

Spatial variability in condition, growth and diet of juvenile plaice (*Pleuronectes platessa*) at sandy beach nursery grounds on the south west coast of Ireland.

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Introduction

Characterization of suitable habitat for settlement of juvenile flatfish is important for the management of nursery areas. Food availability is one important determinant of habitat quality that can affect condition and growth, and thus survival, of flatfish (Gibson, 1994).

Temporal variation in plaice diet (Amara et al., 2001) and trophic niche overlap between different flatfish species (Beyst et al., 1999; Cabral et al., 2002) has been widely studied whereas levels of intraspecific variation in plaice diet at small spatial scales are relatively unknown.

This study investigates how diet, growth and condition of juvenile plaice vary over small spatial scales. It also serves as a pilot study to assess the importance of scale for characterizing habitat quality for flatfish over a larger scale.

Methods

Juvenile plaice were collected from 3 beaches and from 3 replicate hauls on each beach using a beach seine on 3 consecutive days in September 2007 (Fig. 1 & 2).

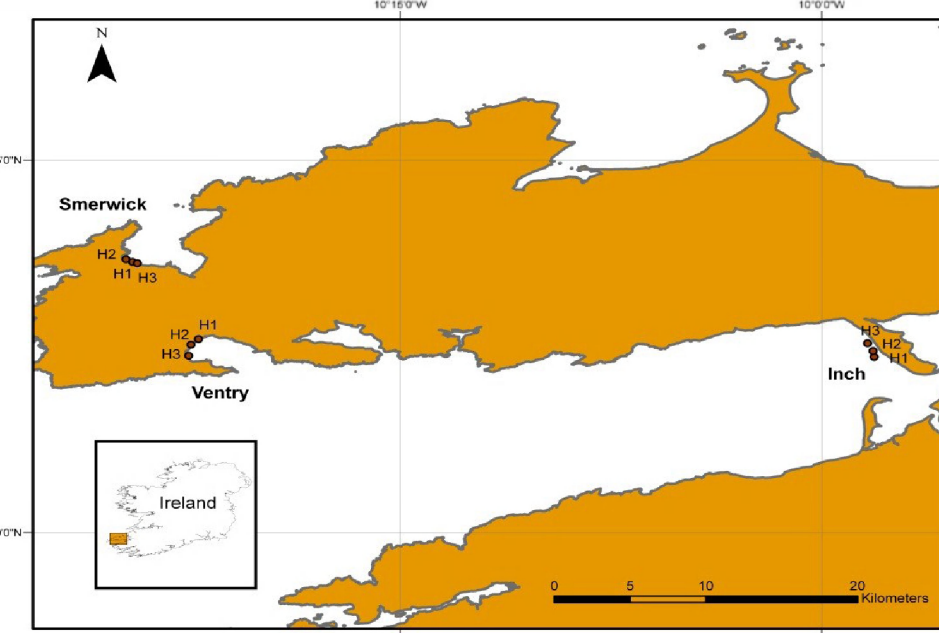


Fig. 1: Beach seine locations and position of 3 hauls (H1-H3) per beach.



Fig. 2: Beach seine used to sample flatfish.



Fig. 3: Gut content analysis flatfish.

6 Plaice of 7-9 cm were randomly selected per haul and kept frozen until analysis:

- Length, weight and morphological characteristics were measured and Fulton's condition factor was calculated.
- Otolith microstructure analysis was used to assess recent daily growth rates.
- Gut content analyses of the total 54 juvenile plaice were carried out (Fig. 3).

Results

SPATIAL VARIATION IN GROWTH

Fulton's K condition Index and mean otolith Increment width over the last 5 days (Fig. 4) were significantly higher for fish from Inch compared to fish from Ventry ($p=0.0037$ and $p=0.0029$, respectively; Two-way nested ANOVA with Tukey pairwise comparisons). There was no difference between hauls for either variable ($p>0.05$).

