



Contents lists available at ScienceDirect

Journal of King Saud University – Science

journal homepage: www.sciencedirect.com

Original article

The adverse health effects of increasing microplastic pollution on aquatic mammals



Ghulam Nabi^{a,*}, Shahid Ahmad^{a,1}, Sana Ullah^b, Sahib Zada^d, Maliha Sarfraz^e, Xinle Guo^{c,*}, Muhammad Ismail^f, Kunyuan Wang^{g,*}

^a Ministry of Education Key Laboratory of Molecular and Cellular Biology, Key Laboratory of Animal Physiology, Biochemistry, and Molecular Biology, College of Life Sciences, Hebei Normal University, Shijiazhuang 050024, China

^b Department of Zoology, Division of Science and Technology, University of Education, Lahore 54000, Pakistan

^c Academy of Plateau Science and Sustainability, College of Life Sciences, Qinghai Normal University, Xining 810008, China

^d Department of Environmental Engineering, Guangdong Technion Israel Institute of Technology, Shantou 515063, China

^e Department of Zoology, Wildlife, and Fisheries, University of Agriculture, Faisalabad 38000, Pakistan

^f Department of Zoology, Islamia College, Peshawar 25120, Pakistan

^g Key Laboratory of Adaptation and Evolution of Plateau Biota, Qinghai Provincial Key Laboratory of Animal Ecological Genomics, Northwest Institute of Plateau Biology, Chinese Academy of Sciences, 23rd Xinning Road, Xining 810008, China

ARTICLE INFO

Article history:

Received 28 December 2021

Revised 6 February 2022

Accepted 27 March 2022

Available online 31 March 2022

Keywords:

Aquatic mammals

Aquatic ecosystem

Heavy metals

Microplastic

Pesticides

ABSTRACT

Microplastics (MPs), an emerging ubiquitous pollutant in the aquatic ecosystem, pose serious health concerns to the survival of aquatic fauna, especially top predators (e.g., aquatic mammals). It is challenging to investigate the toxicological profile of MPs in aquatic mammals due to their diverse toxicological behaviour, physico-chemical properties, and other technical and ethical issues. This study reviewed the current burden of MPs in the aquatic ecosystem, the occurrence of MPs in the various tissues of aquatic mammals, its composition (heavy metals, pesticides, pathogens), and possible health effects on individual and population levels in aquatic mammals. Aquatic mammals are constantly exposed to MPs directly and indirectly via the food-web. The MPs and a wide range of toxic heavy metals, pesticides, and pathogens added during manufacturing or adsorbed from the surrounding environments are bioaccumulated in aquatic mammals for years. Due to their long life-span and heavy body masses, these pollutants can cause several serious health issues in aquatic mammals that can drastically reduce the population size and ultimately can cause extinction, especially in vulnerable populations. Still today the toxicological profile of MPs and its presence in other deep tissues largely remains unknown in aquatic mammals. We therefore suggest a global assessment of the risks associated with the consumption of MPs by aquatic mammals and the presence of MPs in their habitats.

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1. Introduction

Aquatic mammals are the charismatic megafauna and prime sentinel species due to their long-life spans, apex position in the

trophic web, long-term coastal residents, and remarkable fat reserves that serve as depots for anthropogenic toxins (Moore, 2008; Bossart, 2011). These mammals have an essential role in influencing the structure and function of the marine environment (Moore, 2008). Currently, many aquatic mammals are of conservation concern due to escalating anthropogenic activities, including plastic and other chemical pollution (Avila et al., 2018; De Sá et al., 2018; Nabi et al., 2018a; Nabi et al., 2018b; Meaza et al., 2021; Nabi et al., 2021). However, there is limited knowledge of the many anthropogenic contaminants in aquatic ecosystems regarding the relationship between the widespread microplastics (MPs) and the well-being of aquatic mammals. The key objective of this study is to provide an overview of the MPs bioaccumulation and their possible health consequences in aquatic mammals.

* Corresponding authors.

E-mail addresses: ghulamnabiqau@gmail.com (G. Nabi), Yuexinfeiyang1@163.com (X. Guo), wanghekunyuan@189.cn (K. Wanghe).

¹ These authors contributed equally as first author.

Peer review under responsibility of King Saud University.



Production and hosting by Elsevier

<https://doi.org/10.1016/j.jksus.2022.102006>

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2. Rising plastic pollution in aquatic ecosystem

Plastic waste is a modern societal and ecological issue due to its convenience and ubiquitous use in daily life and associated long-term harmful effects on the ecosystems and organisms after careless disposal (Galloway et al., 2017). Plastic constitutes approximately 60–80% of the debris in a marine environment and is currently considered as a significant threat to marine biodiversity above all other environmental hazards, including different types of pollution, climate change, resource overexploitation, and invasive species (Derraik, 2002; Halpern et al., 2008; Nabi et al., 2018a; Nabi et al., 2018b). They are transported far and wide by winds, rivers, and ocean currents. Every year, approximately 5–13 million tonnes of plastic debris end up in the ocean ranging from the coastline, seafloor, mid-ocean gyres, and ultimately, in remote polar regions (Jambeck et al., 2015; Van Sebille et al., 2015; Peeken et al., 2018; Mendes et al., 2021; Osorio et al., 2021). Currently, an estimated 24.4 trillion plastic pieces ($\sim 57.8 \times 10^4$ tonnes) are found in the world's upper ocean (Isobe et al., 2021), and by the year 2025, the weight of plastic debris in the marine environment could increase to 250 million metric tonnes (Jambeck et al., 2015). Furthermore, during the ongoing COVID-19 pandemic, a large quantity of plastic waste has been generated globally. According to Peng et al. (2021), over eight million tons of plastic wastes have been generated globally amidst the COVID-19 pandemic, with over 25,000 tons entering the ocean. In total, 11 million tons of mismanaged plastic waste will be generated by the COVID-19 pandemic; among them, 34,000 tons of plastic waste will enter the ocean (Peng et al., 2021). Sun et al. (2021) reported that approximately 1370 trillion MPs would enter the global coastal marine environments by 2020 from surgical masks discarded throughout the world. Plastic waste is resistant to oxidative damage, heat, and degradation due to its physical and chemical nature. It persists for a long time in the marine ecosystem and is therefore characterized as an environmental contaminant (Andrady, 2011; Vroom et al., 2017).

3. Microplastics in aquatic ecosystem

MPs (<5 mm) are persistent synthetic particles ubiquitously found in a wide range of aquatic ecosystems (Bravo Rebollo et al., 2013; Vroom et al., 2017). Depending on their shape, density, and composition, MP can sink, be neutrally buoyant, and float (Hernandez-Gonzalez et al., 2018). The density of MPs in the ocean surface water ranged from 5 to 70 particles per m^{-2} (Ter Halle et al., 2017), whereas in coastal water, it is six items m^{-3} (Cole et al., 2011). In rivers surface water, the density of MPs is very high, ranging from 1,580 to 57,665 particles/ m^3 (Osorio et al., 2021). The marine environment is a major sink for MPs (plastic soup) where they are found in two different ways either primary MPs such as microspheres or MPs pellets and secondary MPs resulting from the degradation of larger fragments (Fig. 1; Andrady, 2017; Ross et al., 2021). MPs originate from various sources, including macro-plastics degradation (by UV light, physical abrasion, wave action), shipping spills, and wastewater discharge from industries and other sources containing microbeads and microfibers (Nelms et al., 2018; Du et al., 2021). Currently, research interest is developing to investigate the consequences of MPs on the aquatic eco-health and the organism within it. Due to their small size, MPs are ingested by a variety of taxa, including aquatic mammals either accidentally (indiscriminate feeding strategies users, e.g., filter feeders), misidentification of MPs for prey, via inhalation, and indirectly due to the trophic transfer (raptorial feeding strategy users, e.g., dolphins) by consuming contaminated prey (Hocking et al., 2017; Nelms et al., 2018; Meaza et al., 2021). Despite direct inges-

tion of MPs, macro-plastics ingested are fragmented into smaller pieces within the gastro-intestinal tract (GIT) of aquatic mammals (Nelms et al., 2018; Huang et al., 2021). According to Lusher et al. (2016), 11% of mesopelagic fish in the Irish waters contained MPs and approximately, 463 million MPs could be ingested by a single striped dolphin annually.

4. MPs in aquatic mammals

To date, approximately 3876 species have been reported to be affected by marine debris (https://litterbase.awi.de/interaction_detail; date accessed, April 4, 2022). Among them, being an apex predator, plastic debris ingestion has been reported in 123 aquatic mammalian species, including 36% of seals (12 of 33), 59% of whales (47 of 80), and overall, 63% of all cetacean species (Bergmann et al., 2015; Fossi et al., 2018). However, due to logistic, technical, and ethical issues, few studies have recently reported the presence of MPs in the GIT (Bravo Rebollo et al., 2013; Besseling et al., 2015; Lusher et al., 2015; Lusher et al., 2018; Hernandez-Gonzalez et al., 2018; Van Franeker et al., 2018; Xiong et al., 2018; Nelms et al., 2019; Zhu et al., 2019; Moore et al., 2020; Haave et al., 2021) and fecal samples ((Eriksson and Burton, 2003; Perez-Venegas et al., 2018; Nelms et al., 2018; Donohue et al., 2019; Hudak and Sette, 2019; Perez-Venegas et al., 2020) in more than 21 species of aquatic mammals across the globe (Table 1). The total number of MPs/individuals in these aquatic mammals ranges up to 411 in the stomach (Hernandez-Gonzalez et al., 2018) and 584 in the scats (Donohue et al., 2019). Furthermore, the presence of MPs in the grey seals living in a sanctuary (Nelms et al., 2018), where anthropogenic pollution is low, suggests the omnipresence of MPs even in environmentally controlled areas. The size range of MPs reported in aquatic mammal's scats and GIT content are highly heterogeneous and range from 0.1 to 5 mm (Meaza et al., 2021). Fibers shape MPs are the most abundant MPs in the GIT (Hernandez-Gonzalez et al., 2018; Xiong et al., 2018; Zhu et al., 2019; Nelms et al., 2019), while fragment shape MPs are ubiquitous in the scats of aquatic mammals (Eriksson and Burton, 2003; Hudak and Sette, 2019; Donohue et al., 2019; Perez-Venegas et al., 2020). Different colours of MPs, including green, white, black, and blue, are reported in the GIT and scats (Xiong et al., 2018; Lusher et al., 2018; Hernandez-Gonzalez et al., 2018; Perez-Venegas et al., 2020), while scats often additionally contained yellow, brown, purple, and red colour MPs (Meaza et al., 2021). The most common polymer types reported are polyether-sulfone, nylon, cotton, polyester, polyethylene, polypropylene, and ethylene-propylene (Nelms et al., 2019; Meaza et al., 2021). However, others include cellophane, alkyd resin, low-density polyethylene, polyamide resin, acrylic, polyvinyl chloride, polystyrene, cellulose, polycarbonate, and polyolefin, have also been reported in aquatic mammals (Meaza et al., 2021). The current knowledge linked to the toxicities of MPs in aquatic mammals is scanty; however, the lack of studies should not be interpreted as evidence of no risk. MPs toxicities could be detrimental, especially to endangered and critically endangered species populations dwelling in highly polluted areas. Furthermore, considering the escalating rise of plastic debris in water bodies, the morbidities and mortalities in aquatic mammals due to MPs will become more evident if comprehensive studies are carried out using advanced sensitive techniques.

