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Geographical and taxonomic patterns in aerobic traits of marine ectotherms

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The metabolism and hypoxia tolerance of marine ectotherms play key roles in limiting species geographical ranges, but underlying traits have only been directly measured for a small fraction of biodiversity. Here we diagnose and analyse spatial and phylogenetic patterns in hypoxia tolerance and its temperature sensitivity at ecologically active metabolic rates, by combining a model of organismal oxygen (O₂) balance with global climate and biogeographic data for approximately 25 000 animal species from 13 phyla. Large-scale spatial trait patterns reveal that active hypoxia tolerance is greater and less temperature sensitive among tropical species compared to polar ones, consistent with sparse experimental data. Species energetic demands for activity vary less with temperature than resting costs, an inference confirmed by available rate measurements. Across the tree of life, closely related species share similar hypoxia traits, indicating that evolutionary history shapes physiological tolerances to O₂ and temperature. Trait frequencies are highly conserved across phyla, suggesting the breadth of global aerobic conditions selects for convergent trait diversity. Our results support aerobic limitation as a constraint on marine habitat distributions and their responses to climate change and highlight the under-sampling of aerobic traits among species living in the ocean's tropical and polar oxythermal extremes.

This article is part of the theme issue 'The evolutionary significance of variation in metabolic rates'.

1. Introduction

The distribution of species across space and time is strongly influenced by physiological sensitivities to environmental conditions. In the ocean, oxygen (O₂) is increasingly recognized as a constraint on thermally and metabolically available habitat [1–4], even outside of O₂ minimum zones, and in waters with O₂ well above the traditional definition of hypoxic (less than 60 μM) [5,6]. Species sensitivity to low O₂ levels varies with temperature (T) and body size (B), owing to the thermal and allometric dependence of metabolism and physiological O₂ supply [7–9]. Limitation of growth by O₂ remains a leading hypothesis for the Temperature Size Rule [10–14].

Species hypoxia tolerance traits, such as the ratio of organismal O₂ supply and metabolic demand rates and their dependence on temperature, body size, and activity level can be used to map available aerobic habitat in the ocean [4,9,15–19]. Aerobic habitat barriers predicted from these traits and observed climate fields commonly align with the geographical range boundaries of species across ocean basins, latitudes and depths. Measured traits can thus help predict how the lowest O₂ level inhabited by species varies as a function of ocean temperature. They also mediate critical responses to climate change, including present and historical habitat exclusion [16,18,20], intra-specific body size variations [14], and marine mass extinction [21]. These traits are also expected to be strong determinants of future extinction risk from anthropogenic climate warming under ongoing O₂ loss

[22,23]. Quantifying how they vary across the ocean and the tree of life is key for a mechanistic understanding of biogeographic, taxonomic and evolutionary responses to climate.

While the traits governing temperature-dependent hypoxia tolerance have been estimated for dozens of marine species from direct respirometry experiments [9], they still represent a small fraction of known marine species [24], with the potential for regional, phylogenetic, and experimental biases. Hypoxia traits estimated from biogeographic observations reflect those directly measured in laboratory experiments, modified by ecological factors, including differences between resting versus active metabolism. Here we pair a model of organismal O₂ balance with global biogeographic data and climatological measurements of temperature and O₂ to diagnose temperature-dependent hypoxia traits from a database of 24 852 marine animal species (figure 1a), and analyse trait correlations, spatial patterns across the ocean and phylogenetic signals across the tree of life.

2. Methods

The minimum partial pressure of oxygen (pO₂) required to maintain metabolism is a measure of physiological vulnerability to hypoxia (figure 1b,c). A species minimum pO₂ threshold can vary with temperature and increases with activity level relative to the resting state, where it is defined as V_h (in atm) at an arbitrary reference temperature (see the electronic supplementary material, table S1 for definitions of mathematical symbols). The ecological activity of a viable population in nature is vulnerable to O₂ limitation at a higher pO₂ level, V_h*Φ_{crit}, where Φ_{crit} is the ratio of sustained to resting metabolic rate and falls between 1 (i.e. no active costs) and the ratio of maximum to resting metabolic rate, defined by the factorial aerobic scope (FAS) and commonly measured in animal physiology [9,27,28]. This active hypoxia vulnerability of a species is inversely related to its active hypoxia tolerance, which we denote A_{eco} (1/atm), such that A_{eco} = 1/(V_h*Φ_{crit}), following analogous definitions for resting hypoxia tolerance and vulnerability at minimum metabolic rate, i.e. A_o = 1/V_h [9] (figure 1d). Thus, for an ecologically active population, the minimum pO₂ in a metabolically viable environment (pO₂^{act}) can be described by:

$$pO_2^{\text{act}} = \frac{1}{A_{\text{eco}}} f(E_{\text{eco}}, T), \quad (2.1)$$

where $f(E_{\text{eco}}, T)$ is the variation in active hypoxia tolerance as a function of temperature and the species-specific thermal sensitivity, E_{eco} . Variations in hypoxia tolerance owing to body size are relatively small [14,29], and we therefore neglect the allometric term here for simplicity (but see the electronic supplementary material).

