

This report not to be cited without prior reference to the council*

010

 THÜNEN
Digitalization sponsored
by Thünen-Institut

International Council for the
Exploration of the Sea



C.M. 1991/F:2

**REPORT OF THE WORKING GROUP ON MASS REARING OF
JUVENILE MARINE FISH TO THE MARICULTURE COMMITTEE OF ICES**

Gent, Belgium 31 August - 2 September, 1991.

This document is a report of a Working Group of the International Council for the Exploration of the Sea and does not necessarily represent the views of the Council. Therefore, it should not be quoted without consultation with the General Secretary.

***General Secretary
ICES, Palægade 2-4
DK-1261 Copenhagen K
Denmark.**

TABLE OF CONTENTS

1.	PARTICIPATION	2
2.	TERMS OF REFERENCE	3
3.	AGENDA	3
4.	EGG, LARVAL AND JUVENILE QUALITY	4
	Identification of quality criteria	4
	Development of research strategies	5
	Recommendations	6
5.	NUTRITION	7
	Recommendations	9
6.	STANDARDIZED PROCEDURES FOR EXPERI- MENTAL FRY PRODUCTION	9
	Recommendations	10
7.	HYGIENE STRATEGIES	11
	Recommendations	11
8.	NEXT MEETING	12

APPENDIX I. List of participants and addresses.

APPENDIX II. Bibliography on nutrition and histology.

APPENDIX III. I.C.E.S. Study on (n-3) HUFA Requirements
in Marine Fish Larvae (draft).

1. PARTICIPATION

The Working Group convened its fourth meeting at Gent, Belgium on August 31 - 2 September, 1991. Members attending the meeting were:

Belgium: P. Sorgeloos. Denmark: J. G. Støttrup. Canada: J.A. Brown, G. Goff, K. Waiwood. Faroe Islands: I. Fjallstein. France: B. Chatain. Germany: G. Quantz. Norway: D. Danielssen, I. Holmefjord, I. Huse, E. Kjørsvik, I. Lein, A. Mangor-Jensen, K. Naas, Y. Olsen, G. Rosenlund. Portugal: P. Pausao-Ferreria. Spain: J. Iglesias, I. Martinez, G. Minkoff, J.B. Peleteiro. Sweden: P.-O. Larsson. UK: B.R. Howell.

During 1990, several members attended by invitation the World Aquaculture Society annual meeting in Halifax, Canada, where a Larviculture Task Force within the WAS

chaired by Patrick Sorgeloos (Belgium) and David Bengtson (USA) was formed. Several WAS/LTF members also attended part of this meeting. Thus, apart from the ICES members, the following were present:

Belgium: T. Bosteels, Ph. Dhert, A. Komis, P. Lavens, Ph. Léger, I. Roelants, G. Van Stappen, L. Verdonck, F. Volckaert. Denmark: N.E. Poulsen. France: S. Kaushik. Germany: H. Segner. Greece: G.M. Robbins, E. Sweetman, J. Sweetman. Israel: Y. Barr, L. Samuel, A. Tandler, O. Zmora. Japan: K. Muroga, M. Tanaka. Mexico: A. Abreu Gobrois. Netherlands: J. Verreth. Norway: G. Adoff, A. Folkvord. Scotland: J. Dye, J.R. Sargent. Singapore: T.J. Lam, C.L. Lim, J. Walford. S.Africa: T. Hecht. Sweden: J. Pickova. Taiwan: H.-Y. Chien. Thailand: D. Fegan, P. Menasveta, S. Piyatiratitivorakul, J.-F. Rees. USA: H. Ako, D. Bengtson, J.G. Holt, S. Kraul, J.A. Tellock.

See Appendix I for addresses.

I. Huse, Norway, (chairman) and J.G. Støttrup, Denmark, kindly served as rapporteurs for the meeting.

2. TERMS OF REFERENCE

The ICES Working Group on Mass Rearing of Juvenile Marine Fish met to work according to the following terms of reference (ICES C.M. 1990/F:65):

- a) prepare a report describing standardized procedures and conditions for experimental fry production of turbot and sea bass as model species, including criteria in addition to growth and survival, for the evaluation of the quality of eggs, larvae and juveniles;
- b) describe nutritional requirements of marine fish, primarily for fatty acids and amino acids and collect information on the function of individual compounds in the organisms;
- c) advice on alternative strategies to the use of antibiotics in the control of microflora in culture systems.

3. AGENDA

During the meeting, three plenary sessions and one concurrent group session were held with the following subjects covered:

- A) Egg, larval and juvenile quality (E. Kjørsvik, Norway)
- B) Nutrition (P. Sorgeloos, Belgium and J. Verreth, Holland)
J. Sargent: Lipid metabolism
S. Kaushik: Protein metabolism

- C) Standardized procedures and conditions for experimental fry production of model species. (B. Chatain, France and J. G. Støttrup, Denmark)
- D) Hygiene strategies (G. Minkoff, Spain)

In the first two plenary sessions each topic was introduced by 1-2 short presentations, followed by discussion. The nutrition session also included two invited speakers on the main topics; lipid and protein metabolism.

Each subject was discussed in more detail in smaller groups during the concurrent sessions.

The recommendations were discussed and revised during the final plenary session.

4. EGG, LARVAL AND JUVENILE QUALITY

Convener: Elin Kjørsvik

Rapporteurs: Bari Howell, Kenneth Waiwood

Considerable variability in the performance of eggs and larvae is observed during the mass rearing of marine fish. Much of this variability may be attributable to egg quality defined as *the potential of the egg at the completion of fertilization to produce viable fry*. It will consequently embrace any effects associated with the quality of sperm, a subject that has been studied little in marine fish but one that can not be ignored.

There is an urgent need among practitioners to identify characteristics of eggs and larvae that reflect their quality as well as to improve our understanding of the mechanisms involved in generating the observed variability. The Working Group consequently recognized two important aspects for future work:

- I The requirement for making simple, rapid predictions of subsequent performance of eggs and larvae.
- II The development of strategies for the investigation of factors and mechanisms that influence egg quality.

I. IDENTIFICATION OF QUALITY CRITERIA

Ia Eggs

The following parameters have been or may prove to be useful indicators of egg quality in several marine species:

- fertilization rate
- hatching rate
- buoyancy

- cleavage pattern (morphology)
- mortality rates (at different stages)
- relative developmental rates
- turgor pressure (egg hardness)
- chorion appearance / fertilization process
- oil globule distribution
- opacity
- 'stress' tests (eg. mechanical shock)

Of these, it was considered that fertilization and hatching rates, buoyancy and cleavage patterns offered the greatest potential for routine assessment and that these measurements should be included in all assessments of quality. Nevertheless, it was recognized that, because egg quality may vary in response to a variety of factors, there may not be a universal indicator of egg quality and that different parameters may need to be used according to species and procedures used.

Ib Larvae

It is important that larvae quality is assessed at a defined developmental stage. the onset of exogenous feeding would be an appropriate stage and is the last stage at which larval quality can realistically be attributed to maternal influence. This should not, however, preclude eventual studies of effects on later developmental stages.

