

# Assessment of trophic relationships between symbiotic tropical ophiuroids using C and N stable isotope analysis

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Analyses of the natural abundance of carbon and nitrogen stable isotopes were performed to investigate the feeding habits of two ophiuroids, *Ophiomastix venosa* and *Ophiocoma scolopendrina*, and to assess the potential benefit obtained by the symbiotic *Ophiomastix venosa* juveniles. A tracer experiment was also carried out to clarify the contribution of algae to the nitrogen uptake amongst the tested ophiuroids. Our results suggest that *Ophiocoma scolopendrina* adults occupy a higher position in the food web than *Ophiomastix venosa* and mainly feed on neuston. In contrast, *O. venosa* adults feed on the alga *Sargassum densifolium* and on organic matter associated with sediment. Free juveniles and symbiotic juveniles of *O. venosa* have intermediate  $\delta^{13}\text{C}$  values between both adult species. The high proportion of  $^{13}\text{C}$  in the symbiotic juveniles compared to the one in their conspecific adults indicates that their diet slightly differs from the latter and is closer to that of *Ophiocoma scolopendrina*. This raises the hypothesis that symbiotic juveniles steal neuston from their associated host, *O. scolopendrina*.

## INTRODUCTION

*Ophiocoma scolopendrina* and *Ophiomastix venosa* are tropical ophiuroids that co-occur on the Great Reef of Toliara located at the south-west of Madagascar. Although they both live on the boulder tract of the reef, *Ophiocoma scolopendrina* adults inhabit parts of the reef—the domes—that emerge at low tide, while *Ophiocoma venosa* adults are found in channels that are always immersed. These species are involved in a symbiosis where *O. venosa* juveniles live attached to *Ophiocoma scolopendrina* adults (Fourgon, 2006). This symbiosis seems facultative for both ophiuroids as some *Ophiomastix venosa* juveniles, of the same size than symbiotic juveniles, live freely in the channels of the Great Reef. 'Babysitting' symbiosis has been reported in brittle stars from many localities in the Indo-Pacific Ocean. Such a symbiosis, that occurred between the subtidal *Ophiomastix annulosa* and the intertidal *Ophiocoma scolopendrina*, was first described from Sesoko Island (Okinawa, Japan) by Hendler et al. (1999): *Ophiomastix annulosa* juveniles were observed in the bursa and on the disk of *Ophiocoma scolopendrina* adults. It would seem advantageous for these juveniles to take refuge in the intertidal on a large, abundant, widespread, mobile, calcified animals that occupied moist, sheltered crevices. Hendler et al. (1999) also suggested that these symbiotic juveniles would have at their disposal food from the host's arms. They considered this type of association as a brood parasitism.

*Ophiocoma scolopendrina* has been described to employ several methods for food gathering depending on currents and tide levels: it can be a deposit feeder or suspension feeder; it also feeds on the neuston (i.e. the microscopic organisms that inhabit the air-water interface) (Magnus, 1967). Nothing has previously been published about the feeding habits of *Ophiomastix venosa*.

Gut content analysis is the commonest and easiest method for determining the type of food ingested by an organism. However, when applied to invertebrates, it often fails to identify ingested particles. Yet, gut content analysis provides only 'snapshots' of the diet and only takes into account the ingested food rather than the assimilated food. On the other hand, stable isotope methods are increasingly used as tracers to study food web structure (e.g. Lepoint et al., 2000; McCutchan et al., 2003). Stable isotopes of nitrogen and carbon of an animal tissue integrate dietary components that are assimilated over a much longer period of time. The strong enrichment in  $^{15}\text{N}$  of a consumer related to its prey has been used in many studies as a tool to define the trophic levels of organisms (Post, 2002), the enrichment in  $^{13}\text{C}$  is generally used to determine the relative contributions of different potential sources to the diet of a consumer (Phillips & Koch, 2002).