5. Why MPs are dangerous for aquatic mammals

Several hundreds of chemicals are either added or adsorbs by the MPs, including phthalates, organochlorine contaminants (HCB, DDTs, PCBs), persistent organic pollutants, organophospho-

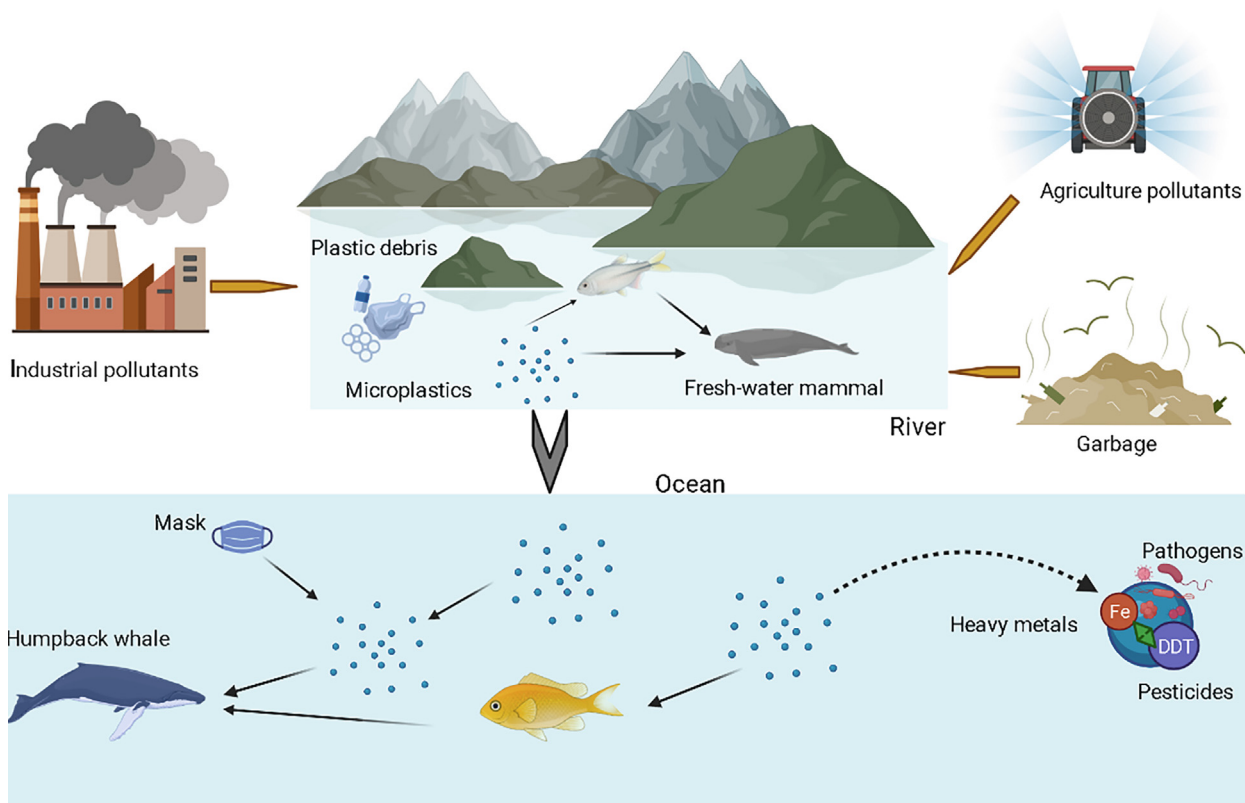


Fig. 1. The primary and secondary microplastics loaded with pathogens and a wide range of chemical pollutants from various sources entered the aquatic ecosystem. The aquatic mammals ingest the microplastics directly from the environment and via the food-web. (Biorender.com)

rus esters, dioxins, antibiotics, tributyltin, bisphenol A, aluminium oxide, chromium, lead, cadmium, antimony, and other chemical ingredients (Ylitalo et al., 2005; Fossi et al., 2014; Fossi et al., 2016; Kedzierski et al., 2018; Campanale et al., 2020; Barrick et al., 2021; Meaza et al., 2021). Due to the contaminant's hydrophobicity, these contaminants have a greater affinity for MPs than natural sediments and seawater. Therefore, MPs act as a vector of pollutants and toxins, which, along with the biomagnification and bioaccumulation process, makes MPs a potential threat to the well-being of aquatic mammals (De Sá et al., 2018). MPs ingestion can cause a reduction in energy reserves, feeding capacity and reproductive outputs, detrimental alterations in the GIT physiology, depressed immune system, vulnerability to diseases, oxidative stress, cytotoxicity, and differential expression of genes, mainly studied in lower trophic animals (Hall et al., 2006; Kedzierski et al., 2018; Nabi et al., 2019; Amin et al., 2020; Meaza et al., 2021; Ugwu et al., 2021). MPs ingestion can alter the kidney histomorphology, induce inflammation, and disrupt several biomarkers (Meng et al., 2022). Furthermore, MPs exposure can cause oxidative and epithelial damage in the gut, immune cell toxicity, microbial disorder, and reduction of the mucous layer.

6. Pollutants in MPs and its biomagnification and effects on aquatic mammals

The MP's larger surface area-to-volume ratio and hydrophobic surface make them a suitable sorbent for many toxic chemicals, including heavy metals and organic chemicals (Verla et al., 2019; Wang et al., 2020). The loaded MPs, therefore, act as a conveyor ("Carrier effects") of toxic pollutants (Verla et al., 2019; Wang et al., 2020) and increase the bioaccumulation of sorbed pollutants

in aquatic mammals directly and indirectly via the food web. Unlike seawater, gut surfactants accelerate the release of hydrophobic organic pollutants ("releaser effects") from MPs and bioaccumulate them in the different tissues (Lee et al., 2019). In aquatic mammals, MPs are only reported in the GIT and scats (Table 1); however, a large number of metals and organic pollutants bioaccumulated in the different tissues (Lungs, liver, kidneys, skin, muscle, blubber, bone, testes, ovaries, blood) of many aquatic mammalian species have been reported across the world (Table 2). On the one side, pollutants bioaccumulation in different tissues depends on the nature of MPs. However, on the other hand, MPs/nano-plastics in aquatic mammals, like other organisms, may cross the cell membrane and blood-brain barrier, enter into the blood circulation and bioaccumulate in the other vital internal organs (brain, liver, kidney, placenta) along with other pollutants (Prieti et al., 2014; Shelver and Banerjee, 2021). Further studies are needed to confirm the presence of MPs in these deep tissues of aquatic mammals. In addition, further studies in aquatic mammals need to investigate MPs in the internal tissues and their correlation with other pollutants. Furthermore, the transplacental transfer of heavy metals, pesticides, and other pollutants to the fetuses of aquatic mammals (Teigen et al., 1999; Alonso et al., 2015; Zanuttini et al., 2019) and MPs to the fetuses of other species in mammals (Fournier et al., 2020; Ragusa et al., 2021) also provides a hint that MPs could bioaccumulate in aquatic mammals during fetal development. Accumulation of heavy metals and organic pollutants causes several ill effects, including congenital disabilities, endocrine disruption, immune system dysfunction, vascular damage, nervous system disorders, kidney and gastrointestinal dysfunction, and cancer (Nabi et al., 2017a,b; Nabi et al., 2020a; Balali-Mood et al., 2021). Pesticides can cause an increase in oxidative stress, producing superoxide, and alter different liver func-

Table 1
Microplastics reported in different species of aquatic mammals globally.

Species	Location	Sample	References
White-beaked dolphin Striped dolphin Risso's dolphin Pygmy sperm whale Harbour seal Harbour porpoise Grey seal Short-beaked common dolphin Bottlenose dolphin Atlantic white-sided dolphin	British coast	GIT	Nelms et al., 2019
True's beaked whale Cuvier's beaked whale Common dolphin Striped dolphin Harbour porpoise Killer whale Bottlenose dolphin	Ireland	GIT	Lusher et al., 2015 Lusher et al., 2018
Humpback whale Harbour porpoise Harbour seal	Netherland	Intestine Stomach GIT	Besseling et al., 2015 Van Franeker et al., 2018 Bravo Rebolledo et al., 2013
East Asian finless porpoises Indo-Pacific humpback dolphins	China	Intestine	Xiong et al., 2018 Zhu et al., 2019
Common dolphin	Iberian Peninsula	Stomach	Hernandez-Gonzalez et al., 2018
Beluga whale	Canada	GIT	Moore et al., 2020
Otters	Norway	Stomach	Haave et al., 2021
Northern fur seals Harbour seal, gray seal	USA	Scats	Donohue et al., 2019 Hudak and Sette, 2019
Fur seals	Macquarie Island	Scats	Eriksson and Burton, 2003
	Guafo Island		Perez-Venegas et al., 2018
Otariids	Peru and Chile	Scats	Perez-Venegas et al., 2020
Grey seals	UK	Scats	Nelms et al., 2018

tions, including detoxification, metabolism, and immunity, resulting in an increase in ROS in the liver (Chen et al., 2021). In response to liver toxicity, hepatocyte inflammation, degeneration, mitochondrial dysfunction, and apoptosis occur (Karami-Mohajeri et al., 2017).

The combination of heavy metals to heavy metals, heavy metals to pesticides, and pesticides with pesticides act synergistically and exhibits higher toxicity than a single molecule alone. When combined, the toxic hazards of a single component can be modified, resulting in unexpected adverse health consequences. The combined interactions of these xenobiotics (heavy metals, pesticides, etc.) in MPs could produce synergistically significant adverse effects on neurotoxicity, cytotoxicity, immunotoxicity, infertility, and several metabolic functions (Singh et al., 2017). Given aquatic mammals' long life-span and their exposure to a mixture of xenobiotics, the synergistic interactions among xenobiotics in aquatic environments could have several negative effects on health, and further research is needed to address these issues. Unfortunately, aquatic mammals have convergently lost the traits, including *Paraoxonase 1*, which encodes a bloodstream enzyme that hydrolyzes organophosphate compounds (Meyer et al., 2018). The loss of this enzyme and the escalating rise of organophosphate compounds in MPs pose severe risks to vulnerable populations in the modern environment.