The following parameters were identified as being worthy of further evaluation:

- deformities
- pigmentation
- survival
- growth
- synchrony in development
- development of stages with time
- yolk absorption rate
- disease resistance
- 'stress tests' (eg. salinity, air exposure)
- 'performance tests' (eg. behavior, response to stimuli)

II DEVELOPMENT OF RESEARCH STRATEGIES

IIa Factors affecting egg quality

The following broodstock-related factors were identified as potential determinants of egg quality:

- time within the spawning season (batch spawners)
- fish age
- nutrition
- stripping/handling/fertilization
- over-ripening

- induced ovulation /spawning (hormones)
- environmental conditions (eg. temperature, salinity, light)
- water quality (eg. O₂, NH₃)

I Ib Mechanisms and manifestations of egg quality

The investigation of the following factors may provide an insight into the mechanisms involved in the determination of egg quality:

- energy charge (ATP/ADP)
- time dependent changes in, for example, hormones (eg. thyroxine), enzymes and nutrients
- cell cycles (eg. regulation of cell cleavage)
- chromosome aberrations/karyotyping of chromosomes

It was stressed that the value of 'snap-shot' analyses were often limited and that more effort should be devoted to studies of dynamic processes during development.

I Ic Research priorities

The following priority areas for future research were identified:

1. Standardization of methods and criteria.

An effort should be made to link the empirical routine criteria to the results of more advanced scientific investigations in order to more fully understand the underlying mechanisms involved in the determination of egg quality.

2. Development of methods for determining the timing of ovulation and the post-ovulatory age of eggs. Such methods may be related to:

- rates of drinking / salt excretion
- ultrasonic observation
- ovulatory cycles
- refinement of methods for natural spawning

These studies would be facilitated by an understanding of the hormonal control mechanisms which could even lead to the development of 'fish-ovulation test kits'.

3. Sperm quality assessment which should also include effects of storage methods.

RECOMMENDATIONS

The following recommendations were proposed and accepted by the Working Group:

1. A standardized protocol for assessment of egg and larval quality should be devised.

2. The design and distribution of a proforma data sheet to record egg production methods, the characteristics of eggs at fertilization and their subsequent performance.
3. Research should be encouraged in the following areas:
 - a) broodstock management procedures
 - b) reproduction mechanisms of the fish
 - c) ovulation/over-ripening mechanisms
 - d) time dependent compositional changes during the egg and larval stages.

5. NUTRITION

Convener: Patrick Sorgeloos & Johan Verreth

Rapporteur: Patrick Sorgeloos & Dave Bengtson

There is an urgent need to develop an appropriate protocol for nutritional experiments in fish larviculture. In that respect the following proposal was worked out:

- a1) A standard live diet should be used consisting of:
 - ICES-reference Artemia (small nauplii with high content of n-3 HUFAs) as starter feed (to be selected and made available by the Artemia Reference Center, Belgium), followed by
 - Great Salt Lake (Utah, USA) Artemia enriched by a standard protocol using:
 - ICES-standard emulsion as external standard
 - local enrichment procedure as internal standard
- a2) Regarding the green-water technique and Brachionus culturing, each laboratory should use the procedure they feel appropriate for the species cultured. Applied procedures should be well documented
- a3) A protocol should be developed so that nutritional tests can be performed under standardized, optimal conditions, including at least:
 - removal of all remaining food at least once a day
 - identification of feeding levels
- a4) Experimental conditions need to be optimized in order to realize maximum growth for use as reference conditions in later dose-response studies
- a5) Experimental results should be evaluated in relation to the maximum growth potential and in relation to a low growth under known diet-deficient conditions

- a6) The following parameters should be considered for evaluating the experimental results:
- survival
 - size of the fish, e.g. length, dry weight and any other mass-related parameter
 - stress resistance
 - size variation
 - time and size until certain developmental stage, such as gastric pH change, metamorphosis, pigmentation, organ development
 - RNA/DNA ratio
- a7) The need is expressed that a literature search should be conducted on the technology, formulation and use of standard reference inert diets for larvae. Dr. J. Walford has accepted to take up this responsibility.
- b1) The procedure for quantitative (n-3) HUFA analysis as prepared by Ph. Léger has been improved since the Palavas WG-meeting, will be further amended to exclude the use of carcinogens (benzene) and will be submitted for final evaluation to J.Sargent. The WG recommends that this method should be proposed for adoption as a standard ICES-procedure for (n-3) HUFA analysis.
- b2) Regarding the (n-3) HUFA requirement study proposed at the WG meeting in Vigo (1989) experimental results are available for sea bass, sea bream, turbot, striped bass, red drum, and summer flounder. It appears that best results are obtained with the medium (n-3) HUFA emulsion. Sometimes better results were obtained with high (n-3) HUFA emulsion, although analytical data show that enrichment levels obtained with medium may equal those obtained with high, which indicated that enrichment success might have been different. It is therefore essential along with the biological tests, analysis for the enriched prey is taken into consideration. The WG recommends the experimental results should be compiled and proposed in a final report (draft as Appendix II).
- b3) Since it appears from the previous study that much variability exists in (n-3) HUFA enrichment success with *Brachionus* and *Artemis*, even when using the same enrichment emulsions, and the present standard procedures, it is important to estimate present variability in (n-3) HUFA enrichment procedures. In this respect and in order to be able to advise optimal and standard enrichment procedures, an enrichment intercalibration exercise should be performed using an ICES-reference emulsion (made available by the Artemia Reference Center, Belgium) and Great Salt Lake (Utah, USA) *Artemia*. Protocols for experimental and analytical procedures will be prepared by J. Sargent and P. Sorgeloos.
- c) A bibliography on nutrition and histology has been compiled by D.A. Bengtson.

RECOMMENDATIONS

The following recommendations were proposed and accepted by the Working Group:

1. A protocol should be developed so that nutritional tests can be performed under standardized, optimal conditions.
2. It is recommended that a reference diet should be included in nutritional studies and this should consist of:
 - ICES-reference Artemia as starter feed (to be selected and made available by the Artemia Reference Center, Belgium), followed by
 - Great Salt Lake (Utah, USA) Artemia enriched by a standard protocol (to be developed), using:
 - ICES-standard emulsion as external standard
 - local enrichment procedures as internal standard
3. Results should be evaluated in terms of both maximal growth and some low-growth reference
4. Experimental results should be evaluated following well-defined criteria
5. To standardize (n-3) HUFA enrichment procedures, an enrichment intercalibration exercise should be performed.
6. A literature search should be conducted on the technology, formulation and use of standard reference inert diets for larvae (J. Walford).

6. STANDARDIZED PROCEDURES FOR EXPERIMENTAL FRY PRODUCTION

Conveners & rapporteurs: Beatrice Chatain & Josianne G. Støttrup

It was decided that the standardized system should serve as a reference protocol rather than a rigid experimental procedure. That is, the protocol should ascribe to each physical parameter a certain value and quality control within which effects of variation are negligible. Where actual experiments deviate from this protocol, it would then serve as a reference protocol.

The protocol could contain certain general laboratory procedures in sufficient detail to enable an operator to carry them out. It should be intended for experience biologists and laboratory staff, familiar with these types of experiments.