The gut content analysis of *O. venosa* and *Ophiocoma scolopendrina* that we performed failed to give any information on their diet (unpublished observations). Therefore natural abundances of C and N stable isotopes in these ophiuroids were studied and are here presented in order to (i) investigate the feeding habits of *O. scolopendrina* and *Ophiomastix venosa* on the Great Reef of Toliara and (ii) determine if *O. venosa* symbiotic juveniles could take a nutritional advantage in the symbiosis.

## MATERIALS AND METHODS

### Sampling location

All samplings were made at low tide on the boulder tract of the Great Reef of Toliara (Madagascar) in March 2002. The boulder tract is characterized by the presence of rocky domes measuring between 100 to 200 m long and

~10 m width. They consist of accumulations of dead corals emerging at low tides. Rocky domes are separated from each other by permanently immersed tidal channels (water height at low tide: ~50 cm) paved with living corals, dead corals and sand. The sampling area covered a rocky dome located at 23°25'00''S and 43°39'23''E, and its adjacent northern tidal channel.

#### *Natural isotopic ratios*

Four series of ophiuroids were collected: (i) *Ophiocoma scolopendrina* adults and (ii) *Ophiomastix venosa* symbiotic juveniles (collected on the discs of their *Ophiocoma scolopendrina* hosts) were sampled on the rocky dome; (iii) free juveniles; and (iv) adults of *Ophiomastix venosa* were sampled in the tidal channel. The free and symbiotic juveniles were approximately of the same size (disc diameter between 5 and 9 mm;  $P > 0.05$ ; Mann–Whitney  $U$ -test). Free juveniles are *O. venosa* individuals that never entered an association with *Ophiocoma scolopendrina*: they would metamorphose in the channels and not on the domes like the symbiotic juveniles (Fourgon, 2006). We also collected four potential food sources for these ophiuroids. Amongst the algae present in the ecosystem, we sampled *Sargassum densifolium* that was the most abundant (fresh biomass of ~91.6 g/m<sup>2</sup>), and *Ulva pertusa* (fresh biomass of ~0.24 g/m<sup>2</sup>) that was frequently encountered near the ophiuroids. Though the investigated ophiuroids do not eat algae, they may ingest pieces of dying algae when they decompose in organic matter. The two other potential food sources sampled was the neuston, collected at the air-water interface at rising tide when it formed a dense organic film, and the organic matter associated with sediment, sampled in the tidal channel.

Five ophiuroid individuals of each category and five samples of each potential food source were collected in order to measure their natural ratios of C and N stable isotope. The ophiuroids were kept overnight in 60-l tanks filled with filtered seawater to facilitate the evacuation of their gut contents. The symbiotic juveniles were separated from their host the following day. All samples were dried for 48 h at 50°C.

#### *<sup>15</sup>N tracer experiment*

Five ophiuroid individuals of each of the four categories (namely *O. scolopendrina* adults, *Ophiomastix venosa* adults, free *O. venosa* juveniles and symbiotic *O. venosa* juveniles) were collected, brought to the laboratory and placed in four separated 60-l tanks of filtered seawater, with bubbled air. Symbiotic juveniles were not separated from their hosts. Every other day, for one month, 10 g of <sup>15</sup>N enriched *Sargassum densifolium* and 30 g of freshly collected sediment were placed for 2 h in each tank.

To obtain <sup>15</sup>N enriched food, one thallus of *S. densifolium* was kept for 6 d in an aerated 12-l tank filled with a <sup>15</sup>N labelled ammonium sulphate solution (75 μM; 99.0% <sup>15</sup>N) (Eurisotop, France). This solution was changed every two days. The labelled alga was then thoroughly rinsed with filtered seawater, ground into pieces of ~1 mm<sup>2</sup> and kept for one week at 4°C. A sample of labelled food was dried for 48 h at 50°C to verify the incorporation of <sup>15</sup>N in algal tissue.

After each feeding session, all tanks were emptied, thoroughly rinsed with seawater and filled with newly filtered

seawater. After the entire experiment had ended, the ophiuroids were kept in the tanks for one night, without feeding, to allow evacuation of the gut contents, and dried the following day.