Hypoxia tolerance is commonly observed to vary nonlinearly with temperature (figure 1d) and recent experimental data indicate it can even be non-monotonic, reaching a maximum tolerance (minimum vulnerability) at a temperature optimum [16,25,30]. The functional form of $f(E_{\text{eco}}, T)$ must thus accommodate a wide range of temperature dependencies. The exponential Arrhenius function of temperature provides a natural starting point, because it describes many chemical and biological rates, including metabolism (O₂ demand) and diffusion (O₂ supply), as well as external ventilation and internal circulation over a limited temperature range:

$$f(E, T) = \exp\left(\frac{-E}{k_B} \left[\frac{1}{T} - \frac{1}{T_{\text{ref}}}\right]\right), \quad (2.2)$$

where E is an activation energy (in electron volts, eV) and represents the thermal sensitivity of the rate of any process, or the

ratio of such rates. The function, f , increases with temperature when $E > 0$ and decreases with temperature when $E < 0$. For resting hypoxia tolerance, which is defined by the ratio of O₂ supply to resting metabolic demand (electronic supplementary material) [4,9], its temperature sensitivity, E_o , is the difference between the effective activation energies for metabolic demand (E_d) and O₂ supply, E_s ($E_o = E_d - E_s$) [9]. This formulation captures much of the thermal variation in resting hypoxia tolerance from laboratory measurements.

To represent active hypoxia tolerance across the full range of ocean temperatures, the temperature sensitivity of the Arrhenius equation requires two distinct modifications. First, metabolic rates needed to fuel ecological activity will elevate hypoxia vulnerability by the ratio of sustained to resting metabolic rates, Φ_{crit}, which in principle may also vary with temperature. We assume that this ratio also follows an Arrhenius function of temperature, with a distinct temperature sensitivity, denoted $E_{\Phi_{\text{crit}}}$, a species trait that may differ from that of resting metabolism (E_d). Including this additional component of ecologically critical energy demand, the net temperature dependence of active hypoxia tolerance, which we denote E_{eco} , becomes a sum $E_{\text{eco}} = E_o + E_{\Phi_{\text{crit}}} = E_d - E_s + E_{\Phi_{\text{crit}}}$. To the degree that the ratio of energetic expenditure on activity versus maintenance is constant across a species temperature range, $E_{\Phi_{\text{crit}}}$ will equal 0 and the E_{eco} diagnosed from biogeographic data should be equal to that of resting hypoxia tolerance derived from laboratory experiments (figure 1b,c). By contrast, if the energy spent on ecological activity increases less with temperature than resting costs, the ratio of active to resting energy will decline ($E_{\Phi_{\text{crit}}} < 0$), and E_{eco} should be lower than its physiological counterpart, E_o . Thus, the comparison of biogeographically estimated E_{eco} with laboratory derived E_o can reveal new information about how temperature influences species allocation of energy to activity relative to maintenance needs.

A second modification of the simple Arrhenius function to describe hypoxia tolerance arises from the multiple steps in the O₂ supply chain, which can have distinct activation energies (E_s values). This leads to an effective E_o that itself can vary across a species temperature range (dashed blue line 2, figure 1d) [25]. The complexity of these physiological processes can be represented in a dynamic organismal model of metabolism and O₂ transport that reproduces observed thermal optima in hypoxia tolerance [25]. The diversity of resulting hypoxia tolerance curves can be simply parameterized by allowing E_o to be a linear function of T , with slope $(\frac{dE_o}{dT})$ (electronic supplementary material, equation S4), which captures the multistep nature of O₂ supply with minimal additional parameters (dotted blue line 1, figure 1d).

We estimated the ecophysiological traits (A_{eco} and E_{eco} , and $\frac{dE_o}{dT}$) that define the minimum pO₂ to sustain active aerobic metabolism by combining a model of organismal O₂ supply to demand rates, the metabolic index (Φ ; electronic supplementary material, equation S1, S8), with species geospatial occurrence data and long-term mean observed fields of ocean temperature and O₂ (electronic supplementary material) [31–33]. Geospatial occurrences ($n = 20\,441\,987$) were downloaded from the Ocean Biodiversity Information System (OBIS; <https://obis.org/>) for 25 090 marine ectothermic animal species (figure 1a; electronic supplementary material, table S2) and paired to gridded hydrographic data (1° × 1°) from monthly climatological fields from the World Ocean Atlas. Traits were diagnosed by projecting species geospatial occurrences onto the temperature and partial pressure of oxygen (T-pO₂) state-space that they inhabit, using a standard statistical categorization metric, the F1-score (electronic supplementary material, equation S10), to maximize the skill of Φ in predicting the observed lower pO₂ threshold (pO₂^{act}; electronic supplementary material, equation S9) above which occurrences are common and below which they are rare, as a function of ocean temperature (electronic supplementary material, figure S1) [18]. In total, this procedure yields trait estimates for