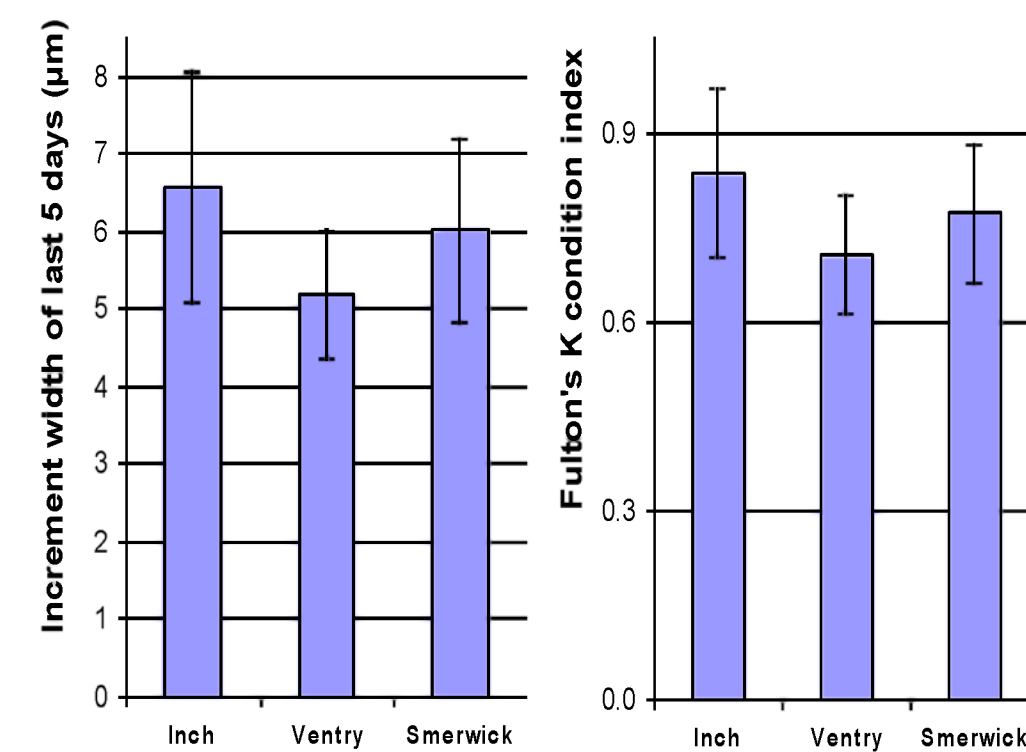


Fig. 4: Averages of growth measures per beach.

Mouth gape was not correlated with fish length but showed higher values for Smerwick ($4.05\text{mm}\pm 0.12$) compared to Inch ($3.41\text{mm}\pm 0.08$) and Ventry ($3.42\text{mm}\pm 0.14$) ($p<0.001$ and $p=0.001$ respectively; Two-way nested ANOVA with Tukey pairwise comparisons).

ANCOVA with fish length included as a covariate showed that otolith size was significantly correlated with fish length ($p<0.001$) and that fish from Ventry had larger otoliths at a given length compared to fish from Inch ($p=0.002$, Fig. 5 & 6). This reflects slower growth of plaice in Ventry compared to those in Inch on a longer term.

