Assessing connectivity in young flatfish and its implementation in fisheries management



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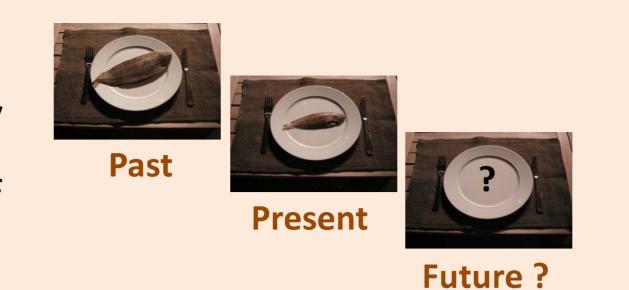
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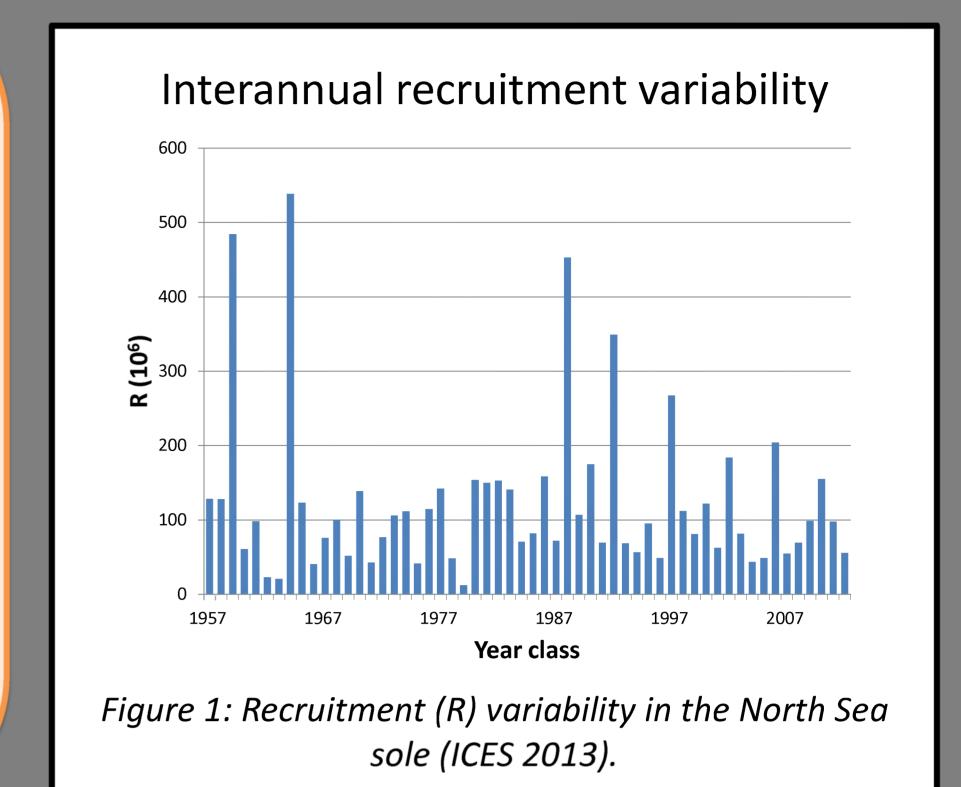


INTRODUCTION & AIMS

• Overfishing and selective fishing lead to a loss of genetic diversity as well as changed life cycles. Given the commercial importance of the North Sea sole species, conservation efforts have to be made.



- Interannual variability in recruitment* (Fig. 1) and dispersal (Fig. 4) remain poorly explained. However sustainable stock* management requires an upgraded biological assessment of population structure and connectivity patterns.
- Early life stages are of critical importance for recruitment and they provide a unique opportunity to reveal connectivity patterns.

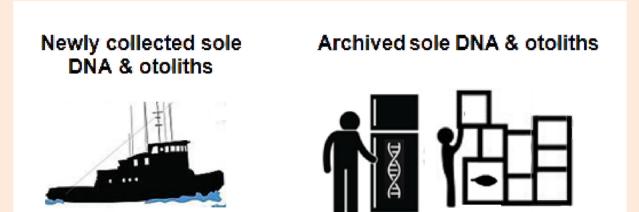


OBJECTIVES

- 1. Does larval dispersal vary in time and space?
- 2. What biotic and abiotic factors are driving larval connectivity?
- 3. Can we predict the impact on recruitment of changes in physical and biological drivers?