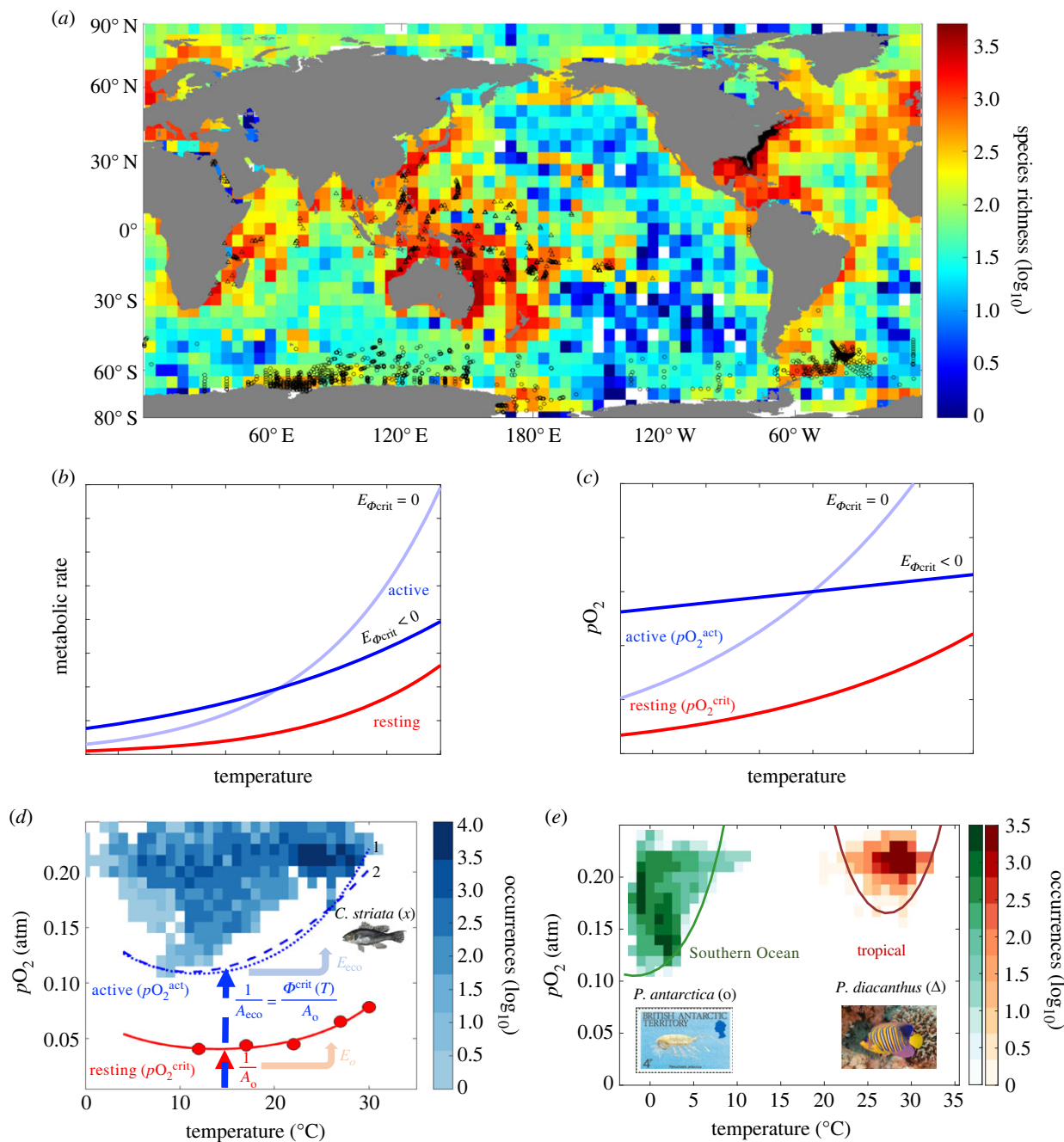


Figure 1. Temperature-dependent hypoxia traits estimated from biogeographic observations. Traits are diagnosed from paired geospatial occurrences, and temperature and partial pressure of oxygen (T-pO₂) observations for globally diverse species (a; species richness). (b,c) Active and resting metabolic rates increase with temperature (b), giving rise to temperature-dependent lower pO₂ thresholds required to maintain metabolism, pO₂^{act} and pO₂^{crit}, respectively (c). If the ratio of active to resting metabolic rate is constant versus temperature ($E_{\phi_{crit}} = 0$), pO₂^{act} and pO₂^{crit} vary similarly with temperature (compare light blue and red lines). By contrast, if activity is less temperature sensitive than resting costs ($E_{\phi_{crit}} < 0$), pO₂^{act} varies less with temperature than pO₂^{crit} (compare dark blue and red lines). (d,e) Species occurrences in T-pO₂ state-space (shading) reveal how the minimum pO₂ inhabited by a species in the ocean (pO₂^{act}) varies with T. This active T-dependence (E_{eco}) reflects both the resting hypoxia tolerance (E_o) measured by pO₂^{crit} (red points in d) and the ratio of active to resting metabolic costs ($E_{\phi_{crit}}$). Active hypoxia tolerances (A_{eco}) at the reference T (15°C) are reduced relative to the resting state (A_o) by the ratio of active to resting metabolic rates (Φ_{crit}). T-dependences can vary across a species temperature range owing to the multi-step nature of O₂ supply, as captured by including multiple Arrhenius functions (dashed line 2 in d) [25], or by allowing E_o and E_{eco} to