#### *Isotope analysis*

All dried samples were ground with a mortar and pestle into a homogeneous powder. They were divided in two equal subsamples. The subsample intended for C stable isotopes analysis, was slightly acidified in HCl 1M to remove inorganic carbonates, rinsed and oven-dried for 48 h at 50°. The second non-acidified subsample was used to analyse N stable isotopes.

Isotopic and elemental measurements were performed in triplicates with an Optima mass spectrometer (Micro-mass, UK) coupled to a C–N–S elemental analyser (Carlo Erba, Italy). Natural stable isotope ratios were expressed in  $\delta$  notation according to the following formula:

$$\delta X = [(R_{\text{sample}}/R_{\text{standard}}) - 1] \times 1000,$$

where  $X$  is <sup>13</sup>C or <sup>15</sup>N and  $R$  is the corresponding ratio <sup>13</sup>C/<sup>12</sup>C or <sup>15</sup>N/<sup>14</sup>N. Carbon and nitrogen ratios are expressed relative to the Vienna Pee Dee Belemnite (VPDB) standard and to atmospheric nitrogen, respectively. Routine measurements are precise to within 0.3‰ for both <sup>13</sup>C and <sup>15</sup>N.

The abundance of <sup>15</sup>N in samples produced in the tracer experiment was expressed in atom % <sup>15</sup>N. This represents the proportion of <sup>15</sup>N atoms relative to the total N atoms (<sup>14</sup>N + <sup>15</sup>N).

#### *Data treatment*

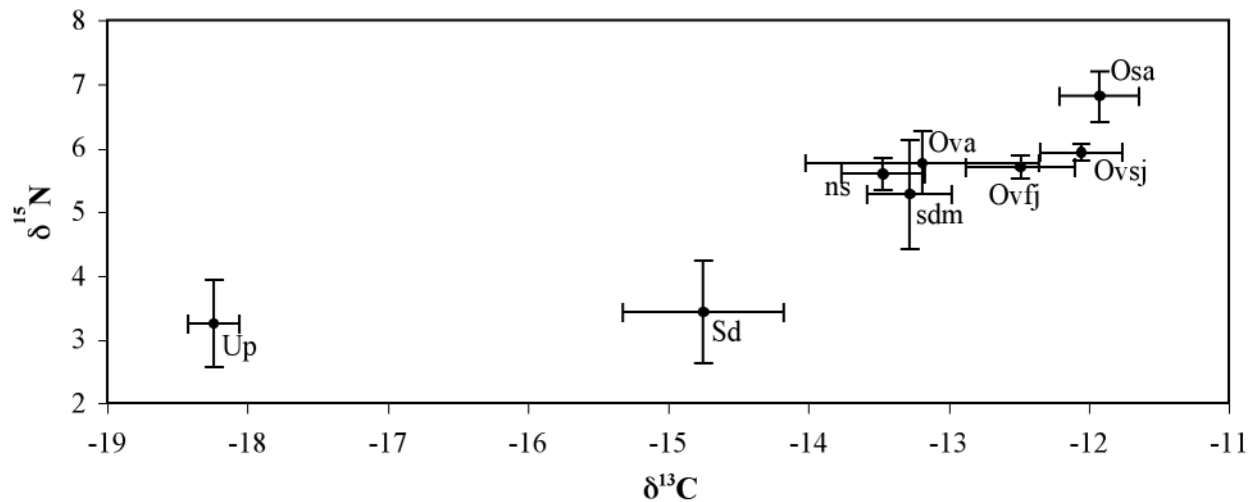
Analysis of variance followed by post-hoc multiple comparison tests were used to compare the data among different categories of ophiuroids and among different food sources. Results were considered significant for  $P < 0.05$ .