7. EDCs in MPs and aquatic mammals

Over 1000 chemicals reported as endocrine disruptor chemicals (EDCs) are found globally in different ecosystems (Kassotis and Trasande, 2021). EDCs comprise the MPs themselves (Amereh et al., 2020) and a diverse group of fungicides, pesticides, metals, phytoestrogens, pharmaceutical agents, nonylphenols, plasticizers, industrial chemicals etc., (Yilmaz et al., 2020; Ullah et al., 2021a;

Ullah et al., 2021b) are transported by MPs to remote areas including the Arctic (Barrick et al., 2021; Meaza et al., 2021; Routti et al., 2021) and bioaccumulate in aquatic mammals (Routti et al., 2021; Table 2). Most EDCs are lipophilic; they bioaccumulate in adipose tissues, are resistant to metabolism, slowly excreted, and, therefore, have a long half-life in the body (Rodprasert et al., 2021). These EDCs can bind to the body's endocrine receptors, leading to activation, blocking, or alteration of natural hormone synthesis and degradation through various mechanisms, resulting in abnormal, lack, or false hormonal signals that can increase or inhibit normal endocrine function (Yilmaz et al., 2020; Rodprasert et al., 2021). Aquatic mammals are sensitive to the toxicological effects of EDCs (Fossi and Marsili, 2003). Several studies have reported the effects of diverse group of EDCs and their effects on aquatic mammals (Table 3). However, the impacts of EDCs are challenging to assess because their adverse effects develop latently and manifest at a later age (Kassotis and Trasande, 2021). EDCs affect the reproductive system and survival through endocrine disruption and immune suppression, which have population-level effects in a region (Sonne et al., 2010). In addition, the EDCs can impair development, reproduction, thyroid functions and metabolism, cause cancer, neurological issues, and gestational disorders including, fetal growth restriction, miscarriage, and preterm birth (Desforges et al., 2018; Murphy et al., 2018; He et al., 2021; Schjenken et al., 2021; He et al., 2021).

EDCs, when used in combination, result in outcomes that cannot be predicted from their individual behavior (Delfosse et al., 2021). For example, a mixture of phthalates such as DEHP + DBP (Tian and Li, 2010), polyfluoroalkyl substances (Preston et al., 2020), and fenvalerate and thiophanate-methyl (Wang et al., 2021), significantly disrupt the thyroid glands normal function relative to the individual chemical exposure. Several EDCs chemicals produce estrogen-like effects (Guo et al., 2021) and 66 of 200 pesticides tested reported antiandrogenic activity (Kojima et al.,

Table 2
Pollutants linked with microplastics reported in aquatic mammals.

Species	Tissues	Pollutants	References
Bottlenose dolphin			Becker, (2000)
Common dolphin			
Striped dolphin			Nielsen et al., (2000)
Risso's dolphin			
Atlantic spotted dolphin	Lung, liver, kidney,	Cadmium, Cobalt, Iron, Manganese, Magnesium,	Frodello and Marchand, (2001)
white-beaked dolphin	skin, muscle, blubber	Sliver, Nickle, Selenium, Zinc, Copper, Lead,	
Spinner dolphin	bone, testes, ovaries,	Thallium, Mercury, Arsenic, Chromium, Strontium,	Ikemoto et al., (2004)
Rough-toothed dolphin	blood	Tin, Vanadium, Aluminium, Titanium, Uranium,	Haynes et al., (2005)Kannan et al., (2005)
Indo-Pacific humpback dolphin		Antimony, Beryllium, Gold, Barium, Lithium,	
Irrawaddy dolphins		Molybdenum, Platinum	Das et al., 2006
Hector's dolphin			
Gervais' beaked whale			Gerpe et al., 2007
Pygmy sperm whale			
Pygmy killer whale			McHugh et al., (2007)
Dwarf sperm whale			
melon-headed whale		Aldrin, Dieldrin, DDT, DDD, DDE, PCB, Chlordane,	Fair et al., (2010)
Cuvier's beaked whale		Toxaphene, Heptachlor epoxides, Bisphenol A,	
Humpback whale		Atrazine, Diethyl phthalate, Nonylphenol	Niño-Torres et al., (2010)
Killer whale		ethoxylate, Triclosan, Perfluoroalkyl phosphinic	
False killer whale		acids, Organochlorine, Dicofol,	Stockin et al., (2010)
Beluga whale		Polychlorinated biphenyls, Polybrominated	Aubail et al., (2013)
Bowhead whale		diphenyl ethers, Endosulfan, Dioxin related	
Fin whale, minke whale		compounds, Pyrethroid pesticides, Ultra-violet	Siegal-Willott et al., (2013)
Pilot whale		filters	
Dall's porpoise			Alonso et al., (2015)
Harbour porpoise			
Yangtze finless porpoise			De Silva et al., 2016
Northern fur seals			
Southern Sea lion			Hansen et al., (2016)
Ringed seal			
Harbor seal			Takeuchi et al., (2016)
Walrus			Mackintosh et al., (2016)
Polar bear			
Manatees			Gui et al., (2017)
Dugong			Aznar-Alemany et al., (2017)
			Wise et al., 2019
			Xiong et al., (2019)
			Zanuttini et al., (2019)
			Page-Karjian et al., (2020)

2004). The combined mixture of cypermethrin and malathion (Guo et al., 2021) and a mixture of phthalate ester (Gao et al., 2017) synergistically increase the estrogenic effects and disrupt the endocrine signaling system.

When EDCs interact with the thyroid hormone axis and estrogen receptors, they form reactive oxygen species (ROS) and increase oxidative stress, resulting in impaired reproductive growth (Park et al., 2020). The Hypothalamic-pituitary-adrenal (HPA) axis, which controls the stress system, immune system, emotions, and reproduction, may be involved in these interactions (Salgado-Freiría et al., 2018); however, there are a limited number of EDCs mixture studies related to HPA axis effects. Based on limited studies, exposure to various EDCs mixtures considerably increases ROS production, stress response genes, immune-related genes, and oxidative stress enzymes in vertebrates (Hamid et al., 2021). Furthermore, there is limited literature about the deleterious epigenetic multi- and transgenerational effects of EDCs mixture. In a few studies, rats developmentally exposed to a mixture of EDCs showed multi- and transgenerational effects, including a significant increase in kidney diseases, reproductive and metabolic disorders, and behavioural alterations (López-Rodríguez et al., 2021).

Considering the significant effects of EDC on wildlife health and existence, it is essential to identify MPs dense regions and pay serious attention to the aquatic wildlife within, especially threatened populations. It is also important to know how many EDC are in the MPs? Where do they come from, and what is their mechanism of action? When used individually or in combination, what are their deleterious effects during development, adulthood, and across generations?

8. MP-linked chemical pollutants and cancer in aquatic mammals

Even at low concentrations, chronic exposure to heavy metals promotes chronic inflammation and subsequently develops carcinogenesis (Balali-Mood et al., 2021). Among the principal metals, including lead, aluminum, silver, arsenic, beryllium, copper, chromium, mercury, cadmium, nickel, radium, and cobalt reported in aquatic mammals (Table 2), can cause lung, brain, skin, liver, gastrointestinal tract, larynx, urinary bladder, kidney, prostate cancer, and leukemia (Cruz et al., 2021; Gallicchio and Harper, 2021). Similarly, several heavy metals, pesticides, and biocides bioaccumulated in aquatic mammals can cause cancer in the endocrine

Table 3
Endocrine disruptors and their effects on aquatic mammals.

Species	Endocrine disruptors	Targeted endocrine glands/effects	References
Blue whales	Phthalates		He et al., 2021
Fin whales	POPs		Kimberley et al., 2021
Grey seals	PBDE		Routti et al., 2019
Male walruses	PCBs	Thyroid glands	Villanger et al., 2011
Beluga whales	POC		Hall et al., 2003
Northern elephant seals			Beckmen et al., 1997
Killer whale			Desforges et al., 2018
Harbor porpoises			Murphy et al., 2018
Common dolphin	PCBs	Reproductive failure	Murphy et al., 2015
Habor seal	Organochlorines		Reijnders,1999
Gray and Ringed seal			
Beluga whale			
Grey-seals, Ringed seals	POP	Sex hormones	Troisi et al., 2020
	PCB		
Bottlenose dolphin	HOC	Testosterone	Trego et al., 2019
Short-beaked common dolphins		Testosterone	Trego et al., 2018
Harbor seals	PCBs	Testosterone, progesterone	Troisi and Mason, 2000
Bottlenose dolphins	DDT	Androgen, cortisol	Galligan et al., 2019
			Ropstad et al., 2006
		Thyroid glands	Braathen et al., 2004
Polar bears	PCBs	Progesterone	Oskam et al., 2004
	Organochlorines	Testosterone	Haave et al., 2003
		Cortisol	Oskam et al., 2003
			Skaare et al., 2001
Beluga whale	PAH	Gonadal cancers	
Pilot whale			Murphy et al., 2018
Blue whale			Martineau et al., 2002
Fin whale			
Harbor porpoise			
Bottlenose dolphin			
Other cetaceans			

PCBs, Polychlorinated Biphenyls; POPs, Persistent Organic Pollutants; POC, Persistent Organohalogen Compound; PBDE, Polybrominated Diphenyl Ethers; HOC, Halogenated Organic Compounds; PAH, Polycyclic Aromatic Hydrocarbons.

glands, including the thyroid, testes, ovary, prostate, and mammary glands (Lauretta et al., 2019). In addition, heavy metals disrupt many cellular functions, such as growth, proliferation, differentiation, damage DNA repair systems, apoptosis, and genomic stability (Balali-Mood et al., 2021). A review of the mechanisms of action reveals that these metals exert similar toxic effects: they produce ROS, enzyme inactivation, weaken antioxidant defenses, oxidative stress, and ultimately lead to carcinogenesis (Balali-Mood et al., 2021).

Currently, 168 pesticides are classified as potential carcinogens (Schwingl et al., 2021). Several carcinogenic pesticides have the potential to cause genitourinary, gastrointestinal, central nervous system, pulmonary system, renal, and endocrine glands cancer (Varghese et al., 2020; Cazzolla Gatti, 2021; Schwingl et al., 2021) are bioaccumulated in aquatic mammals (Table 2). Pesticide-mediated carcinogenesis may involve several mechanisms, including inflammation, oxidative stress, immunotoxicity, endocrine disruption, and genotoxicity (Gangemi et al., 2016). Although cetaceans are more anticancer resistant relative to other mammals (Tejada-Martinez et al., 2021), however, studies have reported neoplastic incidents in 40 species of aquatic mammals (Kitsoulis et al., 2020) and suggested cancer as one of the significant causes of death in aquatic mammals (Martineau et al., 2002; Gulland et al., 2020). For example, beluga whales exposed to a high concentration of polycyclic aromatic hydrocarbons in Canada have a percentage of cancer similar to humans (Martineau et al., 2002). Similarly, in California sea lions, the prevalence of urogenital cancer linked with organic chemical pollutants is one of the highest amongst mammals (Gulland et al., 2020). To our knowledge, till date, no studies are available linking MPs directly with cancer in aquatic mammals despite the rising concentration of MPs and

the increasing incidence of cancer in aquatic mammals. Although it is challenging to study cancer in aquatic mammals due to their inaccessibility, lack of exposure history to MPs and co-contaminants, and logistical, ethical and legal constraints on experimentation. However, understanding the link between cancer and MPs and its co-contaminant in aquatic mammals will not only help in biodiversity conservation, but due to their long lifespans, large body masses, and long-term exposure to MPs and associated chemical contaminants, aquatic mammals may represent an ideal animal model to understand the mechanism of carcinogenesis.

