.
- Shaw, J.C., Øverli, Ø., 2012. Brain-encysting trematodes and altered monoamine activity in naturally infected killifish *Fundulus parvipinnis*. *J. Fish. Biol.* 81, 2213–2222. <https://doi.org/10.1111/j.1095-8649.2012.03439.x>.
- Shirakashi, S., Goater, C., 2005. Chronology of parasite-induced alteration of fish behaviour: effects of parasite maturation and host experience. *Parasitology* 130, 177–183. <https://doi.org/10.1017/S0031182004006432>.
- Shirakashi, S., Goater, C.P., 2001. Brain-encysting parasites affect visually-mediated behaviours of fathead minnows. *Ecoscience* 8, 289–293. <https://doi.org/10.1080/11956860.2001.11682655>.
- Siegmund, I., Franjola, R., Torres, P., 1997. Diplostomatid metacercariae in the brain of silversides from lake rihine, Chile. *J. Wildl. Dis.* 33, 362–364. <https://doi.org/10.7589/0090-3558-33.2.362>.
- So, F.W., Wittrock, D.D., 1982. Ultrastructure of the metacercarial cyst of *Ornithodiplostomum ptychocheilus* (Trematoda: Diplostomatidae) from the brains of fathead minnows. *Trans. Am. Microsc. Soc.* 101, 181. <https://doi.org/10.2307/3225772>.
- Springer, A.D., Easter, S.S., Agranoff, B.W., 1977. The role of the optic tectum in various visually mediated behaviors of goldfish. *Brain Res.* 128, 393–404. [https://doi.org/10.1016/0006-8993\(77\)90166-4](https://doi.org/10.1016/0006-8993(77)90166-4).
- Timi, J.T., Martorelli, S.R., Haydeé Sardella, N., 1999. Digenetic trematodes parasitic on *Engraulis anchoita* (Pisces: Engraulidae) from Argentina and Uruguay. *Folia Parasitol.* 46, 132–138.
- van Beest, G.S., Villar-Torres, M., Raga, J.A., Montero, F.E., Born-Torrijos, A., 2019. In vivo fluorescent cercariae reveal the entry portals of *Cardiocephaloides longicollis* (Rudolphi, 1819) Dubois, 1982 (Strigeidae) into the gilthead seabream *Sparus aurata* L. *Parasit. Vectors* 12, 92. <https://doi.org/10.1186/s13071-019-3351-9>.

- van Beest, G.S., Montero, F.E., Padrós, F., Raga, J.A., Born-Torrijos, A., 2022. The versatility of simplicity: structures of *Cardiocephaloides longicollis* used for different purposes during cercarial transmission. Integr. Comp. Biol. icac102. <https://doi.org/10.1093/icb/icac102>.
- Vermaak, A., Smit, N.J., Kudlai, O., 2021. Molecular and morphological characterisation of the metacercariae of two species of *Cardiocephaloides* (Digenea: Strigeidae) infecting endemic South African klipfish (Perciformes: Clinidae). Folia Parasitol. 68 <https://doi.org/10.14411/fp.2021.007>.
- Weale, R.A., 1982. Focus on Vision. Harvard University Press, Cambridge.
- Wullimann, M.F., Rupp, B., Reichert, H., 1996. Neuroanatomy of the Zebrafish Brain: a Topological Atlas. Birkhäuser Verlag, Basel.