## RESULTS

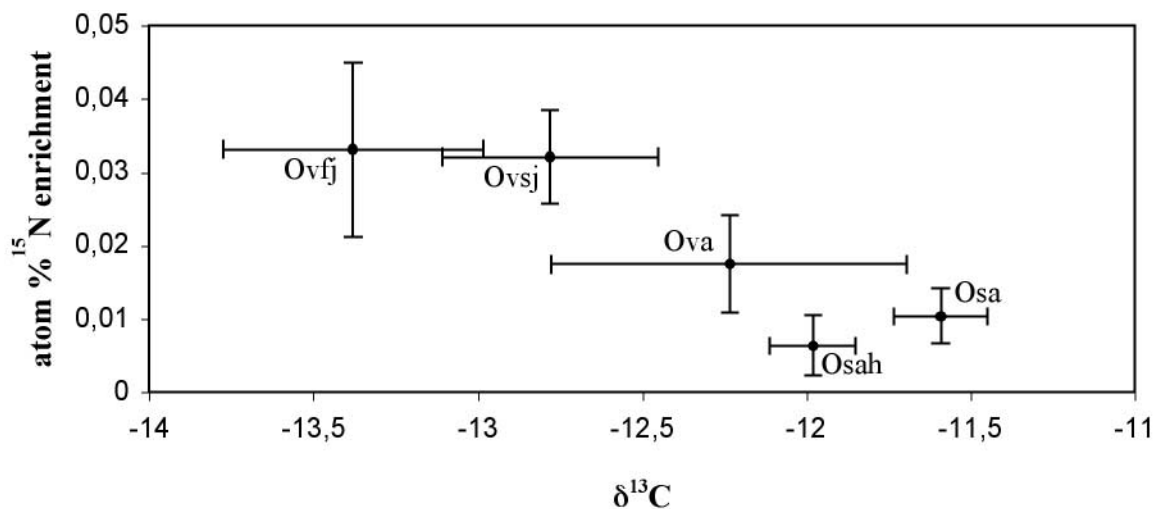
#### *Natural isotopic ratios*

Amongst the ophiuroids tested, *Ophiomastix venosa* adults possess the lowest mean  $\delta^{13}\text{C}$  value (−13.2‰) that is significantly different from the  $\delta^{13}\text{C}$  value of *Ophiocoma scolopendrina* adults (−11.9‰) ( $P < 0.01$ ) (Figure 1). *Ophiomastix venosa* free and symbiotic juveniles have intermediate  $\delta^{13}\text{C}$  signatures that are not significantly different from each other (−12.5 and −12.1, respectively) ( $P > 0.05$ ). However, while both juvenile categories do not significantly differ from *Ophiocoma scolopendrina* adults in respect with their  $\delta^{13}\text{C}$  values ( $P > 0.05$ ), only the symbiotic juveniles significantly differ from their conspecific adults ( $P < 0.02$ ).

Concerning the natural abundance of N stable isotopes, *O. scolopendrina* adults have the highest  $\delta^{15}\text{N}$  signature (6.81‰) ( $P < 0.01$ ) (Figure 1). The  $\delta^{15}\text{N}$  values of adults, free and symbiotic juveniles of *Ophiomastix venosa* do not differ significantly from each other (5.8, 5.7 and 5.9‰, respectively) ( $P > 0.05$ ).



**Figure 1.** Natural abundances of C and N stable isotope in four ophiuroids categories and four potential food sources. Osa, *Ophiocoma scolopendrina* adults; Ova, *Ophiomastix venosa* adults; Ovfj, *O. venosa* free juveniles; Ovsj, *O. venosa* symbiotic juveniles; ns, neuston; Sd, *Sargassum densifolium*; sdm, organic matter associated with sediment; Up, *Ulva pertusa*.



**Figure 2.** Enrichment in atom %  $^{15}\text{N}$  in ophiuroids after the tracer experiment compared with natural values, and their abundances of C stable isotopes. Osa, *Ophiocoma scolopendrina* adults; Ova, *Ophiomastix venosa* adults; Ovfj, *O. venosa* free juveniles; Ovsj, *O. venosa* symbiotic juveniles.