9. MPs-linked pathogens and risk of infectious diseases in aquatic mammals

MPs in the aquatic environments provide a novel microhabitat and protective ecological niche (plastisphere) with high selectivity and plasticity to a variety of distinct communities of pathogens, including antimicrobial-resistant pathogens (Amaral-Zettler et al., 2020; Bowley et al., 2021; Gkoutselis et al., 2021; Pham et al., 2021). These novel pathogens on MPs are dispersed over long distances by waves, currents, and winds to alien communities, transmitted to potential hosts, and can cause disease outbreaks (Bowley et al., 2021). The plastisphere acts as hotspots of antimicrobial resistance and houses 100–5000 times more antimicrobial resistance bacteria than the surrounding water (Yang et al., 2019; Zhang et al., 2020). Furthermore, the plastisphere communities enhance the metabolic pathways that contribute to infectious diseases (Sun et al., 2020). Therefore, the MPs not only act as a hub and a carrier for microbial pathogens but also enrich pathogenic strains through pathogenicity islands and antimicrobial properties

during horizontal gene transfer (Bowley et al., 2021). One study reported direct pathogen transfer via plastic (Rotjan et al., 2019). Many studies have postulated pathogen transfer via plastic in several organisms (Viršek et al., 2017; Lamb et al., 2018; Frère et al., 2018; Amaral-Zettler et al., 2020). However, there are no studies if pathogen-colonized MPs can transfer pathogens and cause disease in aquatic mammals even though aquatic mammals ingest a considerable quantity of MPs directly and indirectly via the food-web as an apex predator. From a conservation and wildlife management point of view, it is a vital area of research to understand the role of MPs as a pathogen vector, especially in the vulnerable populations of aquatic mammals native to the MPs dense regions.

Microorganisms on the plastisphere can use the added or adsorbed metals (e.g., iron, cobalt, copper, zinc, manganese) as micronutrients (Begg, 2019; Amaral-Zettler et al., 2020). These metals are also essential for the ultimate survival and virulence of many pathogenic microorganisms (Begg, 2019). MPs and loaded pollutants can cause pulmonary cytotoxicity, disrupt the protective pulmonary barriers (Dong et al., 2020; Facciola et al., 2021), and allow the entry of opportunistic and newly introduced pathogens to the deep parts of the lungs leading to respiratory infections and other infectious diseases in aquatic mammals (Nabi et al., 2020b; Bowley et al., 2021). Furthermore, metals such as mercury and cadmium have immunosuppressive effects and are linked with a higher rate of infectious diseases in aquatic mammals (Bennett et al., 2001; Pellissó et al., 2008). Currently, Labrado (2019) reported a positive correlation between a higher number of diseased animals and MPs per square kilometer in aquatic mammals. Similarly, Nelms et al. (2019) found that aquatic mammals that died due to infectious disease had a higher number of MPs relative to other drivers of mortality. Both viruses and bacteria that can cause infectious diseases, especially pneumonia, are the leading cause of death in aquatic mammals (Nabi et al., 2020b). During different infectious pneumonia outbreaks in the past, the mortality of thousands of individuals in different aquatic mammal species has been reported across the globe (Nabi et al., 2020b). All the evidence indicates that MPs in aquatic mammals can act as a vector for pathogen transmission. However, studies are required to understand the extent to which MPs can act as a vector for pathogens and toxicants from the aquatic environment into the tissues and their pathogenicity and virulence potential in aquatic mammals.

10. Conclusion

This review represents a warning signal of the rising level of MPs and their associated toxicants and pathogens in the aquatic environment and its potentially hazardous effects on aquatic top predator mammals. It suggests an urgent need for continuous monitoring to avoid reductions in population and biodiversity in both fresh-water and marine environments. Further studies are needed to deeply investigate the MPs burden globally in the aquatic environment, their accumulation in the different tissues of aquatic mammals, and possible physiological dysfunctions.

CRedit authorship contribution statement

Ghulam Nabi: Writing – original draft. **Shahid Ahmad:** Writing – review & editing. **Sana Ullah:** Writing – review & editing. **Sahib Zada:** Writing – review & editing. **Maliha Sarfraz:** Writing – review & editing. **Xinle Guo:** Funding acquisition. **Muhammad Ismail:** Writing – review & editing. **Kunyu Wanghe:** Funding acquisition.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This research was funded by Natural Science Foundation of Qinghai Province (2022-ZJ-936Q) and Investigation of Aquatic Biological Resources in the Aquatic Germplasm Resources Conservation Area of the Qinghai Section of the Yangtze River No. E039831D01.

References

- Alonso, M.B., Feo, M.L., Corcellas, C., Gago-Ferrero, P., Bertozzi, C.P., Marigo, J., Flach, L., Meirelles, A.C., Carvalho, V.L., Azevedo, A.F., Torres, J.P., 2015. Toxic heritage: Maternal transfer of pyrethroid insecticides and sunscreen agents in dolphins from Brazil. *Environ. Pollut.* 207, 391–402.
- Amaral-Zettler, L.A., Zettler, E.R., Mincer, T.J., 2020. Ecology of the plastisphere. *Nat. Rev. Microbiol.* 18, 139–151.
- Amereh, F., Babaei, M., Eslami, A., Fazelpour, S., Rafiee, M., 2020. The emerging risk of exposure to nano (micro) plastics on endocrine disturbance and reproductive toxicity: From a hypothetical scenario to a global public health challenge. *Environ. Pollut.* 261, 114158.
- Amin, M., Yousuf, M., Attaullah, M., Nabi, G., Buner, I.D., Ahmad, N., Ullah, U., Khan, Z., 2020. Comparative acute toxicity of organophosphates and synthetic pyrethroid pesticides in vivo exposed freshwater fish *Oreochromis niloticus* (Linnaeus, 1758). *Aquat. Ecosyst. Health. Manag.* 23, 366–372.
- Andrady, A.L., 2011. Microplastics in the marine environment. *Mar. Pollut. Bull.* 62, 1596–1605.
- Andrady, A.L., 2017. The plastic in microplastics: a review. *Mar. Poll. Bull.* 119, 12–22.
- Aubail, A., Méndez-Fernandez, P., Bustamante, P., Churlaud, C., Ferreira, M., Vingada, J.V., Caurant, F., 2013. Use of skin and blubber tissues of small cetaceans to assess the trace element content of internal organs. *Mar. Poll. Bull.* 76, 158–169.
- Avila, I.C., Kaschner, K., Dormann, C.F., 2018. Current global risks to marine mammals: taking stock of the threats. *Biol. Conserv.* 221, 44–58.
- Aznar-Alemany, Ò., Giménez, J., de Stephanis, R., Eljarrat, E., Barceló, D., 2017. Insecticide pyrethroids in liver of striped dolphin from the Mediterranean Sea. *Environ. Pollut.* 225, 346–353.
- Balali-Mood, M., Naseri, K., Tahergorabi, Z., Khazdair, M.R., Sadeghi, M., 2021. Toxic mechanisms of five heavy metals: mercury, lead, chromium, cadmium, and arsenic. *Front. Pharmacol.* 12, 643972.
- Barrick, A., Champeau, O., Chatel, A., Manier, N., Northcott, G., Tremblay, L.A., 2021. Plastic additives: challenges in ecotox hazard assessment. *Peer. J.* 9, e11300.
- Becker, P.R., 2000. Concentration of chlorinated hydrocarbons and heavy metals in Alaska arctic marine mammals. *Mar. Poll. Bull.* 40, 819–829.
- Beckmen, K.B., Lowenstine, L.J., Newman, J., Hill, J., Hanni, K., Gerber, J., 1997. Clinical and pathological characterization of northern elephant seal skin disease. *J. Wildl. Dis.* 33, 438–449.
- Begg, S.L., 2019. The role of metal ions in the virulence and viability of bacterial pathogens. *Biochem. Soc. Trans.* 47, 77–87.
- Bennett, P.M., Jepson, P.D., Law, R.J., Jones, B.R., Kuiken, T., Baker, J.R., Rogan, E., Kirkwood, J.K., 2001. Exposure to heavy metals and infectious disease mortality in harbour porpoises from England and Wales. *Environ. Pollut.* 112, 33–40.
- Bergmann, M., Gutow, L., Klages, M., 2015. Marine anthropogenic litter. Springer Nature.
- Besseling, E., Foekema, E.M., Van Franeker, J.A., Leopold, M.F., Kühn, S., Rebolledo, E. B., Heße, E., Mielke, L.J., Ijzer, J., Kamminga, P., Koelmans, A.A., 2015. Microplastic in a macro filter feeder: humpback whale Megaptera novaeangliae. *Mar. Poll. Bull.* 95, 248–252.
- Bossart, G.D., 2011. Marine mammals as sentinel species for oceans and human health. *Vet. Pathol.* 48, 676–690.
- Bowley, J., Baker-Austin, C., Porter, A., Hartnell, R., Lewis, C., 2021. Oceanic hitchhikers—assessing pathogen risks from marine microplastic. *Trends. Microbiol.* 29, 107–116.
- Braathén, M., Derocher, A.E., Wiig, Ø., Sørmo, E.G., Lie, E., Skaare, J.U., Jenssen, B.M., 2004. Relationships between PCBs and thyroid hormones and retinol in female and male polar bears. *Environ. Health. Perspect.* 112, 826–833.
- Bravo Rebolledo, E.L., Van Franeker, J.A., Jansen, O.E., Brasseur, S.M.J.M., 2013. Plastic ingestion by harbour seals (*Phoca vitulina*) in The Netherlands. *Mar. Poll. Bull.* 67, 200–202.
- Campanale, C., Massarelli, C., Savino, I., Locaputo, V., Uricchio, V.F., 2020. A detailed review study on potential effects of microplastics and additives of concern on human health. *Int. J. Environ. Res. Publ. Health.* 17, 1212.