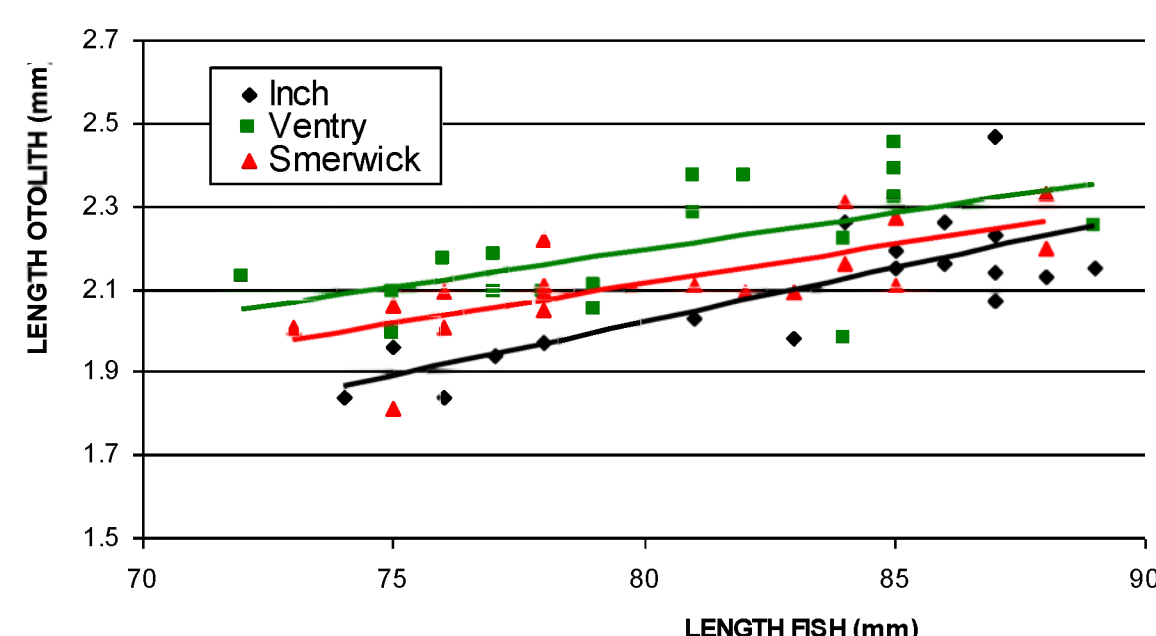


Fig. 5: Average Otolith length and Fish length ratio per beach.