The following protocol was proposed and accepted by the Working Group for the standardization of procedures for experimental fry production of turbot and sea bass:

PROTOCOL

1. **Introductory information**
 - water quality; description of water treatment and analytical procedures
 - egg and larval quality; how to assess the egg and larval quality

2. **Method**
 - definitions
 - principles of rearing methods
 - replicates and controls
 - experimental design
 - description of experimental parameters
 - experimental data requirements
 - description of experimental procedures
 - conditions for judging the validity of the experiment

3. **Report**
 - information to be included in the report

B. Chatain, France provided information on sea bass rearing techniques as a basis for discussion and J.G. Støttrup, Denmark that on turbot. Due to the complexity of the subject and the short time available, many important details were not discussed, nor was there time to work out a comprehensive draft of the protocol at the meeting. The two reports on sea bass and turbot will therefore be compiled by each of the two conveners and sent to the WG members for further comments before the final draft can be submitted. At the earliest this may be for the ICES Statutory Meeting, 1992.

It was proposed that the final draft be submitted as an ICES Cooperative Research Report.

RECOMMENDATIONS

The following recommendations were proposed and accepted by the Working Group:

1. **Research priorities should include:**
 - a) the elucidation of the role of algae in rearing systems.
 - b) an assessment of the effects of the quality and quantity of light on larval performance (including effects of tank color/reflectivity)
 - c) the development of an empirical feeding model to provide a basis for hatchery feeding strategies

2. Composition, source and shelf-life of ingredients and storage conditions when commercial products are used in nutritional studies, should be provided by the manufacturers.
3. Research reports should include starvation (100%) times in clear water under the same experimental conditions as control groups.

7. HYGIENE STRATEGIES

Convener: Gidon Minkoff
Rapporteur: Øivind Bergh

RECOMMENDATIONS

The following recommendations were proposed and accepted by the Working Group:

- A. Marine finfish hatcheries are recommended to take the following measures:
 1. Disinfect eggs, employing methods adapted to the species in question
 2. Divide the hatchery into production zones, physically separated by infection barriers. Movement of material should be carried out down the production line. Duplicate facilities are advisable.
 3. Periodic shut-down of the plant should be carried out in order to disinfect the facilities.
 4. As the live food is seen as a major vector of contaminants, it is advisable to use methods such as rinsing to reduce the associated bacterial abundance.
 5. For monitoring bacterial growth, florescence microscopy is recommended if possible, otherwise marine agar and TCBS.
- B. There is a need for research on:
 - 6a. Bacterial populations associated with different life stages under different conditions
 - 6b. Interactions between larvae and associated bacteria including both pathogens and probiotics, taking into account the nutritional condition of the larvae, as well as their physical rearing conditions.
 - 6c. Procedures to improve system stability with regards to bacterial populations
 - 6d. The development of the immune system in early life stages.

8. NEXT MEETING

The Working Group on Mass Rearing of Juvenile Marine Fish recommends that the group should continue its work and meet in Bergen on June 25-26, 1993 with Ingvar Huse as Chairman.

The following terms of reference were suggested by the group for the Working Group meeting in 1993. The group should meet to work towards the establishment of:

- a) a protocol for standardized monitoring of egg and larval quality
- b) an inter-laboratory investigation of egg and larval quality
- c) a protocol for hygiene procedures in rearing systems
- d) a protocol for standard nutrition research taking into account data available on the performance of the proposed standard live diet and experimental procedure as well as the results of the (n-3) HUFA enrichment intercalibration exercise.

Furthermore, the group should meet to:

- e) compile information on standard inert reference diets.

APPENDIX I

List of participants from ICES countries

Belgium

Bosteels, Thomas
Laboratory of Aquaculture &
Artemia Reference Center
University of Ghent
Rozier 44
9000 Gent

Dhert, Ph.
Laboratory of Aquaculture &
Artemia Reference Center
University of Ghent
Rozier 44
9000 Gent

Komis, Antonios
Laboratory of Aquaculture
& Artemia Reference Center
University of Ghent
Rozier 44
9000 Gent

Lavens, Patrick
Laboratory of Aquaculture &
Artemia Reference Center
University of Ghent
Rozier 44
9000 Gent

Léger, Philippe
Artemia Systems NV/SA
Wiedauwkaai 79
9000 Gent

Roelants, I.
Laboratory of Ecology and Aquaculture
Catholic University of Leuven
Naamsestraat 59
B-3000 Leuven

Sorgeloos, Patrick
Laboratory of Aquaculture &
Artemia Reference Center
University of Ghent
Rozier 44, 9000 Gent

Van Stappen, G
Laboratory of Aquaculture
& Artemia Reference Center
University of Ghent
Rozier 44
9000 Gent

Verdonck, Linda
Laboratory of Microbiology
University of Ghent
KL Ledeganckstraat 35
9000 Gent

Volckaert, F.
Laboratory of Ecology
and Aquaculture
Catholic University
of Leuven
Naamsestraat 59
3000 Leuven

Denmark

Poulsen, Niels Eg
Danish Institute of Fisheries
and Marine Research
North Sea Center
P.O. Box 101
9850 Hirtshals

Støttrup, Josianne
Danish Inst. for Fisheries
and Marine Research
North Sea Center
P.O.Box 101
9850 Hirtshals

Canada

Brown, Joseph A
Ocean Science Center
Memorial University
of Newfoundland
St Johns
New Foundland
AIL 557

Goff, Greg
Fisheries Resource Development Ltd
2021 Brunswick St., Suite 317
Halifax
Nova Scotia B3K 2Y5

Waiwood, Kenneth
Biological Station
St. Andrews
New Brunswick
EOG 2X0 Canada

Faroe Islands

Fjallstein, Ingvarð
Fiskirannsóknarstovan
P.O. Box 3051
Noatun
Fr-100 Torshavn

France

Chatain, B.
Gie Recherche Aquacole
Station Expérimentale d'Aquaculture
IFREMER
route de Maguelone
34250 Palavas-les-Flots

Kaushik, S
Laboratoire de Nutrition
des Poissons
Station d'Hydrobiologie INRA
64310 St. Pée-sur-Nivelle

Person-Le Ruyet, J
Centre de Brest
IFREMER
B.P. 70
29280 Plouzané

Germany

Quantz, G.
Institut für Hydrobiologie und
Fishereiwissenschaft
Olbersweg 24
W-2000 Hamburg 50

Segner, Helmuth
Zoologie II
Universität Karlsruhe
Kaiserstrasse 12
7500 Karlsruhe

Netherlands

Verreth, Johan
Department of Fish Culture
and Fisheries
Wageningen Agricultural University
P.O.Box 338
6700 AH Wageningen
The Netherlands

Norway

Adoff, Grethe
Sea Farm AS
5419 Fitjar

Danielssen, Didrik
Institute of Marine Research
Flødeigen Marine Research Station
4817 His

Folkvord, Arild
Fr. Stangsv. 3
5032 Minde

Holmeffjord, Ivar
Institute of Aquaculture Research
AKVAFORSK
6600 Sunndalsøra

Huse, Ingvar
Austevoll Aquaculture Research Station
Institute of Marine Research
5392 Sterebo

Kjersvik, Elin
Dept. of Zoology
University of Trondheim
7055 Dragvoll-Trondheim

Lein, Ingrid
Institute of Aquaculture Research.
AKVAFORSK
6600 Sunndalsra

Mangor-Jensen, Anders
Research Station Austevoll
Institute of Marine Research
5392 Storebc

Naas, Kjell
Austevoll Aquaculture Research Station
Institute of Marine Research
5392 Storebc