- Cazzolla Gatti, R., 2021. Why we will continue to lose our battle with cancers if we do not stop their triggers from environmental pollution. *Int. J. Environ. Res. Publ. Health* 18, 6107.
- Chen, J., Su, Y., Lin, R., Lin, F., Shang, P., Hussain, R., Shi, D., 2021. Effects of acute diquat poisoning on liver mitochondrial apoptosis and autophagy in ducks. *Front. Vet. Sci.* 8, 727766.
- Cole, M., Lindeque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants in the marine environment: a review. *Mar. Poll. Bull.* 62, 2588–2597.
- Cruz, J.V.B., Santos, E.P., Silva, N.J., Lima, F.L.O., Martinelli, P.P., de Vasconcellos Neto, J.R.T., 2021. Influence of heavy metals on cancer accommodation: A literature review. *Res. Soc. Dev.* 10, e45810615992.
- Das, K., De Groof, A., Jauniaux, T., Bouqueneau, J.M., 2006. Zn, Cu, Cd and Hg binding to metallothioneins in harbour porpoises *Phocoena phocoena* from the southern North Sea. *BMC. Ecol.* 6, 1–7.
- De Sá, L.C., Oliveira, M., Ribeiro, F., Rocha, T.L., Futter, M.N., 2018. Studies of the effects of microplastics on aquatic organisms: what do we know and where should we focus our efforts in the future? *Sci. Total. Environ.* 645, 1029–1039.
- De Silva, A.O., Spencer, C., Ho, K.C., Al Tarhuni, M., Go, C., Houde, M., de Solla, S.R., Lavoie, R.A., King, L.E., Muir, D.C., Fair, P.A., 2016. Perfluoroalkylphosphonic acids in Northern pike (*Esox lucius*), double-crested cormorants (*Phalacrocorax auritus*), and bottlenose dolphins (*Tursiops truncatus*) in relation to other perfluoroalkyl acids. *Environ. Sci. Technol.* 50, 10903–10913.
- Delfosse, V., Huet, T., Harrus, D., Granell, M., Bourguet, M., Gardia-Parège, C., Chiavarina, B., Grimaldi, B., Le Mével, S., Blanc, P., Huang, D., Gruszczak, J., Demeneix, B., Cianféroni, S., Fini, J.B., Balaguer, P., Bourguet, W., 2021. Mechanistic insights into the synergistic activation of the RXR-PXR heterodimer by endocrine disruptor mixtures. *Proc. Natl. Acad. Sci. U. S. A.* 118, e2020551118.
- Derraik, J.G., 2002. The pollution of the marine environment by plastic debris: a review. *Mar. Pollut. Bull.* 44, 842–852.
- Desforjes, J.P., Hall, A., McConnell, B., Rosing-Asvid, A., Barber, J.L., Brownlow, A., De Guise, S., Eulaers, I., Jepson, P.D., Letcher, R.J., Levin, M., 2018. Predicting global killer whale population collapse from PCB pollution. *Science* 361, 1373–1376.
- Dong, C.D., Chen, C.W., Chen, Y.C., Chen, H.H., Lee, J.S., Lin, C.H., 2020. Polystyrene microplastic particles: In vitro pulmonary toxicity assessment. *J. Hazard. Mater.* 385, 121575.
- Donohue, M.J., Masura, J., Gelatt, T., Ream, R., Baker, J.D., Faulhaber, K., Lerner, D.T., 2019. Evaluating exposure of northern fur seals, *Callorhinus ursinus*, to microplastic pollution through fecal analysis. *Mar. Poll. Bull.* 138, 213–221.
- Du, S., Zhu, R., Cai, Y., Xu, N., Yap, P.S., Zhang, Y., He, Y., Zhang, Y., 2021. Environmental fate and impacts of microplastics in aquatic ecosystems: a review. *RSC. Adv.* 11, 15762–15784.
- Eriksson, C., Burton, H., 2003. Origins and biological accumulation of small plastic particles in fur seals from Macquarie Island. *AMBIO* 32, 380–384.
- Facciola, A., Visalli, G., Pruiti Ciarello, M., Di Pietro, A., 2021. Newly emerging airborne pollutants: current knowledge of health impact of micro and nanoplastics. *Int. J. Environ. Res. Public Health* 18, 2997.
- Fair, P.A., Adams, J., Mitchum, G., Hulsey, T.C., Reif, J.S., Houde, M., Muir, D., Wirth, E., Wetzel, D., Zolman, E., McFee, W., 2010. Contaminant blubber burdens in Atlantic bottlenose dolphins (*Tursiops truncatus*) from two southeastern US estuarine areas: Concentrations and patterns of PCBs, pesticides, PBDEs, PFCs, and PAHs. *Sci. Total. Environ.* 408, 1577–1597.
- Fossi, M.C., Coppola, D., Baini, M., Giannetti, M., Guerranti, C., Marsili, L., Panti, C., de Sabata, E., Clò, S., 2014. Large filter feeding marine organisms as indicators of microplastic in the pelagic environment: the case studies of the Mediterranean basking shark (*Cetorhinus maximus*) and fin whale (*Balaenoptera physalus*). *Mar. Environ. Res.* 100, 17–24.
- Fossi, M.C., Marsili, L., 2003. Effects of endocrine disruptors in aquatic mammals. *Pure. Appl. Chem.* 75, 2235–2247.
- Fossi, M.C., Panti, C., Baini, M., Lavers, J.L., 2018. A review of plastic-associated pressures: cetaceans of the Mediterranean Sea and eastern Australian shearwaters as case studies. *Front. Mar. Sci.* 5, 173.
- Fossi, M.C., Marsili, L., Baini, M., Giannetti, M., Coppola, D., Guerranti, C., Caliani, I., Minutoli, R., Lauriano, G., Finoia, M.G., Rubegni, F., 2016. Fin whales and microplastics: The Mediterranean Sea and the Sea of Cortez scenarios. *Environ. Pollut.* 209, 68–78.
- S.B. Fournier, J.N. D'Errico, D.S. Adler, S. Kollontzi, M.J. Goedken, L. Fabris, E.J. Yurkow, P. A. Stapleton Nanopolystyrene translocation and fetal deposition after acute lung exposure during late-stage pregnancy. *Part. Fibre. Toxicol.* 17 2020 1 1.
- Frère, L., Maignien, L., Chalopin, M., Huvet, A., Rinnert, E., Morrison, H., Kerninon, S., Cassone, A.L., Lambert, C., Reveillaud, J., Paul-Pont, I., 2018. Microplastic bacterial communities in the Bay of Brest: Influence of polymer type and size. *Environ. Pollut.* 242, 614–625.
- Frodello, J.P., Marchand, B., 2001. Cadmium, copper, lead, and zinc in five toothed whale species of the Mediterranean Sea. *Int. J. Toxicol.* 20, 339–343.
- Gallicchio, V.S., Harper, J., 2021. Role of heavy metals in the incidence of human cancers. *IntechOpen*. <https://doi.org/10.5772/intechopen.98259>.
- Galligan, T.M., Balmer, B.C., Schwacke, L.H., Bolton, J.L., Quigley, B.M., Rosel, P.E., Ylitalo, G.M., Boggs, A.S., 2019. Examining the relationships between blubber steroid hormones and persistent organic pollutants in common bottlenose dolphins. *Environ. Pollut.* 249, 982–991.
- Galloway, T.S., Cole, M., Lewis, C., 2017. Interactions of microplastic debris throughout the marine ecosystem. *Nat. Ecol. Evol.* 1, 1–8.
- Gangemi, S., Gofita, E., Costa, C., Teodoro, M., Briguglio, G., Nikitovic, D., Tzanakakis, G., Tsatsakis, A.M., Wilks, M.F., Spandidos, D.A., Fenga, C., 2016. Occupational and environmental exposure to pesticides and cytokine pathways in chronic diseases. *Internat. J. Mol. Med.* 38, 1012–1020.
- Gao, H.T., Xu, R., Cao, W.X., Qian, L.L., Wang, M., Lu, L., Xu, Q., Yu, S.Q., 2017. Effects of six priority-controlled phthalate esters with long-term low-dose integrated exposure on male reproductive toxicity in rats. *Food. Chem. Toxicol.* 101, 94–104.
- Gerpe, M., Rodríguez, D., Moreno, V., Bastida, R., Aizpãn, J., 2007. Copper, zinc, cadmium, and mercury in Southern sea lions (*Otaria flavescens*) from Argentina. *Aquat. Mamm.* 33, 223.
- Gkoutselis, G., Rohrbach, S., Harjes, J., Obst, M., Brachmann, A., Horn, M.A., Rambold, G., 2021. Microplastics accumulate fungal pathogens in terrestrial ecosystems. *Sci. Rep.* 11, 1–3.
- Gui, D., Yu, R.Q., Karczmarski, L., Ding, Y., Zhang, H., Sun, Y., Zhang, M., Wu, Y., 2017. Spatiotemporal trends of heavy metals in Indo-Pacific humpback dolphins (*Sousa chinensis*) from the western Pearl River Estuary. *China. Environ. Sci. Technol.* 51, 1848–1858.
- Gulland, F., Hall, A.J., Ylitalo, G.M., Colegrove, K.M., Norris, T., Duignan, P.J., Halaska, B., Acevedo Whitehouse, K., Lowenstine, L.J., Deming, A.C., Rowles, T.K., 2020. Persistent contaminants and herpesvirus OthV1 are positively associated with cancer in wild California sea lions (*Zalophus californianus*). *Front. Mar. Sci.* 7, 1093.
- Guo, D., Liu, W., Yao, T., Ma, M., Wang, Q., Qiu, J., Qian, Y., 2021. Combined endocrine disruptive toxicity of malathion and cypermethrin to gene transcription and hormones of the HPG axis of male zebrafish (*Danio rerio*). *Chemosphere* 267, 128864.
- Haave, M., Gomiero, A., Schönheit, J., Nilsen, H., Olsen, A.B., 2021. Documentation of microplastics in tissues of wild coastal animals. *Front. Environ. Sci.* 9, 575058.
- Haave, M., Ropstad, E., Derocher, A.E., Lie, E., Dahl, E., Wiig, Ø., Skaare, J.U., Jenssen, B.M., 2003. Polychlorinated biphenyls and reproductive hormones in female polar bears at Svalbard. *Environ. Health Perspect.* 111, 431–436.
- Hall, A.J., Hugunin, K., Deaville, R., Law, R.J., Allchin, C.R., Jepson, P.D., 2006. The risk of infection from polychlorinated biphenyl exposure in the harbor porpoise (*Phocoena phocoena*): a case-control approach. *Environ. Health Perspect.* 114, 704–711.
- Hall, A.J., Kalantzi, O.I., Thomas, G.O., 2003. Polybrominated diphenyl ethers (PBDEs) in grey seals during their first year of life—are they thyroid hormone endocrine disruptors? *Environ. Pollut.* 126, 29–37.
- Halpern, B.S., Walbridge, S., Selkoe, K.A., Kappel, C.V., Micheli, F., D'Agrosa, C., Bruno, J.F., Casey, K.S., Ebert, C., Fox, H.E., Fujita, R., 2008. A global map of human impact on marine ecosystems. *Science* 319, 948–952.
- Hamid, N., Junaid, M., Pei, D.S., 2021. Combined toxicity of endocrine-disrupting chemicals: A review. *Ecotoxicol. Environ. Saf.* 215, 112136.
- Hansen, A.M., Bryan, C.E., West, K., Jensen, B.A., 2016. Trace element concentrations in liver of 16 species of cetaceans stranded on Pacific Islands from 1997 through 2013. *Arch. Environ. Contam. Toxicol.* 70, 75–95.
- Haynes, D., Carter, S., Gaus, C., Müller, J., Dennison, W., 2005. Organochlorine and heavy metal concentrations in blubber and liver tissue collected from Queensland (Australia) dugong (*Dugong dugon*). *Mar. Pollut. Bull.* 51, 361–369.
- He, S., Li, P., Li, Z.H., 2021. Review on endocrine disrupting toxicity of triphenyltin from the perspective of species evolution: Aquatic, amphibious and mammalian. *Chemosphere* 269, 128711.
- Huang, Z., Weng, Y., Shen, Q., Zhao, Y., Jin, Y., 2021. Microplastic: A potential threat to human and animal health by interfering with the intestinal barrier function and changing the intestinal microenvironment. *Science. Total. Environ* 27, 147365.
- Karami-Mohajeri, S., Ahmadipour, A., Rahimi, H.R., Abdollahi, M., 2017. Adverse effects of organophosphorus pesticides on the liver: a brief summary of four decades of research. *Arch. Higijena. Rada. i toksikologiju* 68, 261–275.
- Routti, H., Harju, M., Lühmann, K., Aars, J., Ask, A., Goksøyr, A., Kovacs, K.M., Lydersen, C., 2021. Concentrations and endocrine disruptive potential of phthalates in marine mammals from the Norwegian Arctic. *Environ. Int.* 152, 106458.
- Hernandez-Gonzalez, A., Saavedra, C., Gago, J., Covelo, P., Santos, M.B., Pierce, G.J., 2018. Microplastics in the stomach contents of common dolphin (*Delphinus delphis*) stranded on the Galician coasts (NW Spain, 2005–2010). *Mar. Pollut. Bull.* 137, 526–532.
- Hocking, D.P., Marx, F.G., Park, T., Fitzgerald, E.M., Evans, A.R., 2017. A behavioural framework for the evolution of feeding in predatory aquatic mammals. *Proc. Royal. Soc. B* 284, 20162750.
- Hudak, C.A., Sette, L., 2019. Opportunistic detection of anthropogenic micro debris in harbor seal (*Phoca vitulina vitulina*) and gray seal (*Halichoerus grypus atlantica*) fecal samples from haul-outs in southeastern Massachusetts. *USA. Mar. Pollut. Bull.* 145, 390–395.
- Ikemoto, T., Kunito, T., Tanaka, H., Baba, N., Miyazaki, N., Tanabe, S., 2004. Detoxification mechanism of heavy metals in marine mammals and seabirds: interaction of selenium with mercury, silver, copper, zinc, and cadmium in liver. *Arch. Environ. Contam. Toxicol.* 47, 402–413.
- Isobe, A., Azuma, T., Cordova, M.R., Cózar, A., Galgani, F., Hagita, R., Kanhai, L.D., Imai, K., Iwasaki, S., Kako, S.I., Kozlovskii, N., 2021. A multilevel dataset of microplastic abundance in the world's upper ocean and the Laurentian Great Lakes. *Microplastics and Nanoplastics* 1, 1–4.