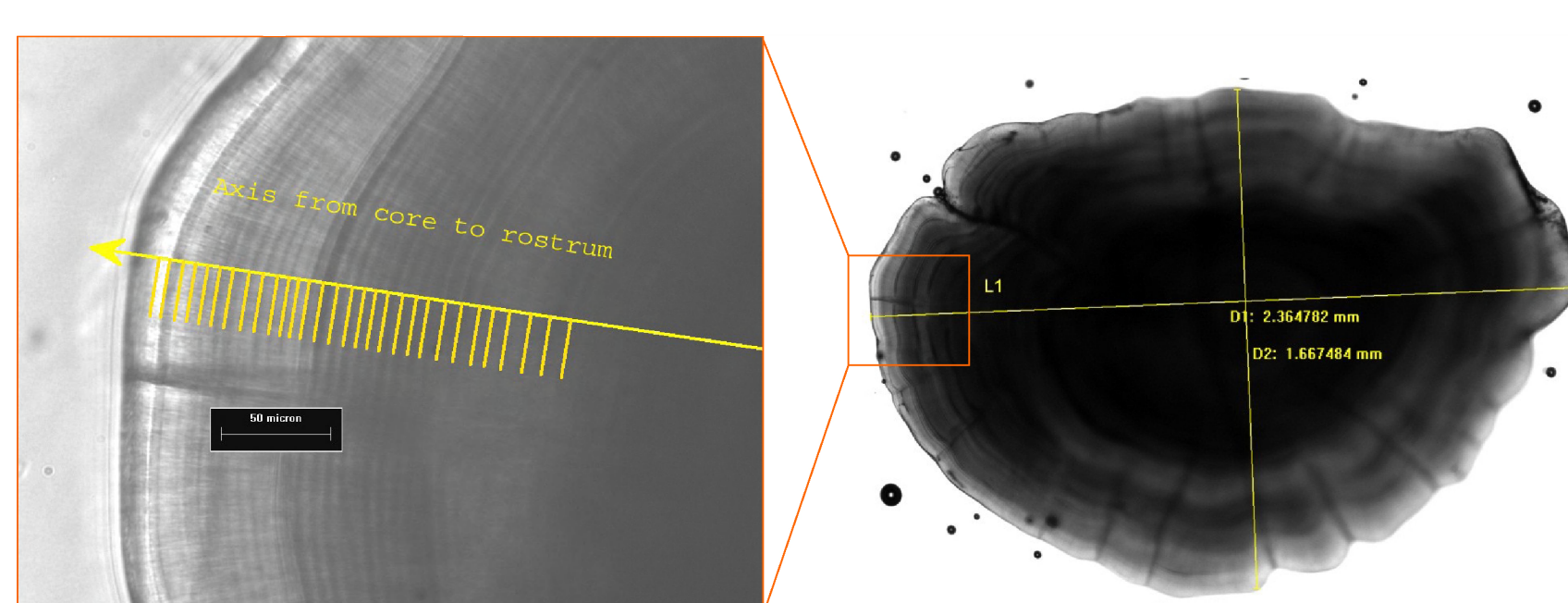


Fig. 6: Otolith daily increment width during the last 30 days.