Amongst the food sources, the organic matter associated with sediment and the neuston have the highest natural  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values ( $-13.3$  and  $-13.5\%$  for  $\delta^{13}\text{C}$ , and  $5.3$  and  $5.6$  for  $\delta^{15}\text{N}$ ; respectively) (significantly different from algae,  $P < 0.02$ ), and no significant difference was observed between them ( $P > 0.05$ ). The algae *Ulva pertusa* and *Sargassum densifolium* do not differ from each other in their  $\delta^{15}\text{N}$  mean values ( $3.2$  and  $3.4\%$ , respectively), but *U. pertusa* has by far the lighter C isotope value than *S. densifolium* ( $\delta^{13}\text{C}$  of  $-18.24$  and  $-14.75\%$ , respectively).

#### $^{15}\text{N}$ tracer experiment

Isotopic analysis of labelled *S. densifolium* revealed that they are successfully enriched in  $^{15}\text{N}$ : their mean  $^{15}\text{N}$  abundance is  $2.73$  atom %  $^{15}\text{N}$  that is more than seven times higher than their natural  $^{15}\text{N}$  abundance ( $0.37$  atom %  $^{15}\text{N}$ ). All ophiuroids are significantly enriched in  $^{15}\text{N}$  compared with their natural  $^{15}\text{N}$  abundances ( $P < 0.05$ ).

*Ophiomastix venosa* free and symbiotic juveniles display the highest enrichments ( $0.033$  and  $0.032$  atom %  $^{15}\text{N}$  in excess relatively to natural  $^{15}\text{N}$  abundance, respectively) (Figure 2). These enrichments are significantly higher than those of all adults ( $P < 0.03$ ) (Figure 2). *Ophiocoma scolopendrina* adults have the lowest enrichment ( $0.010$  atom %  $^{15}\text{N}$ ), although they do not differ significantly from those of *Ophiomastix venosa* adults ( $0.017$  atom %  $^{15}\text{N}$ ) ( $P > 0.05$ ) (Figure 2).

## DISCUSSION

A very large difference is observed in the natural abundances of C and N stable isotopes between adults of *Ophiomastix venosa* and of *Ophiocoma scolopendrina*, the latter having the highest  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values. *Ophiomastix venosa* free and symbiotic juveniles have  $\delta^{15}\text{N}$  values similar to their conspecific adults, but have intermediate values of  $\delta^{13}\text{C}$ .

Since  $\delta^{15}\text{N}$  is related to the trophic level of organisms (Post, 2002),  $\delta^{15}\text{N}$  values of *Ophiocoma scolopendrina* would indicate that this species eats  $^{15}\text{N}$  enriched food and occupies a higher position in the food web than the other sampled ophiuroids. From field observations, the neuston, which forms a dense film below the air-water interface at rising tide, seems to constitute the major component of the diet of *O. scolopendrina*. Indeed, it is abundantly caught by *O. scolopendrina*, which curls the tip of its arms (generally three at once), directing the oral side towards the air-water interface (Magnus, 1967; Chartock, 1983).

Isotope analyses support the importance of the neuston in *O. scolopendrina*'s diet. The analyses, however, indicate trophic shifts ( $\Delta = \delta_{\text{consumer}} - \delta_{\text{diet}}$ ) of 1.21‰ in  $\delta^{15}\text{N}$  and of 1.55‰ in  $\delta^{13}\text{C}$ , that strongly differ from the mean trophic shifts described in aquatic systems of 2.9‰ and 0.47‰, respectively (Vander Zanden & Rasmussen, 2001). Similarly low values for  $\Delta^{15}\text{N}$  are known to occur amongst detritus feeders (Vanderklift & Ponsard, 2003), and particularly in an unnamed ophiuroid (Fry, 1988). However, a large  $\Delta^{13}\text{C}$  is known to occur in benthic detritus feeders because of the presence of bacteria in the substrates that raises the  $^{13}\text{C}/^{12}\text{C}$  ratio of organic matter (McConnaughey & McRoy, 1979). It should, however, be pointed out that the small  $\Delta^{15}\text{N}$  and the large  $\Delta^{13}\text{C}$  observed for *O. scolopendrina* might also result from a selective feeding on the neuston, as suggested for other organisms that feed on particulate organic matter (Hsieh et al., 2000; Vanderklift & Ponsard, 2003).