- Jambeck, J.R., Geyer, R., Wilcox, C., Siegler, T.R., Perryman, M., Andrady, A., Narayan, R., Law, K.L., 2015. Plastic waste inputs from land into the ocean. *Science*. 347, 768–771.
- Kannan, K., Ramu, K., Kajiwar, N., Sinha, R.K., Tanabe, S., 2005. Organochlorine pesticides, polychlorinated biphenyls, and polybrominated diphenyl ethers in Irrawaddy dolphins from India. *Arch. Environ. Contam. Toxicol.* 49, 415–420.
- Kassotis, C.D., Trasande, L., 2021. Endocrine disruptor global policy. *Adv. Pharmacol.* 92, 1–34.
- Kedzierski, M., d'Almeida, M., Magueresse, A., Le Grand, A., Duval, H., César, G., Sire, O., Bruzaud, S., Le Tilly, V., 2018. Threat of plastic ageing in marine environment. Adsorption/desorption of micropollutants. *Mar. Pollut. Bull.* 127, 684–694.
- Kimberley, A.B., Robinson, K.J., Armstrong, H.C., Moss, S.E.W., Scholl, G., Tranganida, A., Eppe, G., Thomé, J.P., Debier, C., Hall, A.J., 2021. Predicting consequences of POP-induced disruption of blubber glucose uptake, mass gain rate and thyroid hormone levels for weaning mass in grey seal pups. *Environ. Int.* 152, 106506.
- Kitsoulis, C.V., Baxevanis, A.D., Abatzopoulos, T.J., 2020. The occurrence of cancer in vertebrates: a mini review. *J. Biol. Res.* 27, 1–2.
- Kojima, H., Katsura, E., Takeuchi, S., Niiyama, K., Kobayashi, K., 2004. Screening for estrogen and androgen receptor activities in 200 pesticides by in vitro reporter gene assays using Chinese hamster ovary cells. *Environ. Health Perspect.* 112, 524–531.
- Labrado, M.L. (2019). Influence of marine pollution on infectious diseases in marine mammals: review and data analysis. Master Thesis, Marine Conservation, Universidad De Oviedo, Oviedo, Spain.
- Lamb, J.B., Willis, B.L., Fiorenza, E.A., Couch, C.S., Howard, R., Rader, D.N., True, J.D., Kelly, L.A., Ahmad, A., Jompa, J., Harvell, C.D., 2018. Plastic waste associated with disease on coral reefs. *Science*. 359, 460–462.
- Lauretta, R., Sansone, A., Sansone, M., Romanelli, F., Appetecchia, M., 2019. Endocrine disrupting chemicals: effects on endocrine glands. *Front. Endocrinol.* 21 (10), 178.
- Lee, H., Lee, H.J., Kwon, J.H., 2019. Estimating microplastic-bound intake of hydrophobic organic chemicals by fish using measured desorption rates to artificial gut fluid. *Sci. Total. Environ.* 651, 162–170.
- López-Rodríguez, D., Aylwin, C.F., Delli, V., Sevrin, E., Campanile, M., Martin, M., Franssen, D., Gérard, A., Blacher, S., Tirelli, E., Noël, A., 2021. Multi-and transgenerational outcomes of an exposure to a mixture of endocrine-disrupting chemicals (EDCs) on puberty and maternal behavior in the female rat. *Environ. Health Perspect.* 129, 087003.
- Lusher, A.L., O'Donnell, C., Officer, R., O'Connor, I., 2016. Microplastic interactions with North Atlantic mesopelagic fish. *ICES J. Mar. Sci.* 73, 1214–1225.
- Lusher, A.L., Hernandez-Milian, G., Berrow, S., Rogan, E., O'Connor, I., 2018. Incidence of marine debris in cetaceans stranded and bycaught in Ireland: Recent findings and a review of historical knowledge. *Environ. Pollut.* 232, 467–476.
- Lusher, A.L., Hernandez-Milian, G., O'Brien, J., Berrow, S., O'Connor, I., Officer, R., 2015. Microplastic and macroplastic ingestion by a deep diving, oceanic cetacean: The True's beaked whale *Mesoplodon mirus*. *Environ. Pollut.* 199, 185–191.
- Mackintosh, S.A., Dodder, N.G., Shaul, N.J., Aluwihare, L.I., Maruya, K.A., Chivers, S.J., Danil, K., Weller, D.W., Hoh, E., 2016. Newly identified DDT-related compounds accumulating in Southern California bottlenose dolphins. *Environ. Sci. Technol.* 50, 12129–12137.
- Martineau, D., Lemberger, K., Dallaire, A., Labelle, P., Lipscomb, T.P., Michel, P., Mikaelian, I., 2002. Cancer in wildlife, a case study: beluga from the St. Lawrence estuary, Québec, Canada. *Environ. Health Perspect.* 110, 285–292.
- Meng, X., Zhang, J., Wang, W., Gonzalez-Gil, G., Vrouwenvelder, J.S., Li, Z., 2022. Effects of nano-and microplastics on kidney: Physicochemical properties, bioaccumulation, oxidative stress and immunoreaction. *Chemosphere* 288, 132631.
- McHugh, B., Law, R.J., Allchin, C.R., Rogan, E., Murphy, S., Foley, M.B., Glynn, D., McGovern, E., 2007. Bioaccumulation and enantiomeric profiling of organochlorine pesticides and persistent organic pollutants in the killer whale (*Orcinus orca*) from British and Irish waters. *Mar. Pollut. Bull.* 54, 1724–1731.
- Meaza, I., Toyoda, J.H., Wise, S.J.P., 2021. Microplastics in sea turtles, marine mammals and humans: A one environmental health perspective. *Front. Environ. Sci.* 8, 575614.
- Mendes, A.M., Golden, N., Bermejo, R., Morrison, L., 2021. Distribution and abundance of microplastics in coastal sediments depends on grain size and distance from sources. *Mar. Pollut. Bull.* 172, 112802.
- Meyer, W.K., Jamison, J., Richter, R., Woods, S.E., Partha, R., Kowalczyk, A., Kronk, C., Chikina, M., Bonde, R.K., Crocker, D.E., Gaspard, J., 2018. Ancient convergent losses of Paraoxonase 1 yield potential risks for modern marine mammals. *Science*. 361, 591–594.
- Moore, R.C., Loseto, L., Noel, M., Etemadifar, A., Brewster, J.D., MacPhee, S., Bendell, L., Ross, P.S., 2020. Microplastics in beluga whales (*Delphinapterus leucas*) from the Eastern Beaufort Sea. *Mar. Pollut. Bull.* 150, 110723.
- Moore, S.E., 2008. Marine mammals as ecosystem sentinels. *J. Mammal.* 89, 534–540.
- Murphy, S., Barber, J.L., Learmonth, J.A., Read, F.L., Deaville, R., Perkins, M.W., Brownlow, A., Davison, N., Penrose, R., Pierce, G.J., Law, R.J., Jepson, P.D., 2015. Reproductive failure in UK harbour porpoises *Phocoena phocoena*: Legacy of pollutant exposure? *Plos. One*. 10, e0131085.
- Murphy, S., Law, R.J., Deaville, R., Barnett, J., Perkins, M.W., Brownlow, A., Penrose, R., Davison, N.J., Barber, J.L., Jepson, P.D. (2018). Organochlorine contaminants and reproductive implication in cetaceans: a case study of the common dolphin. In: *Marine mammal ecotoxicology*, 3–8.
- Nabi, G., Hao, Y., Zeng, X., Jinsong, Z., McLaughlin, R.W., Wang, D., 2017a. Hematologic and biochemical differences between two free ranging Yangtze finless porpoise populations: The implications of habitat. *Plos. One*. 12, e0188570.
- Nabi, G., Hao, Y., Zeng, X., Wang, D., 2017b. Assessment of Yangtze finless porpoises (*Neophocaena asiaorientalis*) through biochemical and hematological parameters. *Zool. Stud* 56, e31.
- Nabi, G., Hao, Y., McLaughlin, R.W., Wang, D. (2018a). The possible effects of high vessel traffic on the physiological parameters of the critically endangered Yangtze Finless Porpoise (*Neophocaena asiaorientalis* ssp. *asiaorientalis*). *Front. Physiol.* 9, 1665.
- Nabi, G., McLaughlin, R.W., Hao, Y., Wang, K., Zeng, X., Khan, S., Wang, D., 2018. The possible effects of anthropogenic acoustic pollution on marine mammals' reproduction: an emerging threat to animal extinction. *Environ. Sci. Pollut. Res. Internat.* 25, 19338–19345.
- Nabi, G., Robeck, T.R., Hao, Y., Wang, D., 2019. Hematologic and biochemical reference interval development and the effect of age, sex, season, and location on hematologic analyte concentrations in critically endangered Yangtze finless porpoise (*Neophocaena asiaorientalis* ssp. *asiaorientalis*). *Front. Physiol.* 10, 792.
- Nabi, G., Li, Y., McLaughlin, R.W., Mei, Z., Wang, K., Hao, Y., Zheng, J., Wang, D., 2020a. Immune responses of the critically endangered Yangtze finless porpoises (*Neophocaena asiaorientalis* ssp. *asiaorientalis*) to escalating anthropogenic stressors in the wild and seminatural environments. *Front. Physiol.* 10, 1594.
- Nabi, G., McLaughlin, R.W., Khan, S., Hao, Y., Chang, M.X., 2020b. Pneumonia in endangered aquatic mammals and the need for developing low-coverage vaccination for their management and conservation. *Anim. Health Res. Rev.* 21, 122–130.
- Nabi, G., Robeck, T.R., Yujiang, H., Tang, B., Zheng, J., Wang, K., Wang, D., 2021. Circulating concentrations of thyroid hormones and cortisol in wild and semi-natural Yangtze finless porpoise (*Neophocaena asiaorientalis*). *Conserv. Physiol.* 9, p.coab034.
- Nelms, S.E., Barnett, J., Brownlow, A., Davison, N.J., Deaville, R., Galloway, T.S., Lindeque, P.K., Santillo, D., Godley, B.J., 2019. Microplastics in marine mammals stranded around the British coast: ubiquitous but transitory? *Sci. Rep.* 9, 1–8.
- Nelms, S.E., Galloway, T.S., Godley, B.J., Jarvis, D.S., Lindeque, P.K., 2018. Investigating microplastic trophic transfer in marine top predators. *Environ. Pollut.* 238, 999–1007.
- Nielsen, J.B., Nielsen, F., Jørgensen, P.J., Grandjean, P., 2000. Toxic metals and selenium in blood from pilot whales (*Globicephala melas*) and sperm whales (*Physeter catodon*). *Mar. Pollut. Bull.* 40, 348–351.
- Niño-Torres, C.A., Zenteno-Savín, T., Gardner, S.C., Urbán, R.J., 2010. Organochlorine pesticides and polychlorinated biphenyls in fin whales (*Balaenoptera physalus*) from the Gulf of California. *Environ. Toxicol.* 52, 381–390.
- Oskam, I., Ropstad, E., Lie, E., Derocher, A., Wiig, Ø., Dahl, E., Larsen, S., Skaare, J.U., 2004. Organochlorines affect the steroid hormone cortisol in free-ranging polar bears (*Ursus maritimus*) at Svalbard, Norway. *J. Toxicol. Environ. Health.* 67, 959–977.
- Oskam, I.C., Ropstad, E., Dahl, E., Lie, E., Derocher, A.E., Wiig, Y., Larsen, S., Wiger, R., Skaare, J.U., 2003. Organochlorines affect the major androgenic hormone, testosterone, in male polar bears (*Ursus maritimus*) at Svalbard. *J. Toxicol. Environ. Health.* A. 66, 2119–2139.
- Osorio, E.D., Tanchuling, M.A.N., Diola, M.B.L.D., 2021. Microplastics occurrence in surface waters and sediments in five river mouths of Manila Bay. *Front. Environ. Sci.* 9, 719274.
- Page-Karjian, A., Lo, C.F., Ritchie, B., Harms, C.A., Rotstein, D.S., Han, S., Hassan, S.M., Lehner, A.F., Buchweitz, J.P., Thayer, V.G., Sullivan, J.M., Christiansen, E.F., Perrault, J.R., 2020. Anthropogenic contaminants and histopathological findings in stranded cetaceans in the Southeastern United States, 2012–2018. *Front. Mar. Sci.* 7, 630.
- Park, C., Song, H., Choi, J., Sim, S., Kojima, H., Park, J., Iida, M., Lee, Y., 2020. The mixture effects of bisphenol derivatives on estrogen receptor and androgen receptor. *Environ. Pollut.* 260, 114036.
- Peeken, I., Primpke, S., Beyer, B., Gütermann, J., Katlein, C., Krumpfen, T., Bergmann, M., Hehemann, L., Gerdt, G., 2018. Arctic sea ice is an important temporal sink and means of transport for microplastic. *Nat. Commun.* 9, 1–2.
- Pellissó, S.C., Munoz, M.J., Carballo, M., Sanchez-Vizcaino, J.M., 2008. Determination of the immunotoxic potential of heavy metals on the functional activity of bottlenose dolphin leukocytes in vitro. *Vet. Immunol. Immunopathol.* 121, 189–198.
- Peng, Y., Wu, P., Schartup, A.T., Zhang, Y., 2021. Plastic waste release caused by COVID-19 and its fate in the global ocean. *Proc. Natl. Acad. Sci. U.S.A.* 118, e2111530118.
- Perez-Venegas, D.J., Seguel, M., Pavés, H., Pulgar, J., Urbina, M., Ahrendt, C., Galbán-Malagón, C., 2018. First detection of plastic microfibers in a wild population of South American fur seals (*Arctocephalus australis*) in the Chilean Northern Patagonia. *Mar. Pollut. Bull.* 136, 50–54.
- Perez-Venegas, D.J., Toro-Valdivieso, C., Ayala, F., Brito, B., Iturra, L., Arriagada, M., Seguel, M., Barrios, C., Sepúlveda, M., Oliva, D., Cárdenas-Alayza, S., Urbina, M.A., Jorquera, A., Castro-Nallar, E., Galbán-Malagón, C., 2020. Monitoring the occurrence of microplastic ingestion in Otariids along the Peruvian and Chilean coasts. *Mar. Pollut. Bull.* 153, 110966.
- Pham, D.N., Clark, L., Li, M., 2021. Microplastics as hubs enriching antibiotic-resistant bacteria and pathogens in municipal activated sludge. *J. Hazard. Mater. Lett.* 2, 100014.