STUDY AREA

(1) TEMPORAL SAMPLING Years 2013 & 2014 + Archives 1975-2008



(2) SPATIAL SAMPLING

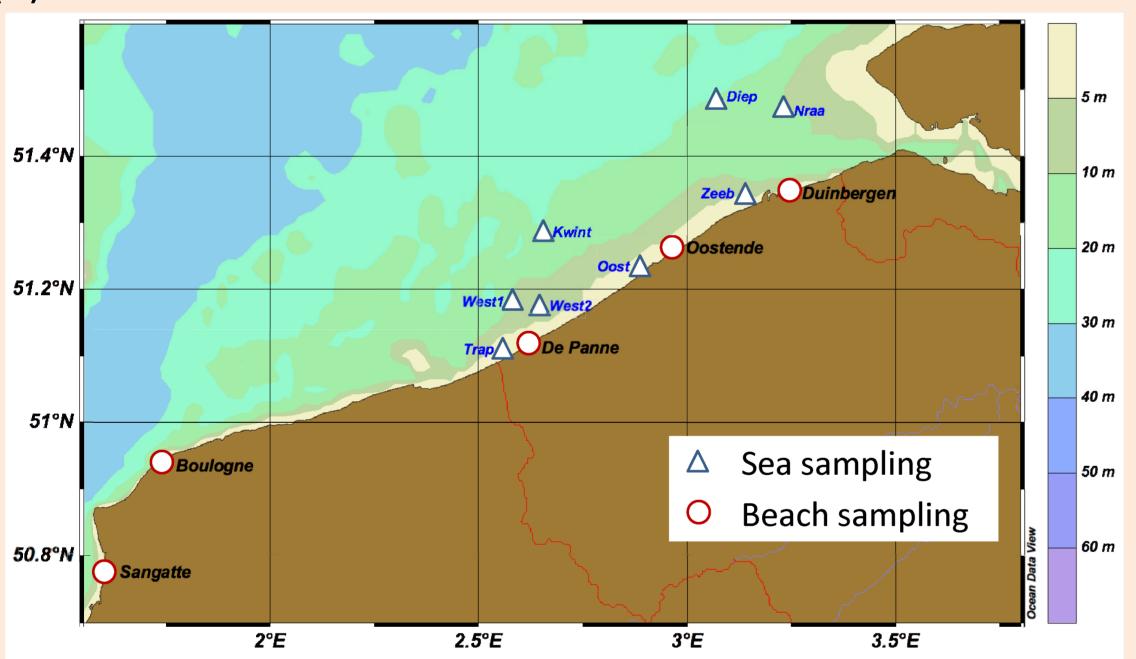


Figure 2: Sampling points of sole larvae, postlarvae and juveniles along the Belgian-French coast. Sea sampling = beam trawl, benthic sledge and WP3 plankton net. Beach sampling = beam trawl on foot.

Fish size limit <10cm. Courtesy of A. Vanden Bavière.

METHODOLOGY

(1) ENVIRONMENTAL CHEMICAL HETEROGENEITY

Tool 1: Microchemistry: Otolith elemental composition

15 chemical elements (e.g., Na, Ba, Mn)

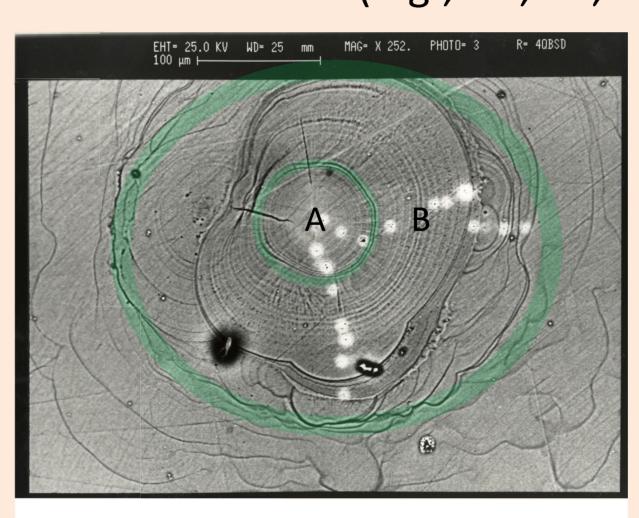


Figure 3: Sole postlarval otolith sampled for ICPMS.

A: natal signature (core of the otolith).

B: capture location signature (edge).

Sample holes in white.

Courtesy of A. Gueffen.