be a linear function of T, with slope ($\frac{dE}{dT}$) (dotted line 1 in d). T-pO₂ occurrences (d,e) and geographical distributions (symbols in a) are shown for: (d) western Atlantic black sea bass (*Centropomus striata*, x) [26], (e) the Southern Ocean copepod (*Paraeuchaeta antarctica*, o), and tropical Indo-Pacific royal angelfish (*Pygoplites diacanthus*, Δ). Traits are estimated by fitting a model of O₂ supply to demand (electronic supplementary material, equation S1, S8; lines) to experimental pO₂^{act} data and inhabited lower pO₂ levels. Illustrations in (d) and (e) are from NOAA FishWatch (https://en.wikipedia.org/wiki/Black_sea_bass#/media/File:Centropomus_striata.png) and WoRMS (<https://www.marinespecies.org/aphia.php?p=image&pic=46884&tid=344974>), respectively, and photograph in (e) is from D. Delso ([https://en.wikipedia.org/wiki/Royal_angelfish#/media/File:Pez_%C3%A1ngel_real_\(Pygoplites_diacanthus\),_parque_nacional_Ras_Muhammad,_Egipto,_2022-03-26,_DD_155.jpg](https://en.wikipedia.org/wiki/Royal_angelfish#/media/File:Pez_%C3%A1ngel_real_(Pygoplites_diacanthus),_parque_nacional_Ras_Muhammad,_Egipto,_2022-03-26,_DD_155.jpg)).

24 852 species from 13 marine animal phyla and 51 classes with global coverage, including cosmopolitan species, and those endemic to the Northern and Southern Hemisphere's high and mid-latitude waters, and the tropics (figure 1a; electronic supplementary material, figure S2).

3. Results

Across species, active hypoxia tolerance and its temperature sensitivity exhibit large-scale geographical patterns (figure 2). Active hypoxia tolerances increase systematically from the