- Preston, E.V., Webster, T.F., Henn, B.C., McClean, M.D., Gennings, C., Oken, E., Rifas-Shiman, S.L., Pearce, E.N., Calafat, A.M., Fleisch, A.F., Sagiv, S.K., 2020. Prenatal exposure to per-and polyfluoroalkyl substances and maternal and neonatal thyroid function in the Project Viva Cohort: A mixtures approach. *Environ. Int.* 139, 105728.
- Prieti, B., Meindl, C., Roblegg, E., Pieber, T., Lanzer, G., Fröhlich, E., 2014. Nano-sized and micro-sized polystyrene particles affect phagocyte function. *Cell. Biol. Toxicol.* 30, 1–16.
- Ragusa, A., Svelato, A., Santacroce, C., Catalano, P., Notarstefano, V., Carnevali, O., Papa, F., Rongioletti, M.C., Baiocco, F., Draghi, S., D'Amore, E., 2021. Plasticenta: First evidence of microplastics in human placenta. *Environ. Int.* 146, 106274.
- Reijnders, P.J. (1999). Reproductive and developmental effects of endocrine-disrupting chemicals on marine mammals. In: *Marine mammals and persistent ocean contaminants: proceedings of the marine mammal commission workshop*, Keystone, Colorado, 12–15 October 1998, pp. 93–100.
- Rodprasert, W., Toppari, J., Virtanen, H.E., 2021. Endocrine disrupting chemicals and reproductive health in boys and men. *Front. Endocrinol.* 12, 706532.
- Ropstad, E., Oskam, I.C., Lyche, J.L., Larsen, H.J., Lie, E., Haave, M., Dahl, E., Wiger, R., Skaare, J.U., 2006. Endocrine disruption induced by organochlorines (OCs): field studies and experimental models. *J. Toxicol. Environ. Health. A* 69, 53–76.
- Ross, P.S., Chastain, S., Vassilenko, E., Etemadifar, A., Zimmermann, S., Quesnel, S.A., Eert, J., Solomon, E., Patankar, S., Posacka, A.M., Williams, B., 2021. Pervasive distribution of polyester fibres in the Arctic Ocean is driven by Atlantic inputs. *Nat. Commun.* 12, 1–9.
- Rotjan, R.D., Sharp, K.H., Gauthier, A.E., Yelton, R., Lopez, E.M., Carilli, J., Kagan, J.C., Urban-Rich, J., 2019. Patterns, dynamics and consequences of microplastic ingestion by the temperate coral, *Astrangia poculata*. *Proc. Royal. Soc. B* 286, 20190726.
- Routti, H., Diot, B., Panti, C., Duale, N., Fossi, M.C., Harju, M., Kovacs, K.M., Lydersen, C., Scotter, S.E., Villanger, G.D., Bourgeon, S., 2019. Contaminants in Atlantic walrus in Svalbard Part 2: Relationships with endocrine and immune systems. *Environ. Pollut.* 246, 658–667.
- Salgado-Freiria, R., López-Doval, S., Lafuente, A., 2018. Perfluorooctane sulfonate (PFOS) can alter the hypothalamic-pituitary-adrenal (HPA) axis activity by modifying CRF1 and glucocorticoid receptors. *Toxicol. Lett.* 295, 1–9.
- Schjenken, J.E., Green, E.S., Overduin, T.S., Mah, C.Y., Russell, D.L., Robertson, S.A., 2021. Endocrine disruptor compounds—A cause of impaired immune tolerance driving inflammatory disorders of pregnancy? *Front. Endocrinol.* 12, 607539.
- Schwingl, P.J., Lunn, R.M., Mehta, S.S., 2021. A tiered approach to prioritizing registered pesticides for potential cancer hazard evaluations: implications for decision making. *Environ. Health.* 20, 1–4.
- Shelver, W., Banerjee, A., 2021. Micro/nanoplastic mediated pathophysiological changes in rodents, rabbits, and chicken. *J. Food. Prot.* 84, 1480–1495.
- Siegal-Willott, J.L., Harr, K.E., Hall, J.O., Hayek, L.A., Aulí-Gómez, N., Powell, J.A., Bonde, R.K., Heard, D., 2013. Blood mineral concentrations in manatees (*Trichechus manatus latirostris* and *Trichechus manatus manatus*). *J. Zoo. Wildl. Med.* 44, 285–294.
- Singh, N., Gupta, V.K., Kumar, A., Sharma, B., 2017. synergistic effects of heavy metals and pesticides in living systems. *Front. Chem.* 5, 70.
- Skaare, J.U., Bernhoft, A., Wiig, O., Norum, K.R., Haug, E., Eide, D.M., Derocher, A.E., 2001. Relationships between plasma levels of organochlorines, retinol and thyroid hormones from polar bears (*Ursus maritimus*) at Svalbard. *J. Toxicol. Environ. Health. A* 62, 227–241.
- Sonne, C., Siebert, U., Gonnens, K., Desforges, J.P., Eulaers, I., Persson, S., Roos, A., Bäcklin, B.M., Kauhala, K., Olsen, M.T., Harding, K.C., 2010. Health effects from contaminant exposure in Baltic Sea birds and marine mammals: A review. *Environ. Int.* 139, 105725.
- Stockin, K.A., Law, R.J., Roe, W.D., Meynier, L., Martinez, E., Duignan, P.J., Bridgen, P., Jones, B., 2010. PCBs and organochlorine pesticides in Hector's (*Cephalorhynchus hectori hectori*) and Maui's (*Cephalorhynchus hectori maui*) dolphins. *Mar. Pollut. Bull.* 60, 834–842.
- J. Sun S. Yang G.J. Zhou K. Zhang Y. Lu Q. Jin P.K. Lam K.M. Leung Y. He Release of microplastics from discarded surgical masks and their adverse impacts on the marine copepod *Tigriopus japonicus* 2021 *Sci. Technol. Lett Environ* 10.1021/acs.estlett.1c00748.
- Sun, X., Chen, B., Xia, B., Li, Q., Zhu, L., Zhao, X., Gao, Y., Qu, K., 2020. Impact of mariculture-derived microplastics on bacterial biofilm formation and their potential threat to mariculture: A case in situ study on the Sungo Bay, China. *Environ. Pollut.* 262, 114336.
- Takeuchi, N.Y., Walsh, M.T., Bonde, R.K., Powell, J.A., Bass, D.A., Gaspard III, J.C., Barber, D.S., 2016. Baseline reference range for trace metal concentrations in whole blood of wild and managed west Indian manatees (*Trichechus manatus*) in Florida and Belize. *Aquat. Mamm.* 42, 440.
- Teigen, S.W., Andersen, R.A., Daee, H.L., Skaare, J.U., 1999. Heavy metal content in liver and kidneys of grey seals (*Halichoerus grypus*) in various life stages correlated with metallothionein levels: Some metal-binding characteristics of this protein. *Environ. Toxicol. Chem.* 18, 2364–2369.
- Tejada-Martinez, D., de Magalhães, J.P., Opazo, J.C., 2021. Positive selection and gene duplications in tumour suppressor genes reveal clues about how cetaceans resist cancer. *Proc. Royal. Soc. B* 288, 20202592.
- Ter Halle, A., Jeanneau, L., Martignac, M., Jardé, E., Pedrono, B., Brach, L., Gigault, J., 2017. Nanoplastic in the North Atlantic subtropical gyre. *Environ. Sci. Technol.* 51, 13689–13697.