Results

SPATIAL VARIATION IN DIET

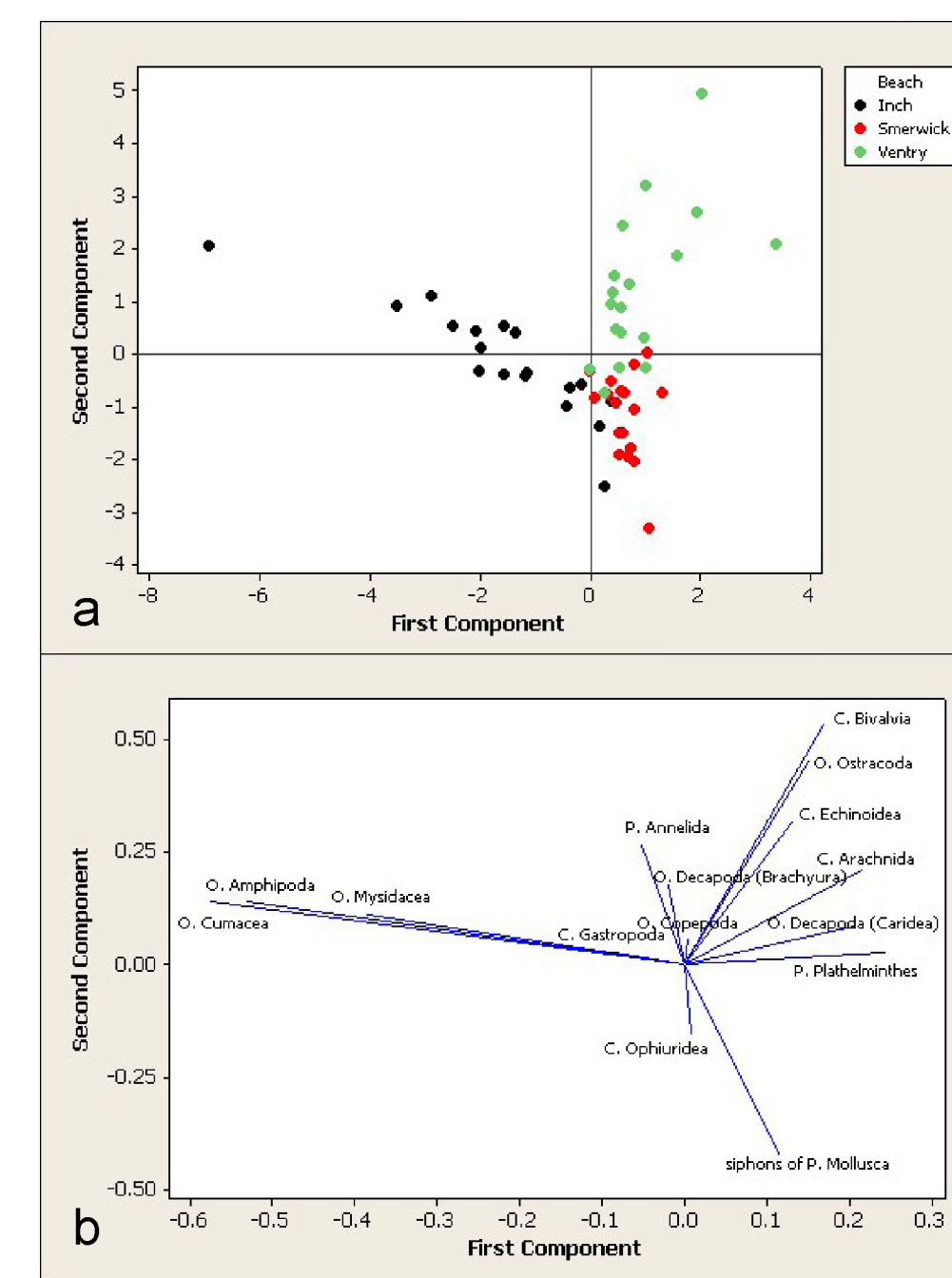


Fig. 7: PCA score plot (a) and loading plot (b) based on PCA of relative measures of prey groups in juvenile plaice, individually indicated by dots in (a).

ANOVA revealed significant variation in the diversity and volume of prey items between replicate hauls within beaches (Table 1)

	Beach Differences	Haul Differences
Tot. Prey	1.63 ns	4.37 **
Taxa	9.87 ***	1.67 ns
Tot. Prey Abundance	2.88 ns	4.05 **
Shannon-Wiener	0.40 ns	4.09 **
Fulness		

Table 1: Results of Two-Way nested ANOVA performed on diet descriptors, with F-value and levels of significance; ns, not significant; ** $p<0.01$; *** $p<0.001$

A Principal Component Analysis (PCA) characterized the 3 different beaches which were distinguished based on prey assemblages (Fig. 7) and prey descriptors (Table 1). **Inch** is characterized by high abundances of Amphipods, Mysids and Cumacea; species rarely abundant at other beaches. **Smerwick** is characterized by a high abundance of predominantly one prey item; siphons of bivalves. **Ventry** reveals a high prey diversity but very low abundances of prey (Table 1 & Fig. 7). However, gut composition also varied over smaller spatial scales (Fig. 8).