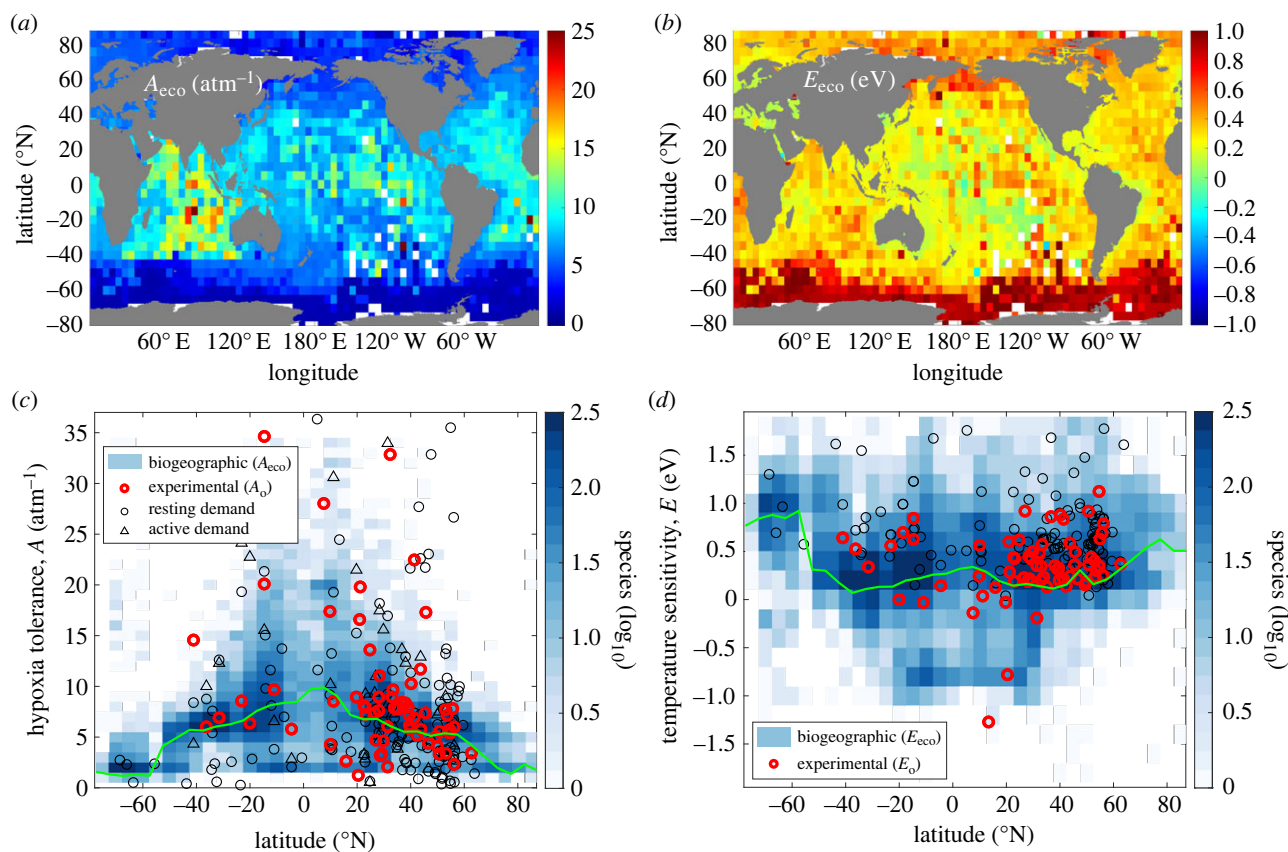


Figure 2. Geographical variation of active hypoxia tolerance and its temperature sensitivity and underlying metabolic traits. (a,b) Maps of interspecies median active hypoxia tolerance (A_{eco}) and its temperature sensitivity (E_{eco}) reveal large-scale gradients in traits, including a polar to tropical increase in A_{eco} and decrease in E_{eco} . (c,d) Diagnosed traits (colours; \log_{10} number of species) vary with latitude similar to available resting traits from experimental data (red points) and can be understood as arising from spatial variations in resting and active metabolic O_2 demand (black circles and triangles, respectively in c) and their temperature dependences (black circles in d) show resting E_{d}), with the potential for contributions from organismal O_2 supply. In (c), metabolic contributions to A_{eco} are predicted from the ratio O_2 supply to demand rates ($A_{\text{eco}} = \alpha_{\text{S}}/(\alpha_{\text{D}}*\Phi_{\text{crit}})$), using latitudinal variations in either measured resting metabolism (α_{D}) (circles) or active metabolism ($\alpha_{\text{D}}*\Phi_{\text{crit}}$) (triangles) while holding other factors, like the efficacy of O_2 supply (α_{S}), at median experimental values (Φ_{crit} data from [9]). Median diagnosed trait values versus latitude are shown as solid green lines. In (c,d), species traits are plotted at their corresponding median latitude. In (c), resting hypoxia tolerances, A_{o} (red), are normalized by a constant Φ_{crit} of 3.

poles to equator, with lowest values throughout the Southern Ocean and a broad maximum across the low latitudes, reflecting the need for species to support active aerobic metabolism at warmer water temperatures and lower ambient pO_2 in the tropics (figure 2a,c). By contrast, the temperature sensitivity of active hypoxia tolerance generally decreases from the poles to (sub)tropics with a weaker secondary peak at the equator, corresponding to the equatorial Pacific upwelling zone (figure 2b,d). This latitudinal decline in E_{eco} indicates the common pattern of a decrease in hypoxia tolerance from warming becomes weakened. Peak E_{eco} values are found in Southern Ocean species, and to a lesser extent those from the north Pacific and Arctic (figure 2b,d), as this-trait provides habitat in cold waters (e.g. *Paraeuchaeta antarctica* in figure 1a,e). Species exhibiting negative E_{eco} which indicates that hypoxia tolerance instead increases with temperature, are most frequently found in the tropics and subtropics (e.g. *Pygoplites diacanthus* in figure 1a,e), but even in these regions, median species E_{eco} remains positive. Diagnosed differences in the distributions of A_{eco} and E_{eco} between the low (less than 30°) and high latitudes (greater than 60°) are statistically significant (two-sample Kolmogorov-Smirnov test, $p < 10^{-3}$) (electronic supplementary material, figure S3). Measured resting hypoxia tolerance (A_{o}) is also higher in the tropics where negative E_{o} values are found

(figure 2c,d), indicating that the geographical structure of resting traits from laboratory respirometry experiments is consistent with patterns in active traits.

Variability in species active hypoxia tolerance also increases from the poles to equator, corresponding to the stronger variance of T and O_2 associated with their vertical gradients (figure 2c). While the central tendency and upper deciles of A_{eco} are higher in the tropics, these regions also host many species with low hypoxia tolerance. Some species endemic to the tropics or high latitudes can appear to require pO_2 above atmospheric levels (i.e. $A_{\text{eco}} < 4.8 \text{ atm}^{-1}$), because hypoxia tolerance is defined at a common reference temperature that lies outside their native range (figure 1e). Low hypoxia tolerance can also reflect species living in supersaturated O_2 conditions found in surface waters or in the abyssal sea, where pO_2 can exceed that of the surface atmosphere (approx. 0.21 atm).