- Tian, X., Li, L., 2010. Combined toxic effects of di-n-butyl phthalate and di-2-ethylhexyl phthalate on fertility in male rats. *J. Environ. Health.* 27, 290–292.
- Trego, M.L., Hoh, E., Kellar, N.M., Meszaros, S., Robbins, M.N., Dodder, N.G., Whitehead, A., Lewison, R.L., 2018. comprehensive screening links halogenated organic compounds with testosterone levels in male *Delphinus delphis* from the Southern California Bight. *Environ. Sci. Technol.* 52, 3101–3109.
- Trego, M.L., Hoh, E., Whitehead, A., Kellar, N.M., Lauf, M., Datuin, D.O., Lewison, R.L., 2019. Contaminant exposure linked to cellular and endocrine biomarkers in Southern California bottlenose dolphins. *Environ. Sci. Technol.* 53, 3811–3822.
- Troisi, G.M., Mason, C.F., 2000. PCB-associated alteration of hepatic steroid metabolism in harbor seals (*Phoca vitulina*). *J. Toxicol. Environ. Health. A* 61, 649–655.
- Troisi, G.M., Barton, S.J., Liori, O., Nyman, M., 2020. Polychlorinated biphenyls (PCBs) and sex hormone concentrations in ringed and grey seals: A possible link to endocrine disruption? *Arch. Environ. Contam. Toxicol.* 78, 513–524.
- Ugwu, K., Herrera, E., Gómez, M., 2021. Microplastics in marine biota: A review. *Mar. Pollut. Bull.* 169, 112540.
- Ullah, S., Li, Z., Hassan, S., Ahmad, S., Guo, X., Wanghe, K., Nabi, G., 2021a. Heavy metals bioaccumulation and subsequent multiple biomarkers-based appraisal of toxicity in the critically endangered *Tor putitora*. *Ecotoxicol. Environ. Saf.* 228, 113032.
- Ullah, S., Ahmad, S., Altaf, Y., Dawar, F.U., Anjum, S.I., Baig, M.M., Fahad, S., Al-Misned, F., Atique, U., Guo, X., Nabi, G., 2021b. Bifenthrin induced toxicity in *Ctenopharyngodon idella* at an acute concentration: A multi-biomarkers-based study. *J. King. Saud. Univ. Sci.* 101752.
- Van Franeker, J.A., Rebolledo, E.L., Hesse, E., Ijseldijk, L.L., Kühn, S., Leopold, M., Mielke, L., 2018. Plastic ingestion by harbour porpoises *Phocoena phocoena* in the Netherlands: Establishing a standardised method. *Ambio* 47, 387–397.
- Van Sebille, E., Wilcox, C., Lebreton, L., Maximenko, N., Hardesty, B.D., Van Franeker, J.A., Eriksen, M., Siegel, D., Galgani, F., Law, K.L., 2015. A global inventory of small floating plastic debris. *Environ. Res. Lett.* 10, 124006.
- Varghese, J.V., Sebastian, E.M., Iqbal, T., Tom, A.A., 2020. Pesticide applicators and cancer: a systematic review. *Rev. Environ. Health.* 36, 467–476.
- Verla, A.W., Enyoh, C.E., Verla, E.N., Nwornorh, K.O., 2019. Microplastic-toxic chemical interaction: a review study on quantified levels, mechanism and implication. *SN. Appl. Sci.* 1, 1–30.
- Villanger, G.D., Lydersen, C., Kovacs, K.M., Lie, E., Skaare, J.U., Jenssen, B.M., 2011. Disruptive effects of persistent organohalogen contaminants on thyroid function in white whales (*Delphinapterus leucas*) from Svalbard. *Sci. Total. Environ.* 409, 2511–2524.
- Viršek, M.K., Lovšin, M.N., Koren, Š., Kržan, A., Peterlin, M., 2017. Microplastics as a vector for the transport of the bacterial fish pathogen species *Aeromonas salmonicida*. *Mar. Pollut. Bull.* 125, 301–309.
- Vroom, R.J., Koelmans, A.A., Besseling, E., Halsband, C., 2017. Aging of microplastics promotes their ingestion by marine zooplankton. *Environ. Pollut.* 231, 987–996.
- Wang, T., Wang, L., Chen, Q., Kalogerakis, J., Ji, R., Ma, Y., 2020. Interactions between microplastics and organic pollutants: Effects on toxicity, bioaccumulation, degradation, and transport. *Sci. Total. Environ.* 748, 142427.
- Wang, Y., Lv, L., Xu, C., Wang, D., Yang, G., Wang, X., Weng, H., Wang, Q., 2021. Mixture toxicity of thiophanate-methyl and fenvalerate to embryonic zebrafish (*Danio rerio*) and its underlying mechanism. *Sci. Total. Environ.* 756, 143754.
- Wise Jr, J.P., Wise, J., Wise, C.F., Wise, S.S., Zhu, C., Browning, C.L., Zheng, T., Perkins, C., Gianios Jr, C., Xie, H., Wise Sr., J.P., 2019. Metal levels in whales from the Gulf of Maine: A one environmental health approach. *Chemosphere* 216, 653–660.
- Xiong, X., Chen, X., Zhang, K., Mei, Z., Hao, Y., Zheng, J., Wu, C., Wang, K., Ruan, Y., Lam, P.K., Wang, D., 2018. Microplastics in the intestinal tracts of East Asian finless porpoises (*Neophocaena asiaeorientalis sunameri*) from Yellow Sea and Bohai Sea of China. *Mar. Pollut. Bull.* 136, 55–60.
- Xiong, X., Qian, Z., Mei, Z., Wu, J., Hao, Y., Wang, K., Wu, C., Wang, D., 2019. Trace elements accumulation in the Yangtze finless porpoise (*Neophocaena asiaeorientalis asiaeorientalis*)—a threat to the endangered freshwater cetacean. *Sci. Total. Environ.* 686, 797–804.
- Yang, Y., Liu, G., Song, W., Ye, C., Lin, H., Li, Z., Liu, W., 2019. Plastics in the marine environment are reservoirs for antibiotic and metal resistance genes. *Environ. Int.* 123, 79–86.
- Yilmaz, B., Terekci, H., Sandal, S., Kelestimur, F., 2020. Endocrine disrupting chemicals: exposure, effects on human health, mechanism of action, models for testing and strategies for prevention. *Rev. Endocr. Metab. Disord.* 21, 127–147.
- Ylitalo, G.M., Stein, J.E., Hom, T., Johnson, L.L., Tilbury, K.L., Hall, A.J., Rowles, T., Greig, D., Lowenstine, L.J., Gulland, F.M., 2005. The role of organochlorines in cancer-associated mortality in California sea lions (*Zalophus californianus*). *Mar. Pollut. Bull.* 50, 30–39.
- Zanuttini, C., Gally, F., Scholl, G., Thomé, J.P., Eppe, G., Das, K., 2019. High pollutant exposure level of the largest European community of bottlenose dolphins in the English Channel. *Sci. Rep.* 9, 12521.
- Zhang, Y., Lu, J., Wu, J., Wang, J., Luo, Y., 2020. Potential risks of microplastics combined with superbugs: Enrichment of antibiotic resistant bacteria on the surface of microplastics in mariculture system. *Ecotoxicol. Environ. Saf.* 187, 109852.
- Zhu, J., Yu, X., Zhang, Q., Li, Y., Tan, S., Li, D., Yang, Z., Wang, J., 2019. Cetaceans and microplastics: First report of microplastic ingestion by a coastal delphinid, *Sousa chinensis*. *Sci. Total. Environ.* 659, 649–654.