Olsen, Yngvar
Centre of Aquaculture
SINTEF
7034 Trondheim

Rosenlund, Grethe
BP Nutrition Aquaculture
Research Centre
P.O.Box 532
4001 Stavanger

Portugal

Pousao-Ferreira, Pedro
Instituto Nacional de
Investigacao das Pescas
Av. 5 de Outubro S/N
8700 Olh

Spain

Iglesias, Jose
Instituto Espanol de Oceanografia
Apdo 1552
36280 Vigo

Martinez, I.
Instituto Espanol de
Oceanografia
Laboratorio de Santander
Apdo 240
39080 Santander

Minkoff, G.
Tinamenor S.A. Pesues
Cantabria

Peleteiro, Jose B
Instituto Espanol de Oceanografia
Apdo 1522
36280 Vigo
Pontevedra

Sweden

Larsson, Per-Olov
Institute of Marine Research
P.O. Box 4
45300 Lysekil

Pickova, Jana
Institute of Marine Research
P.O. Box 4
45900 Lysekil

United Kingdom

Bromage, Niall
Institute of Aquaculture
University of Stirling
Stirling
FK9 4LA
Scotland

Dye, John
Fish Industry Authority
Marine Farming Unit
Ardtoe, Acharacle
Argyll, PH36 4LD

Howell, B. R.
Fisheries Laboratory
MAFF
Benarth Road
Conwy
Gwynedd LL32 8UB
Wales

United States of Amerika

Ako, Harry
Dept of Environmental Biochemistry
University of Hawaii
1800 East West Rd
Honolulu
HI 96822

Bengtson, David
Dept. of Zoology
University of Rhode Island
Kingston, RI 02881

Holt, G Joan
Marine Sciences Institute
The University of Texas at Austin
P.O.Box 1267
Port Aransas
TX 78373

Tellock, Jeff A.
Provesta Corporation
93-D Philips Research Center
Bartlesville
OK 74004

Appendix II

BIBLIOGRAPHY ON NUTRITION AND HISTOLOGY (Draft No. 1) D.A. Bengtson

Prepared for:

ICES Working Group on Mass Rearing of Juvenile Marine Fish
Meeting at Ghent, Belgium, 31 August - 2 September 1991.

- Albertini-Berhaut, J. 1987. L'intestin chez les Mugilidae (Poissons; Teleosteens) a differentes etapes de leur croissance. I. Aspects morphologiques et histologiques. J. Appl. Ichthyol. 3:1-12.
- Al-Maghazachi, S.J. 1983. Histological and histochemical studies on gut development and physiology of larval turbot, Scophthalmus maximus (L.). Ph.D. Dissertation, Liverpool Polytechnic, 237 pp.
- Appelbaum, S., H. Segner and V. Storch. 1986. Electron microscopic study on the influence of different nutritional conditions on liver and white trunk muscles of whitefish (Coregonus lavaretus) larvae. Zool. Anz. 217:54-64.
- Bell, M.V., R.J. Henderson, B.J.S. Pirie and J.R. Sargent. 1985. Effects of dietary polyunsaturated fatty acid deficiencies on mortality, growth and gill structure in the turbot, Scophthalmus maximus. J. Fish Biol. 26:181-191.
- Blaxter, C.H.S., D. Danielssen, E. Moksness and V. Oiestad. 1983. Description of the early development of the halibut Hippoglossus hippoglossus and attempts to rear the larvae past first feeding. Mar. Biol. 74:99-107.
- Burkhardt-Holm, R. Eckmann and V. Storch. 1989. Schädigung des Darmepithels von Coregonenlarven (Coregonus fera) durch Artemia-Fütterung: Eine bakterielle Infektion. J. Appl. Ichthyol. 5:2-11.
- Connes, R. and K. Benhalima. 1984. Ultrastructure de l'intestin du Loup Dicentrarchus labrax L. au cours du developpement larvaire. Bull. Soc. Zool. Fr. 109:19-33.
- Cousin, J.C.B. and F. Baudin-Laurencin. 1985. Morphogenese de l'appareil digestif et de la vessie gazeuse du turbot, Scophthalmus maximus L. Aquaculture 47:305-319.
- Cousin, J.C.B., G. Balouet and F. Baudin-Laurencin. 1986. Alterations histologiques observees chez des larves de turbot (Scophthalmus maximus L.) en elevage intensif. Aquaculture 52:173-189.
- Cousin, J.C.B., F. Baudin-Laurencin and J. Gabaudan. 1987. Ontogeny of enzymatic activities in fed and fasting turbot, Scophthalmus maximus L. J. Fish Biol. 30:15-33.

Dabrowski, K. 1989. Formulation of a bioenergetic model for coregonine early life history. *Trans. Am. Fish. Soc.* 118:138-150.

Eckmann, R. 1985. Histopathological alterations in the intestine of whitefish (Coregonus sp.) larvae reared on zooplankton from Lake Constance. *Dis. aquat. Org.* 1:11-17.

Eckmann, R. 1987. Pathological changes in the midgut epithelium of grayling, Thymallus thymallus L., larvae reared on different kinds of food, and the relation to mortality and growth. *J. Fish Dis.* 10:91-99.

Ehrlich, K.F., J.H.S. Blaxter and R. Pemberton. 1976. Morphological and histological changes during the growth and starvation of herring and plaice larvae. *Mar. Biol.* 35:105-118.

Ferraris, R.P., J.D. Tan and M.C. De la Cruz. 1987. Development of the digestive tract of milkfish, Chanos chanos (Forsk.) : histology and histochemistry. *Aquaculture* 61:241-257.

Gabaudan, J. 1984. Posthatching morphogenesis of the digestive system of striped bass. Ph.D. Dissertation, Auburn University, Auburn, Alabama, USA.

Georgopoulou, U., M.F. Sire and J.M. Vernier. 1986. Absorption intestinale des proteines sous forme macromoleculaire et leur digestion chez la Truite arc-en-ciel. Etude ultrastructurale et biochimique en relation avec la premiere prise de nourriture. *Can. J. Zool.* 64:1231-1240.

Glazebrook, J.S., M.P. Heasman and S.W. de Beer. 1990. Picorna-like viral particles associated with mass mortalities in larval barramundi, Lates calcarifer Bloch. *J. Fish Diseases* 13:245-249.

Govoni, J.J. 1980. Morphological, histological and functional aspects of alimentary canal and associated organ development in larval Leiostomus xanthurus. *Rev. Can. Biol.* 39:69-80.

Govoni, J.J., G.W. Boehlert and Y. Watanabe. 1986. The physiology of digestion in fish larvae. *Env. Biol. Fish.* 16:59-77.

Grizzle, J.M. and M.R. Curd. 1978. Posthatching histological development of the digestive system and swim bladder of logperch, Percina caprodes. *Copeia* 1978(3):448-455.

Il'ina, I.D. and V.I. Turetskiy. 1987. Development of the digestive function in fishes. *Vopr. Ikhtiolog.* 1987(5):835-843.

Iwai, T. 1967a. The comparative study of the digestive tract of teleost larvae - I. Fine structure of the gut epithelium in larvae of ayu. *Bull. Jap. Soc. Sci. Fish.* 33:480-496.