Spatial variations in active hypoxia tolerance and its temperature sensitivity can be understood as arising from underlying metabolic O_2 demand and organismal supply traits (figure 2c,d). Active hypoxia tolerance is equivalent to the ratio of organismal O_2 supply efficacy (α_{S}) to resting metabolic O_2 demand rate (α_{D}), reduced by activity level, Φ_{crit} : $A_{\text{eco}} = \alpha_{\text{S}}/(\alpha_{\text{D}}*\Phi_{\text{crit}})$ (electronic supplementary material) [9]. On their own, observed variations in species resting and

active metabolic rates yield a latitudinal pattern of A_{eco} similar to diagnosed and measured A_{eco} , with central values in hypoxia tolerance from metabolic rate alone increasing from high to low latitudes (black circles and triangles in figure 2c). This is because species temperature-normalized resting and active metabolic rates are observed to increase significantly, albeit weakly, with absolute latitude ($R^2 = 0.09$ and 0.15 , respectively, $p < 0.01$), such that at the same temperature, tropical species with slower metabolisms would have a lower demand for O_2 than more metabolically active, high-latitude ones (electronic supplementary material, figure S4). Gradients in organismal O_2 supply rates versus latitude might also contribute to patterns of A_{eco} , but there are fewer observations of this trait and latitude trends are weak in available data. The temperature sensitivity of resting metabolic rate (E_d ; black circles in figure 2d) and O_2 supply (E_s) also show little apparent trends with latitude but estimates of these parameters are largely missing from the Southern Ocean where the latitude gradient is expected to be strongest based on diagnosed E_{eco} .

On their own, neither T nor pO_2 show much skill in explaining the inhabited T - pO_2 state-space of individual species (figure 1d,e; electronic supplementary material, figure S5). Otherwise, mean species temperature sensitivities (E_{eco}) would centre on 0 (no temperature dependence) or at extreme values ($|E_o| > 2$), approximating pO_2 -independent temperature thresholds (figure 2d). Moreover, spatial variations in diagnosed species A_{eco} and E_{eco} do not reflect the relationship of environmental pO_2 versus T in time and depth from local ocean depth profiles, as would be expected if inferred traits were an artifact of the ocean's vertical stratification or of poorly sampled ranges (electronic supplementary material, figure S6).

The joint distribution of diagnosed E_{eco} and A_{eco} reveals new structure in the relationship between active hypoxia tolerance and its temperature sensitivity (figure 3a). At low active hypoxia tolerance, inter-species variations in E_{eco} increase and display two prominent tails, largely corresponding to tropical ($E_{\text{eco}} < 0$), and polar species ($E_{\text{eco}} > 0$). At high hypoxia tolerance, most commonly found in tropical to mid-latitudes species, inter-species variations in E_{eco} largely converge towards increasingly positive values with larger A_{eco} . The presence of these distinct tails of the joint distribution of active traits is not readily evident in the relatively sparse experimental data of resting traits alone but is supported by it.

If temperature-dependent O_2 thresholds constrain the geographical ranges of marine species and thereby influence population fitness, we might expect species trait frequencies to reflect global aerobic habitat availability. We tested the simple hypothesis that traits providing more voluminous aerobic habitat are found among a larger number of species by computing the global aerobic habitat volumes (m^3) for a given combination of E_{eco} and A_{eco} using the long-term observations of O_2 and T (figure 3b,c). In the warm, shallow ocean (0–100 m), there is a positive association between the E_{eco} and A_{eco} with available habitat. Habitat volumes are largely afforded to species with negative E_{eco} values and low A_{eco} and those with positive E_{eco} and high A_{eco} . By contrast, in colder, deeper waters, the relationship between E_{eco} and A_{eco} with available habitat changes sign—as E_{eco} increases, the A_{eco} with available habitat declines. Across depth, this change in slope in the relationship between E_{eco} versus A_{eco} values with available habitat is consistent with the shape of