(2) LOCAL GENETIC SIGNATURE

Tool 2: SNPs for sole

Screen FishPopTrace's 426 SNPs* for 4 Atlantic populations

Illumina-Veracode Genotyping

200 outlier SNPs

Connectivity matrix

Origin assignment

Correlation analysis between matrices

Temporal stability analysis between old and new samples

Seascape (spatial) analysis

Population structure, relatedness and population origin

SCENARIO TESTING – IPCC PREDICTIONS IPCC

- River runoff, SST, wind & atmospheric pressure, behaviour
- Larval retention (Fig. 4)

Tool 3: Modelling of larval dispersal

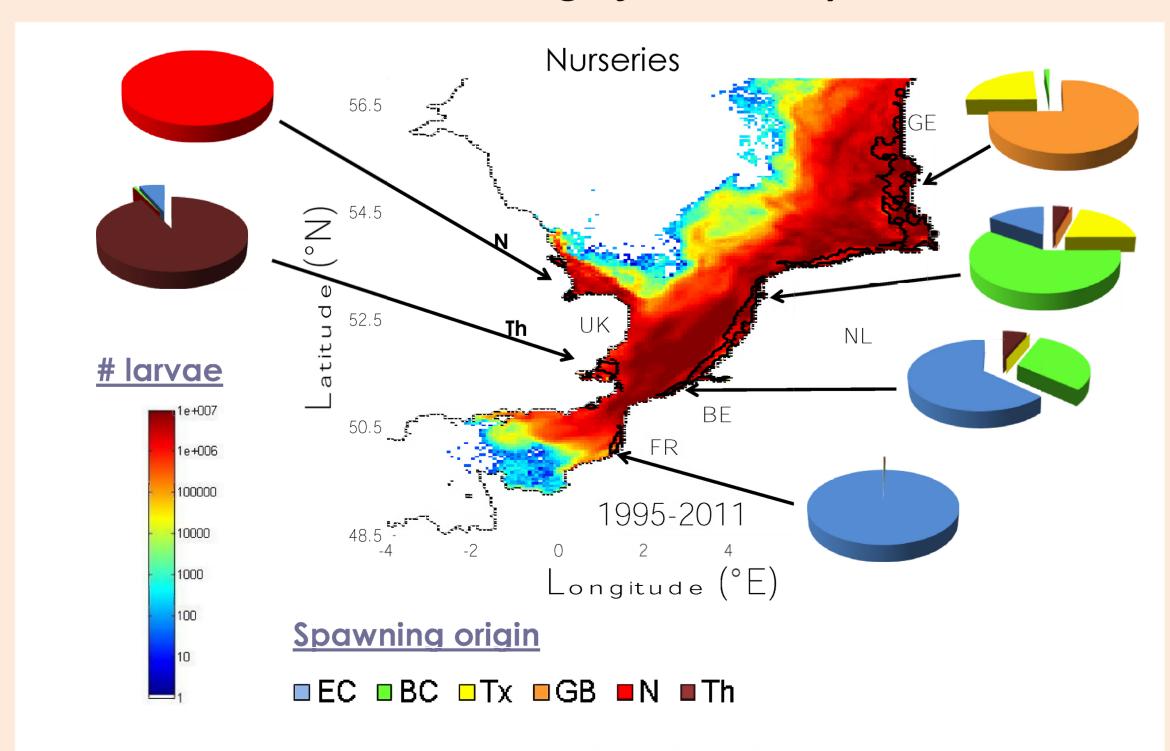


Figure 4: Larval abundance of North Sea sole at the end of larval transport obtained from a bio-hydrodynamic model (Lacroix et al. 2013).

Relative contributions of 6 spawning grounds: EC (Eastern English Channel),

Relative contributions of 6 spawning grounds: EC (Eastern English Channel), BC (off the Belgian coast), Tx (off Texel), GB (inner German Bight), N (Norfolk banks), Th (Thames estuary).

OUTCOMES

- **Individual tracing** with otolith microchemistry and genomics to understand:
 - → dispersal and connectivity
 - → recruitment
- Comparison of bio-hydrodynamic model with in situ data
 - → to assess the relative impacts of hydrodynamics, environment, behaviour and physiology
 - -> to improve the predictability of cohort strength
- Fisheries management at the subpopulation level because of:
 - -> potential collapse of less productive components
 - → the importance of biocomplexity for long term stability and sustainability of fish stocks

VOCABULARY

* Recruitment: the part of fish juveniles that have reached a sufficient length to enter the stock SNPs: Single Nucleotide Polymorphisms are molecular markers

Stock: the economically exploitable part of a population

BIBLIOGRAPHY

Lacroix G., Gregory E. Maes, Loes J. Bolle and Filip A.M. Volckaert. (2013) Modelling dispersal dynamics of the early life stages of a marine flatfish *Solea solea* (L.). Journal of Sea Research 84. pp. 13-25.

ICES Advice 2013, Book 6 – http://www.ices.dk/sites/pub/Publication%20Reports/Advice/2013/2013/sol-nsea.pdf

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