Iwai, T. 1967b. The comparative study of the digestive tract of teleost larvae - II. Ciliated cells of the gut epithelium in pond

- smelt larvae. Bull. Jap. Soc. Sci. Fish. 33:1116-1119.
- Iwai, T. 1968a. The comparative study of the digestive tract of teleost larvae - V. Fat absorption in the gut epithelium of goldfish larvae. Bull. Jap. Soc. Sci. Fish. 34:973-978.
- Iwai, T. 1968b. Fine structure and absorption patterns of intestinal epithelial cells in rainbow trout alevins. Z. Zellforsch. mikroskop. Anat. 91:366-379.
- Iwai, T. 1969. Fine structure of gut epithelium cells of larval and juvenile carp during absorption of fat and protein. Arch. Histol. Jap. 30:183-199.
- Iwai, T. and M. Tanaka. 1968a. The comparative study of the digestive tract of teleost larvae - III. Epithelial cells in the posterior gut of halfbeak larvae. Bull. Jap. Soc. Sci. Fish. 34:44-48.
- Iwai, T. and M. Tanaka. 1968b. The comparative study of the digestive tract of teleost larvae - IV. Absorption of fat by the gut of halfbeak larvae. Bull. Jap. Soc. Sci. Fish. 34:871-875.
- Kjorsvik, E., T. van der Meeren, H. Kryvi, J. Arnfinnson and P.G. Kvenseth. 1991. Early development of the digestive tract of cod larvae, Gadus morhua L., during start-feeding and starvation. J. Fish Biol. 38:1-15.
- Loewe, H. and Eckmann, R. 1988. The ontogeny of the alimentary tract of coregonid larvae: normal development. J. Fish Biol. 33:841-850.
- Louw, E. and M.J. O'Toole. 1977. Larval development of Sardinops ocellata (Pisces:Clupeidae). Ann. S. Afr. Mus. 72:125-145.
- Mahr, K., M. Grabner, R. Hofer and H. Moser. 1983. Histological and physiological development of the stomach in Coregonus sp. Arch. Hydrobiol. 98:344-353.
- Mitchell, L.G., J.G. Nickum and M.T. Long. 1986. Histochemical localization of some digestive enzymes in larval walleyes. Prog. Fish Cult. 48:279-281.
- Moshal'kova, M.I. 1988. Anatomical-histological and functional peculiarities of development of the intestine in the round goby, Neogobius melanostomus, a species with direct type of development. Vopr. Ikhtiol. 1988(5):812-827.
- O'Connell, C.P. 1976. Histological criteria for diagnosing the starving condition in early post yolk sac larvae of the northern anchovy, Engraulis mordax Girard. J. Exp. Mar. Biol. Ecol. 25:285-312.
- O'Connell, C.P. 1980. Percentage of starving northern anchovy,

Engraulis mordax, larvae in the sea as estimated by histological methods. Fish. Bull. 78:475-489.

O'Connell, C.P. 1981. Development of organ systems in the northern anchovy, Engraulis mordax, and other teleosts. Amer. Zool. 21:429-446.

O'Connell, C.P. and P.A. Paloma. 1981. Histochemical indications of liver glycogen in samples of emaciated and robust larvae of the northern anchovy, Engraulis mordax. Fish. Bull. 79:806-812.

Poston, H.A., G.F. Combs, Jr. and L. Leibovitz. 1976. Vitamin E and selenium interrelations in the diet of Atlantic salmon (Salmo salar): gross, histological and biochemical deficiency signs. J. Nutr. 106:892-904.

Rombout, J.H.W.M., C.H.J. Lamers and J.G. Hanstede. 1978. Enteroendocrine APUD cells in the digestive tract of larval Barbus conchionius (Teleostei, Cyprinidae). J. Embryol. Exp. Morphol. 47:121-135.

Rombout, J.H.W.M., H.W.J. Stroband and J.J. Taverne-Thiele. 1984. Proliferation and differentiation of intestinal epithelial cells during development of Barbus conchionius (Teleostei, Cyprinidae). Cell Tissue Res. 236:207-216.

Segner, H. 1985. Influence of starvation and refeeding with different diets on the hepatocyte ultrastructure of juvenile Siganus guttatus Bloch (Teleostei: Siganidae). Zool. Anz. 214:81-90.

Segner, H., P. Burkhardt, E.M. Avila, J.V. Juario and V. Storch. 1987. Nutrition-related histopathology of the intestine of milkfish Chanos chanos fry. Dis. aquat. Org. 2:99-107.

Segner, H. and J.V. Juario. 1986. Histological observations on the rearing of milkfish, Chanos chanos, fry using different diets. J. Appl. Ichthyol. 2:162-173.

Segner, H., R. Rosch, H. Schmidt and K.J. von Poeppinghausen. 1989. Digestive enzymes in larval Coregonus lavaretus L. J. Fish Biol. 35:249-263.

Sinha, G.M. 1976. Comparative morphology, anatomy and histology of the alimentary canal of an Indian freshwater major carp, Labeo calbasu (Hamilton) during the different life-history stages in relation to food and feeding habits. Anat. Anz. 139,S:348-362.

Sinha, G.M. 1979. Histochemical localization of alkaline and acid phosphatases in the alimentary tract of hatchling of a teleost fish, Cirrhinus mrigala (Hamilton). Mikroskopie 35:101-107.

Sinha, G.M. and S.K. Moitra. 1975a. Morpho-histology of the intestine in a freshwater major carp, Cirrhinus mrigala (Hamilton)

during the different life-history stages in relation to food and feeding habits. *Anat. Anz.* 137:395-407.

Sinha, G.M. and S.K. Moitra. 1975b. Functional morpho-histology of the alimentary canal of an Indian freshwater major carp, Labeo rohita (Hamilton) during its different life history stages. *Anat. Anz.* 138:222-239.

Sinha, G.M. and S.K. Moitra. 1976. Studies on the morpho-histology of the alimentary canal of freshwater fishes of India. Part 1. The alimentary canal of young Cirrhinus reba (Ham.) with a comparison with that of the adult in relation to food. *Vest. Ceskoslovenske spol. zool.* 40:221-231.

Smallwood, W.M. and M.B. Derrickson. 1933. The development of the carp, Cyprinus carpio. II. The development of the liver-pancreas, the islands of Langerhans, and the spleen. *J. Morphol.* 55:15-27.

Smallwood, W.M. and M.L. Smallwood. 1931. The development of the carp, Cyprinus carpio. I. The larval life of the carp, with special reference to the development of the intestinal canal. *J. Morphol. Physiol.* 52:217-231.

Storch, V. and J.V. Juario. 1983. The effect of starvation and subsequent feeding on the hepatocytes of Chanos chanos (Forsskal) fingerlings and fry. *J. Fish Biol.* 23:95-103.

Storch, V., W. Stahlin and J.V. Juario. 1983. Effects of different diets on the ultrastructure of hepatocytes of Chanos chanos fry (Chanidae: Teleostei): an electron microscopic and morphometric analysis. *Mar. Biol.* 74:101-104.

Stroband, H.W.J. 1977. Growth and diet dependent structural adaptations of the digestive tract in juvenile grass carp (Ctenopharyngodon idella Val.). *J. Fish Biol.* 11:167-174.

Stroband, H.W.J. and K. Dabrowski. 1981. Morphological and physiological aspects of the digestive system and feeding in freshwater fish larvae. Pp. 355-374 in *Nutrition des poissons* (M. Fontaine, ed.), Editions du CNRS, Paris.