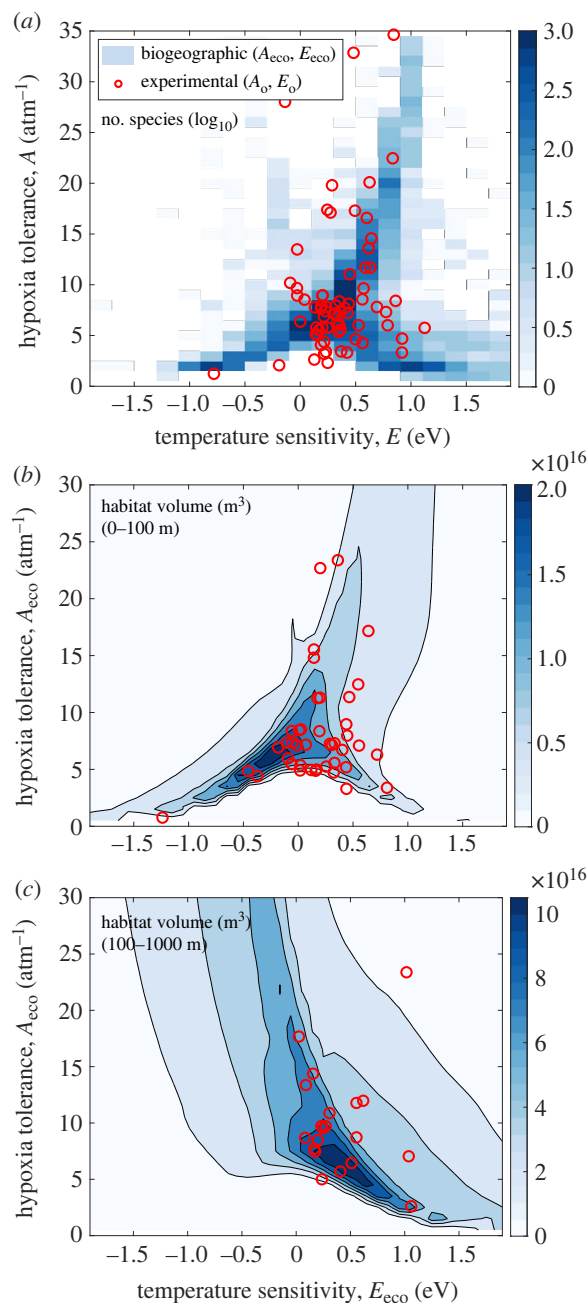


Figure 3. Joint frequency distributions of hypoxia tolerance and its temperature sensitivity compared to habitable ocean volume. (a) The joint frequency distributions of active and resting hypoxia tolerance versus their temperature sensitivities are consistent with those expected if trait frequency is influenced by aerobic habitat availability (b,c). (a) The number of species with a given trait combination of active A_{eco} and E_{eco} estimated from biogeographic data (colours; \log_{10}) is compared to resting traits A_o and E_o derived from laboratory experiments (red points). A_o is normalized by a constant Φ_{crit} of 3. There is a positive relationship between E_{eco} and A_{eco} values with aerobic habitat available in the relatively warm upper 100 m (b) but this relationship changes sign in cooler deeper waters (100–1000 m; c). Available habitat is defined as the water volume (m^3) with $1 \leq \Phi \leq \Phi_{\text{max}}$, where Φ_{max} is the median upper Φ value inhabited by marine species (figure 4b) and assumes a dE/dT of $0.025 \text{ eV}/^\circ\text{C}$ [9]. E_{eco} is plotted at species median inhabited temperatures (a) and at T_{ref} (b,c). In (b,c), laboratory A_o is normalized by the species-specific Φ_{crit} (data from [9]) and laboratory E_o is shifted by the median offset from biogeographic E_{eco} (red points).

the diagnosed E_{eco} versus A_{eco} frequency distribution (figure 3a), and with estimates of active traits based on laboratory data (red points in figure 3b,c), suggesting global