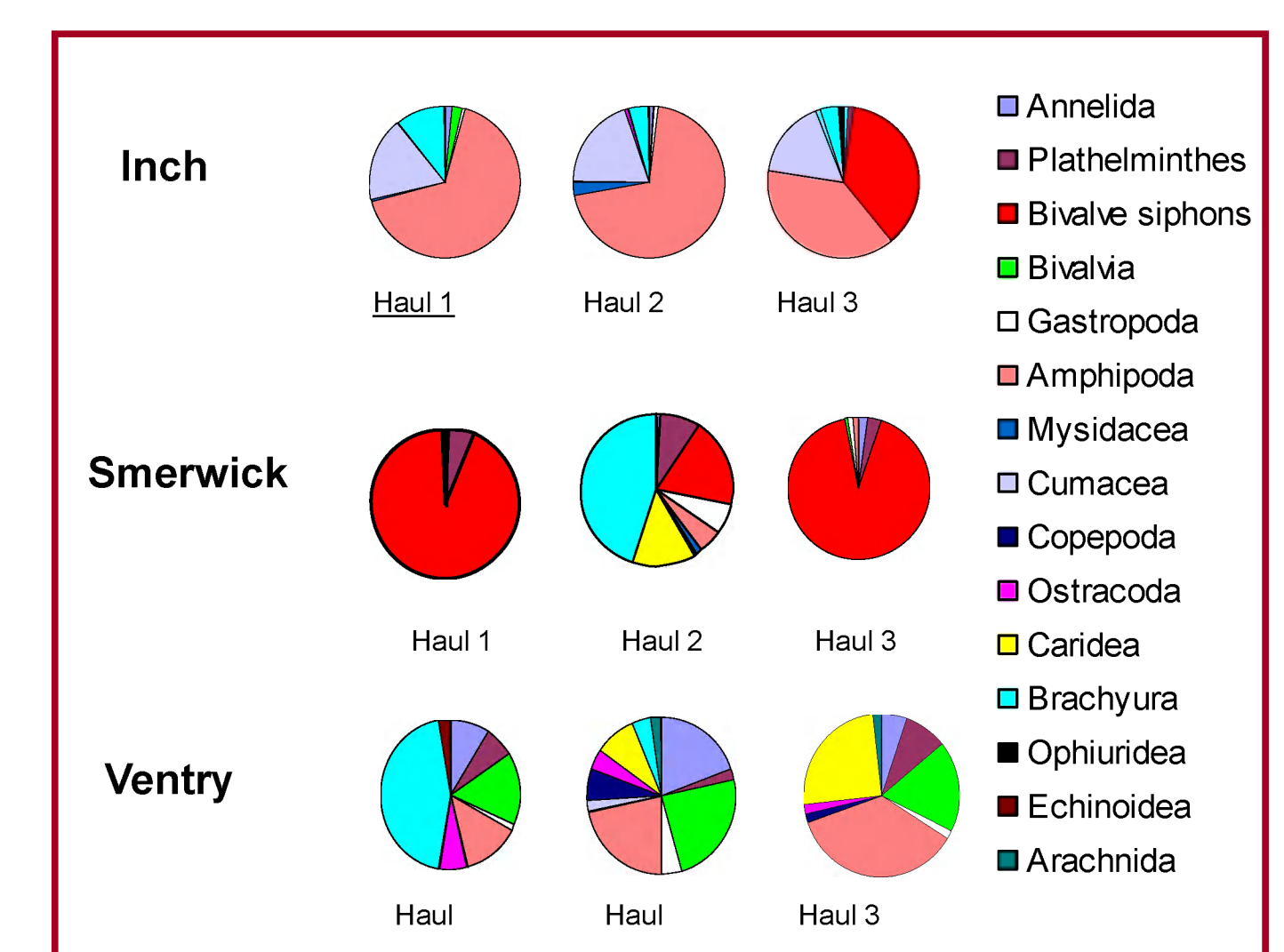


Fig. 8: Pie charts showing the taxonomic breakdown of prey items in the guts of juvenile plaice across replicate hauls on three beaches.

Conclusions

- Condition and growth of juvenile plaice varied over a spatial scale of several km's. This may reflect differences in habitat quality (diet or environmental conditions) between beaches. Further investigation is needed to determine the cause of the variability.
- Mouth gape varied significantly and considerably between beaches. This may reflect spatial variation in feeding behaviours, although further work is needed to confirm this.
- The composition and quantity of juvenile plaice gut contents varied over a small spatial scale (meters).
- Variation in composition of plaice diet was also observed over larger spatial scales (km's).
- The results highlight the importance of considering small scale variation when attempting to link broad scale habitat types to feeding, growth and condition of juvenile flatfish.

Literature cited:

Amara R., Laffargue P., Dewarumez J.M., Maryniak C., Lagardere F. and Luzac C. (2001). Feeding ecology and growth of O-group flatfish (sole, dab and plaice) on a nursery ground (Southern Bight of the North Sea). *Journal of Fish Biology* 58, 788-803.
Beyst B., Cattirjisse A. and Mees J. (1999). Feeding ecology of juvenile flatfishes of the surf zone of a sandy beach. *Journal of Fish Biology* 55, 1171-1186.
Cabral H.N., Lopes M. and Loeper R. (2002). Trophic niche overlap between flatfishes in a nursery area on the Portuguese coast. *Sci. Mar. (Barc.)* 66, 293-300.
Gibson R.N. (1994). Impact of habitat quality and quantity on the recruitment of juvenile flatfishes. *Netherlands Journal of Sea Research*: 32(2), 191-206.

Acknowledgements:

This project was funded through the STRIVE Programme by the Environmental Protection Agency. Many thanks to Paula Haynes, Stephen Comerford, the Marine Institute, BIM, Taighda Mara and the crew of the Commercial Fisheries Research Group for the collaboration during field sampling.