Stroband, H.W.J. and A.G. Kroon. 1981. The development of the stomach in Clarias lazera and the intestinal absorption of protein macromolecules. *Cell Tissue Res.* 215:397-415.

Stroband, H.W.J., H. van der Meer and L.P.M. Timmermans. 1979. Regional functional differentiation in the gut of the grass carp, Ctenopharyngodon idella (Val.). *Histochemistry* 64:235-249.

Stroband, H.W.J. and F.H. van der Veen. 1981. Localization of protein absorption during transport of food in the intestine of the grass carp (Ctenopharyngodon idella Val.). *J. Exp. Zool.* 218:149-156.

Strussmann, C.A. and F. Takashima. 1989. PNR, histology and morphometry of starved pejerrey Odontesthes bonariensis larvae. Nippon Suisan Gakkaishi 55:237-246.

Strussmann, C.A. and F. Takashima. 1989. Effects of temperature upon survival and histological changes of starved pejerrey Odontesthes bonariensis larvae. Nippon Suisan Gakkaishi 55:247-254.

Strussmann, C.A. and F. Takashima. 1990. Hepatocyte nuclear size and nutritional condition of larval pejerrey, Odontesthes bonariensis (Cuvier et Valenciennes). J. Fish Biol. 36:59-65.

Szlaminska, M. 1980. A histochemical study of digestive enzymes in pike larvae. Fish. Mgmt. 11:139-140.

Tanaka, M. 1969a. Studies on the structure and function of the digestive system in teleost larvae - I. Development of the digestive system during prelarval stage. Jap. J. Ichthyol. 16:1-9.

Tanaka, M. 1969b. Studies on the structure and function of the digestive system in teleost larvae - II. Characteristics of the digestive system in larvae at the stage of first feeding. Jap. J. Ichthyol. 16:41-49.

Tanaka, M. 1971. Studies on the structure and function of the digestive system in teleost larvae - III. Development of the digestive system during postlarval stage. Jap. J. Ichthyol. 18:164-174.

Tanaka, M. 1972a. Studies on the structure and function of the digestive system in teleost larvae - IV. Changes in the epithelium related to fat absorption in the anteromedian part of the intestine after feeding. Jap. J. Ichthyol. 19:15-25.

Tanaka, M. 1972b. Studies on the structure and function of the digestive system in teleost larvae - V. Epithelial changes in the posterior gut and protein absorption. Jap. J. Ichthyol. 19:172-180.

Tanaka, M., S. Kawai and S. Yamamoto. 1972. On the development of the digestive system and changes in activities of digestive enzymes during larval and juvenile stages in ayu. Bull. Jap. Soc. Sci. Fish. 38:1143-1152.

Theilacker, G.H. 1978. Effect of starvation on the histological and morphological characteristics of jack mackerel, Trachurus symmetricus. Fish. Bull. 76:403-414.

Theilacker, G.H. 1986. Starvation-induced mortality of young sea-caught jack mackerel, Trachurus symmetricus, determined with histological and morphological methods. Fish. Bull. 84:1-17.

- Umeda, S. and A. Ochiai. 1973. On the development of the structure and function of the alimentary tract of the yellowtail from the larval to the juvenile stage. Bull. Jap. Soc. Sci. Fish. 39:923-930.
- Umeda, S. and A. Ochiai. 1975. On the histological structure and function of digestive organs of the fed and starved larvae of the yellowtail, Seriola quinqueradiata. Jap. J. Ichthyol. 21:213-219.
- Verreth, J.A.J., E. Torreele, E. Spazier, A. van der Sluiszen, J.H.W.M. Rombout, R. Booms and H. Segner. In press. The development of a functional digestive system in the African catfish Clarias gariepinus (Burchell). J. World Aquacult. Soc.
- Vu, T.T. 1976. Etude du developpement du tube digestif des larves de Bar, Dicentrarchus labrax (L.). Arch Zool. Exp. Gen. 117:493-509.
- Vu, T.T. 1980. Etude histologique de l'epithelium du tube digestif du Bar, Dicentrarchus labrax (L.), au cours du developpement post-embryonnaire. Arch. Zool. Exp. Gen. 121:191-206.
- Vu, T.T. 1983. Etude histoenzymologique des activites proteasiques dans le tube digestif des larves et des adultes de Bar, Dicentrarchus labrax (L.). Aquaculture 32:57-69.
- Wang, Z. and F. Takashima. 1984. Histological changes in digestive organs of carp larvae during starvation. II. Liver and pancreas. Suisanzoshoku 32:44-53.
- Wang, Z., F. Takashima, M. Nomura and K. Sakai. 1983. Histological changes in digestive organs of carp larvae during starvation. I. Suisanzoshoku 31:134-142.
- Watanabe, Y. 1981. Ingestion of horseradish peroxidase by the intestinal cells in larvae or juveniles of some teleosts. Bull. Jap. Soc. Sci. Fish. 47:1299-1307.
- Watanabe, Y. 1982a. Ultrastructure of epithelial cells of the anteromedian intestine and the rectum in larvae and juvenile teleosts. Bull. Fac. Fish., Hokkaido Univ. 33:217-228.
- Watanabe, Y. 1982b. Intracellular digestion of horseradish peroxidase by the intestinal cells of teleost larvae and juveniles. Bull. Jap. Soc. Sci. Fish. 48:37-42.
- Watanabe, Y. 1984a. An ultrastructural study of intracellular digestion of horseradish peroxidase by the rectal epithelium cells in larvae of a freshwater cottid fish Cottus nozawae. Bull. Jap. Soc. Sci. Fish. 50:409-416.
- Watanabe, Y. 1984b. Morphological and functional changes in rectal epithelium cells of pond smelt during postembryonic development. Bull Jap. Soc. Sci. Fish. 50:805-814.

Watanabe, Y. 1984c. Postembryonic development of intestinal epithelium of masu salmon (Oncorhynchus masou). Bull. Tohoku Reg. Fish. Res. Lab. 46:1-14.

Watanabe, Y. 1985. Histological changes in the liver and intestine of freshwater goby larvae during short-term starvation. Bull. Jap. Soc. Sci. Fish. 51:707-709.

Watanabe, Y. and N. Sawada. 1985. Larval development of digestive organs and intestinal absorptive functions in the freshwater goby (Chaenogobius annularis). Bull. Tohoku Reg. Fish. Res. Lab. 47:1-10.

Yasunaga, Y. 1972. The development of the digestive gland of the plaice larva, Paralichthys olivaceus. Tokai-ku Suisan Kenkyujo, Kenkyu, Hokoku, No. 69:75-89.

Appendix III

I.C.E.S. Study on (n-3)HUFA Requirements in Marine Fish Larvae.

PHILIPPE LÉGER

Artemia Systems S.A., Wiedauwkaal 79, B-9000 Ghent, Belgium.

DAVE A. BENGTON

University of Kingston,.....,RI, U.S.A.

PATRICK SORGELOOS

Laboratory of Aquaculture and Artemia Reference Center, State University of Ghent, Rozier 44, B-9000 Ghent, Belgium.