*Ophiomastix venosa* adults probably feed upon various nutrient sources, as indicated by the large variance associated with the  $\delta^{13}\text{C}$  mean (see Bearhop et al., 2004; Matthews & Mazumder, 2004). The information on their  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values suggests that *O. venosa* adults have a mixed diet composed of organic matter associated with sediment and algae. In the light of our results, *Sargassum densifolium* would constitute one of the algal components of their diet, *Ulva pertusa*  $\delta^{13}\text{C}$  value being too far from the value recovered for *O. venosa* adults.

$^{15}\text{N}$  enrichment analyses do not reveal significant difference between both adult species, which suggests that *Ophiocoma scolopendrina* is also able to feed on algae when neuston is not available. The contribution of algae to the diet of *O. scolopendrina* is presumably less important than in *Ophiomastix venosa*, as the former had the smallest enrichment in  $^{15}\text{N}$ . Hence, in our tracer experiment, the low  $^{15}\text{N}$  abundance in adults compared to juveniles might not only result from differences in their diet, but also from differences in the tissue turnover rate that would logically be higher in juveniles because of their faster growth, leading to higher  $^{15}\text{N}$  assimilation.

The  $\delta^{13}\text{C}$  values obtained on *O. venosa* juveniles (on both free juveniles and symbiotic juveniles) sampled in the field are significantly lower than those obtained on individuals that fed on *S. densifolium* over one month ( $P < 0.05$  for both categories). This result probably reflects that, in the field, free and symbiotic juveniles, if they feed on *S. densifolium*, have surely other carbon sources that would have a higher  $\delta^{13}\text{C}$  than this alga. On the other hand, the  $\delta^{13}\text{C}$  values of adults (of both species) sampled in the field do not differ significantly from those of individuals that fed on *S. densifolium* over one month ( $P > 0.05$  for both categories).

The high  $\delta^{13}\text{C}$  value in symbiotic juveniles, compared to that in their conspecific adults, indicates that their diet is slightly different. It would be closer to the diet of their *Ophiocoma scolopendrina* hosts. Symbiotic juveniles might obtain neuston by stealing it from their hosts. Indeed, juveniles cling to the host in such a way that the oral side of their arms cross the oral side of *O. scolopendrina*'s arms. This would allow them to pick up food particles that move toward *O. scolopendrina*'s mouth. Such similar stealing behaviour has also been proposed for other ophiuroids, for instance juveniles of *Ophiothrix fragilis* are thought to intercept the food from conspecific adults, *Ophiomaza cacaoica* from crinoids, and *Ophiomastix annulosa* from *Ophiocoma scolopendrina* (Warner, 1969; Clark, 1976; Hendler et al., 1999; respectively). Our results support the proposal (Hendler et al., 1999) that juvenile *Ophiomastix* that are associated with *Ophiocoma scolopendrina* may steal food from the host. That appears to be the case for juvenile *Ophiomastix venosa*. Furthermore, the results indicate that *O. venosa* juveniles ingest the neuston collected by *Ophiocoma scolopendrina*, and perhaps do so selectively in preference to other potential food items.

This work was supported by the Belgian National Fund for Scientific Research (FRFC contract 2.4.583.05). We acknowledge the assistance of J.M. Ouin, F. Mamitiana, J. Ralainirina, P. Manohitsara, N. Fohy, and Prosper. We thank G. Hendler and D. Vaitilingon for insightful comments that improved the quality of this manuscript. This paper is a contribution from the 'Centre Interuniversitaire de Biologie Marine' (CIBIM) and from the MARE centre (University of Liège).

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*Submitted 6 October 2005. Accepted 24 October 2006.*