Abstract

This paper summarizes the results obtained in the framework of a study launched and coordinated by I.C.E.S. (International Council for the Exploration of the Seas). This study aims at establishing requirement levels of the essential fatty acids (n-3)HUFA for the early life stages of marine aquaculture organisms. So far results have been reported for the following species: the giant prawn *Macrobrachium rosenbergii*, seabass *Dicentrarchus labrax*, seabream *Sparus aurata*, turbot *Scophthalmus maximus*, striped bass *Morone saxatilis*, and red drum *Sciaenops ocellatus*.

During its meeting in Vigo (1988, Spain) the I.C.E.S. working group on 'Early Life Stages of Marine Fish Larvae' recommended to '...better identify the qualitative and the quantitative (n-3)HUFA requirements in marine larvae...'. The term (n-3)HUFA refers to long chain highly unsaturated fatty acids, more particularly eicosapentaenoic acid (EPA or 20:5n-3) and docosahexaenoic acid (DHA or 22:6n-3). These fatty acids have an important structural role in the build up of biomembranes and they act on a number of biochemicals called 'eicosanoids' such as leukotrienes and prostaglandins. These molecules act as hormones involved in regulating several vital body functions such as the immune response mechanism and inflammatory processes. The essentiality of these fatty acids for young marine animals has been documented quite extensively the last years. Several publications evidence the qualitative dietary importance of these fatty acids for the larvae of several species of marine organisms. Establishing quantitative requirement levels for commercially important species would be very beneficial for improving and optimizing their larviculture. This is however a complex issue as quantitative dietary requirement levels may differ from species to species and may furthermore be affected by larval age, larval quality, environmental conditions, etc. . Provided these conditions are determined and standardized, still then establishing dietary requirement levels of f.ex. (n-3)HUFA is very difficult as today no reference compound larval diets -with (n-3)HUFA as the only variable- is available. Even when such diet would be available it might not be suitable as the first feeding stages of most marine fish larvae do not accept compound feeds as

a sole diet. For these reasons it was suggested that for the present I.C.E.S. study the commonly used feeding regime based on the use of the live food organisms *Brachionus plicatilis* and *Artemia nauplii* would be applied in a first step. The fatty acid profile of the rotifer *Brachionus plicatilis* and the nauplii of the brine shrimp *Artemia*, especially their content of the (n-3)HUFA, can be modified by applying the technique of enrichment (Léger et al. 1985, 1986; Watanabe et al. 1983). This way the content of (n-3)HUFA can significantly be increased from a few milligrams per gram dry weight to 50 milligrams and more (Léger et al. 1987). These high values are obtained by feeding the live food organisms (n-3)HUFA rich emulsions.

Materials and Methods

For the present study three emulsions have been formulated for the enrichment of the live food organisms. The emulsions had a different content of (n-3)HUFA : low, medium and high (see Table I).

Detailed instructions for rotifer and *Artemia* enrichment were provided by I.C.E.S. to the 14 participants of the study:

1. *Artemia* enrichment

- incubate the daily required amount of *Artemia* cysts (Great Salt Lake strain, 2 g per liter seawater) in natural or artificial seawater:

- salinity = 35 ppt
- temperature = 28°C +/- 1°C
- pH = 8 à 9 throughout hatching; eventually add sodium bicarbonate
- light = > 2000 lux
- aeration = fairly strong

- harvest and rinse the *Artemia* nauplii after 24 h incubation and transfer them to the enrichment vessel filled with filtered seawater at a density of 100 individuals per ml.

- mix (with kitchenblender) 0.3 g enrichment emulsion (per liter enrichment medium) for 30 seconds in a small volume of water and add to the enrichment vessel.

- maintain a temperature of 28°C +/- 1°C.

- ensure high oxygen levels (min. 4 ppm D.O.) by applying an airstone aeration.

- no illumination is required.

- a second ration of 0.3 g enrichment emulsion (per liter enrichment medium) is prepared and added after 16 h enrichment

(between 12 and 18 h).

- after a total enrichment period of 24 h the metanauplii are harvested and thoroughly rinsed; now they are ready for feeding to the larvae.

2. Rotifer enrichment

- harvest and rinse cultured rotifers on an immersed sieve (45 à 60 μm).

- gently transfer the rinsed rotifers into filtered pre-aerated seawater of 25 ppt salinity at a density of 500 rotifers per ml.

- maintain a temperature of $27^{\circ}\text{C} \pm 1^{\circ}\text{C}$.

- maintain a slight aeration as to keep all rotifers well suspended; avoid strong aeration in order to minimize clogging.

- the enrichment media are prepared as above for *Artemia*; instead of two times 0.3 g/l for *Artemia* enrichment two times 0.1 g/l are used for rotifer enrichment; a first ration is added when the rotifers are being transferred to the enrichment tank (t_{0h}) (start of enrichment) and the second ration is added after three hours (t_{3h}) incubation.

- after a total enrichment period of six hours (t_{6h}), gently harvest and rinse the enriched rotifers on an immersed sieve; avoid strong turbulence; now the enriched rotifers are ready for feeding.

By applying the above instructions the content of (n-3)HUFA in rotifers and *Artemia* is altered depending on the type of emulsion used: rotifers and *Artemia* enriched with the low, medium and high (n-3)HUFA emulsion contain respectively low, medium and high levels of (n-3)HUFA (example for *Artemia* see Figure 1 and 2).

Results

The detailed results obtained by the different participating institutes are being published elsewhere as individual papers. A summary of the first results are being presented hereunder.

1. *Dicentrarchus labrax*

Two different laboratories ran the requirement study on seabass (*Dicentrarchus labrax*) larvae.

One study (Corneillie et al., Zoological Institute, University of Leuven, Leuven, Belgium) was conducted on first feeding larvae which during a period of 40 days were offered rotifers and subsequently *Artemia* both enriched with the three emulsions containing

increasing levels of (n-3)HUFA.

A clear impact of the fatty acid composition of the feeds on larval survival was noted: survival after 60 days culturing was 3.3 % +/- 0.1 % in the low (n-3)HUFA group, 12.7 % +/- 0.6 % in the medium (n-3)HUFA group and 13.0 % +/- 0.1 % in the high (n-3)HUFA group. A massive mortality was noted between day 30 and day 60 in the larvae fed the low (n-3)HUFA enriched rotifers and *Artemia*. During this period many larvae in this treatment exhibited the symptoms of the so-called 'whirling disease'. No larvae from the medium and high (n-3)HUFA treatments were affected. Growth in terms of individual dry weight was also affected by the diet composition though significant differences appeared only from the third week onwards (see Fig.4). The same was observed for growth in terms of individual length. Besides survival and growth effects on the morphological development of the larvae were also studied. No relation was found between the diet composition and the occurrence of skeletal deformities, shortening of the opercula nor the degree of swimbladder inflation. A significant interaction was detected however between the (n-3)HUFA content of the diet and the development of the gall bladder: low dietary (n-3)HUFA levels appeared to induce hypertrophy of the gall bladder.

The second study (Martinez and Alcazar, Oceanographic Institute, Mar Menor, Murcia, Spain) was carried out with 35 d old *Dicentrarchus labrax* larvae which were fed the enriched *Artemia* preparations during 29 days. During this stage of larval development noticeable effects of dietary fatty acid content on survival are only observed between the low (n-3)HUFA treatment and both others (see Fig.5). Effects on growth were not significant.

Neither of the two studies carried out fatty acid analysis on the enriched live feed used in the experiment. Hence absolute requirements seabass larvae for n-3HUFA may not be drawn from the above studies. Taking into consideration the values as illustrated in Figures 1 and 2 one could conclude that estimated requirements would be about ... to ... mg n-3HUFA per gram dry feed offered.

2. *Sparus aurata*

Sofar results with seabream larvae are limited to those obtained by Martinez and Alcazar (cfr above). Results relative to the treatments are very comparable to those obtained by the same authors for seabass *D. labrax* (see above). Figure 6 shows again a massive mortality in the low (n-3)HUFA treatment and a relatively high survival in both other treatments.

Koven et al. carried out a two weeks experiment with ...days old seabream larvae fed *Artemia* nauplii enriched with soybean oil (no n-3HUFA) supplemented with increasing levels of SUPER SELCO (*Artemia* Systems S.A., Ghent Belgium; SUPER SELCO contains an equivalent concentration of n-3HUFA as the I.C.E.S. High emulsion). The supplementation of n-3HUFA in the *Artemia* increased survival in the bream larvae significantly though a levelling off could be noted atmg/g (see Fig....). On the contrary however, growth

continued to improve along with the level of n-3HUFA in the diet indicating that for growth requirements are above ...mg n-3HUFA per gram dry feed offered (see Figure....).

3. *Macrobrachium rosenbergii*

Two studies have been performed evaluating the effects of feeding newly hatched *M. rosenbergii* larvae a sole diet of *Artemia* nauplii enriched with the I.C.E.S. emulsions. A first study by Buzzi et al. (1989) carried out at the Sterling University - Tropical Prawn Unit- indicated that best results in terms of survival, growth and metamorphosis rate were achieved when the prawn larvae were fed the highest n-3HUFA containing diet (High-emulsion). The results were significantly better than when feeding the Medium enriched nauplii. Lowest culture performance was observed in the treatment receiving Low n-3HUFA *Artemia* (see Figures....). Fatty acid profiles of the enriched *Artemia* nauplii are given in Table.... From these results the authors concluded that for meeting the n-3HUFA requirements of *M. rosenbergii* larvae the diet offered should contain at leastmg n-3HUFA per gram dry weight.

In a second study Devresse et al. (1990) performed the same experiment at the *Artemia* Reference center (University of Ghent, Belgium). Their results indicated that best culture performance (in terms of growth, survival, vitality and metamorphosis rate) was achieved when offering *Artemia* nauplii enriched with the Medium emulsion. No further improvement was noticed when nauplii were offered enriched with the High n-3HUFA emulsion. From the fatty acid analysis (see Table...) the authors conclude that the requirements of *M. rosenbergii* larvae should be in between 5.2 and 37.5 mg per gram dry diet offered - probably toward the higher end. In this sense these results agree with those obtained by Buzzi et al. (1989). This further correlates with the results obtained by Sandifer and Joseph (1976) who improved the performance of a postlarval diet by increasing the n-3HUFA content to 19 mg/g. Considering n-6 fatty acids Devresse et al. (1990) could not confirm a requirement in *M. rosenbergii* larvae as was identified in postlarvae by Reigh and Stickney (1989).

Romdhane et al. (in preparation) carried out an experiment feeding newly hatched *M. rosenbergii* larvae during an increasingly shorter (delayed) period with *Artemia* enriched with SELCO (*Artemia* Systems S.A., Ghent, Belgium). n-3HUFA levels of SELCO enriched nauplii correspond to those obtained when using the Medium I.C.E.S. emulsion. From their results (see Figure...) the authors conclude that for optimal results (in terms of growth, survival, stress resistance and metamorphosis rate) prawn larvae should be fed n-3HUFA enriched *Artemia* nauplii preferably from first feeding onwards.

4. *Morone saxatilis*

One study on larval striped bass (*Morone saxatilis*) has been

conducted by Lemm and Lemarie (1991, in press). Four days post-hatch Tennessee striped bass larvae were cultured during 14 days on a diet consisting of freshly hatched or enriched (I.C.E.S. emulsions) *Artemia* nauplii. The authors report on the fatty acid profiles of the emulsions, the *Artemia*, and the fish larvae (values of n-3HUFA in *Artemia* are represented in Table....).

Fatty acid	LOW HUFA		MEDIUM HUFA		HIGH HUFA		unenr
	emul	Art.	emul	Art.	emul	Art.	
20:5n-3	0.0	2.6	9.7	8.2	25.0	12.5	3.8
22:6n-3	0.0	0.0	7.8	3.1	36.9	9.4	0.0
Σn3HUFA	0.0	2.6	19.1	11.3	66.5	22.8	3.8

As can be noticed from Table... n-3HUFA levels in *Artemia* reflect those of the enrichment emulsions; they are however inferior to those presented in Table.... Total lipid levels as reported by the authors decrease during enrichment (about 20 %) which would indicate that the enrichment procedure was not optimal. Culture results are summarized in Table...

<i>Artemia</i> treatment	length	survival	swimbladder inflation
Unenriched	9.3 +/- 1.0	5	(29)
LOW HUFA	9.8 +/- 0.9	23	26
MED. HUFA	10.3 +/- 0.8	64	33
HIGH HUFA	10.2 +/- 0.8	48	32

Growth and survival were significantly improved in the fish larvae fed the n-3HUFA enriched *Artemia*. A n-3HUFA content of 11.3 % of total fatty acids in *Artemia* appears to fulfill the requirements of first feeding striped bass larvae. Increasing the level to 22.8 % did not further improve the performance. Swimbladder inflation was not significantly affected by the dietary n-3HUFA content.

6. *Sciaenops ocellatus*

Craigh, Holt and Arnold (in press) report on experiments assessing the effects of feeding enriched rotifers, using the I.C.E.S. emulsions, on the growth and fatty acid composition of red drum (*Sciaenops ocellatus*) larvae. Fish were cultured during ten days and no *Artemia* was fed. The contents of n-3HUFA in the enriched rotifers are given in Table....

Fatty acid	CONTROL	LOW n-3HUFA	MEDIUM n-3HUFA	HIGH n-3HUFA
20:5n-3		1.5	8.6	10.7
22:6n-3		0.0	6.7	12.5
Σ n-3HUFA		1.5	15.3	23.2

Growth rates were best in the larvae fed the n-3HUFA enriched rotifers (medium and high n-3HUFA emulsions); they were however not significantly different from the control treatment fed rotifers grown on a mixture of algae, fish oil and yeast. Poorest results were recorded in the fish fed the low n-3HUFA enriched rotifers. The authors conclude that 22:6n-3 significantly affect growth in red drum larvae. XXXXXXXXXXXX--control rotifers---XXXXXX

Ten day old red drum larvae exhibit a fairly constant fatty acid profile independent of the diet offered. An extended trial covering the subsequent culturing phase including *Artemia* would allow to verify the evolution of the fatty acid profile in red drum larvae beyond the rotifer stage. More pronounced differences between treatments could eventually show up when reserve fatty acid pools would get depleted.

(*)

Conclusions

Reference List

List of Figures + Figures

List of Tables + Tables

7. SUNNER FLOUNDER (*Paralichthys dentatus*)
Zentgraf & Beumton (Larvi '51)

Biochemical composition enriched *Artemia*
Fernandez-Lucas et al (Larvi '81)