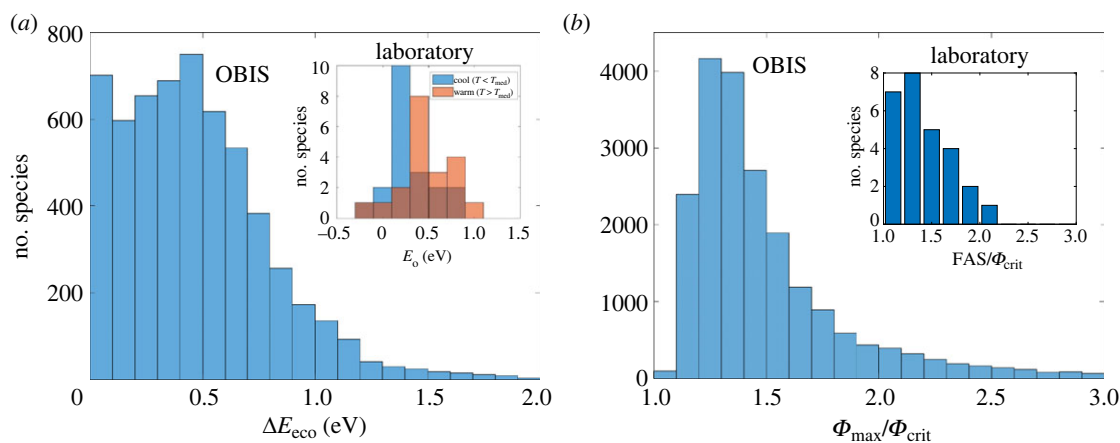


Figure 4. Measures of ‘cold edge’ habitat limits shaped by Φ . (a) Frequency distribution of the change in species E_{eco} (ΔE_{eco}) from cold to warm habitat limits. Across species, differences in the temperature sensitivity of active hypoxia tolerance (E_{eco}) over the inhabited T range are qualitatively consistent with differences observed from direct laboratory data of resting hypoxia tolerance (E_o) above and below the median experimental temperature (T_{med}) (inset in (a); data from [14]) and predicted in a mechanistic organismal model that includes multi-step O_2 supply [25]. The effective T sensitivity of active hypoxia tolerance at the warm (T_w) and cold (T_c) range edges is estimated as $E_{\text{eco}w/c} = E_{\text{eco}}(T_{\text{ref}}) + \frac{dE}{dT}(T_{w/c} - T_{\text{ref}})$, where T_w is the 95th and T_c is the 5th percentile of inhabited temperatures, and $E_{\text{eco}}(T_{\text{ref}})$ and $\frac{dE}{dT}$ are species-specific traits. (b) Frequency distribution of species maximum occupied Φ values (Φ_{max}) relative to Φ_{crit} (95th percentile). The median species inhabits waters with up to 1.4 times as much Φ as needed for sustained ecological activity. The ratio of maximum metabolic rate to resting metabolic rate ($\text{FAS} = \text{MMR} : \text{RMR}$) measured in laboratory experiments is also a factor of $\approx 40\%$ higher than the Φ_{crit} required to sustain a species population in the environment (inset in (b); data from [9]).

aerobic habitat volume acts as a selective pressure on the global frequency of species traits.

The inferred trait distributions allow a more comprehensive examination of the role of Φ at the cold edge of species geographical ranges. For some species, the lower limit of inhabited pO_2 increases at cooler temperatures, indicating a shift in the thermal sensitivity of O_2 supply and thus the potential for hypoxia tolerance to restrict habitat even at cold edges (figure 1*d,e*). We computed the effective T sensitivity of active hypoxia tolerance at warm and cold range edges (figure 4*a*). We find a mean change in effective E_{eco} (ΔE_{eco}) across species temperature ranges of approximately 0.4 eV, from just below 0 in cold water to approximately 0.4 eV at the warm edge, which is about twice the mean difference in resting E_o in cold versus warm water from laboratory measurements ($\Delta E_o = 0.2$ eV; inset figure 4*a*) [14], and is owing to the larger $\frac{dE}{dT}$ in diagnosed traits (mean 0.05 ± 0.03 s.d., eV/°C) compared to laboratory data ($\frac{dE}{dT} \approx 0.022$, 0.013–0.036 range, eV/°C) [9]. A change in E_{eco} across temperature is qualitatively consistent with the switch from O_2 supply limitation via diffusion ($E_s \approx 0.05$ eV) in warm water to ventilation and/or internal circulation ($E_s \approx 0.4$) in cold water [25].

Even for species without a thermal optimum in hypoxia tolerance, Φ has the potential to influence habitat at species cold and/or high O_2 edges via physiological and ecological mechanisms apart from direct aerobic limitation of metabolic demand. We diagnosed the ratio of highest to lowest occupied Φ for each species ($\Phi_{\text{max}}/\Phi_{\text{crit}}$), which displays a well-defined frequency distribution with a median of approximately 1.4 (1.2 to 2.2, inter-decile range; figure 4*b*), indicating that the typical species inhabits waters with up to 40% higher O_2 supply than is required for population sustenance. The ratio of maximum metabolic rate to resting metabolic rate (MMR : RMR) measured in laboratory experiments is also a factor of approximately 1.4 higher than the Φ_{crit} required to sustain a species population in the environment (inset figure 4*b*) [9,34].

Diagnosed global trait distributions also allow for evolutionary analyses of trait variation across taxa. Taxonomic

signals in A_{eco} and E_{eco} might be expected because related species share similar morphological structures and physiological constraints on metabolism and O_2 supply, factors that govern hypoxia tolerance [9] and which could be influenced by evolutionary history if species in different groups evolved under distinct climate conditions [35].

We mapped taxonomic variations in A_{eco} and E_{eco} across the phylogenetic tree of marine animals using the Open Tree of Life (<https://tree.opentreeoflife.org>) following methods of [36] (figure 5). We find that both hypoxia tolerance and its temperature sensitivity have a significant phylogenetic signal (Pagel’s lambda, $\lambda = 0.74$ and 0.53 , respectively and $p < 10^{-3}$), consistent with results for directly measured resting hypoxia tolerance in fishes. This can be observed on the phylogenetic tree, where shared lineages often show similar values of A_{eco} and E_{eco} . At higher taxonomic ranks towards the tree root, differences between lineages are reduced. Across phyla, distributions of E_{eco} and A_{eco} show convergence (figure 5*b,d*), indicating that most variation in traits occurs within this higher taxonomic rank, consistent with distributions of resting hypoxia traits [9]. Instead, if taxonomic trait variation primarily occurred at the phylum level, geographical niche differentiation across phyla would be expected. For example, phyla with high active hypoxia tolerances that increase with T would be restricted to warm, low O_2 conditions while those with lowest tolerances and high E_{eco} would only be found in cold waters, in contrast to observed global biodiversity patterns of individual phyla, which have widespread distributions [39]. These observations suggest convergence of species within cosmopolitan taxa, like phyla, on a similar variance in traits across species to be able to inhabit the full breadth of oxythermal conditions present in the ocean. Convergence in diagnosed traits might also arise through ecological interactions which have potential to shape the realized niches of species not directly limited by temperature-dependent hypoxia, but which are tied to species that are so constrained.

The temperature-dependences of active and resting hypoxia tolerance estimated from biogeographic and

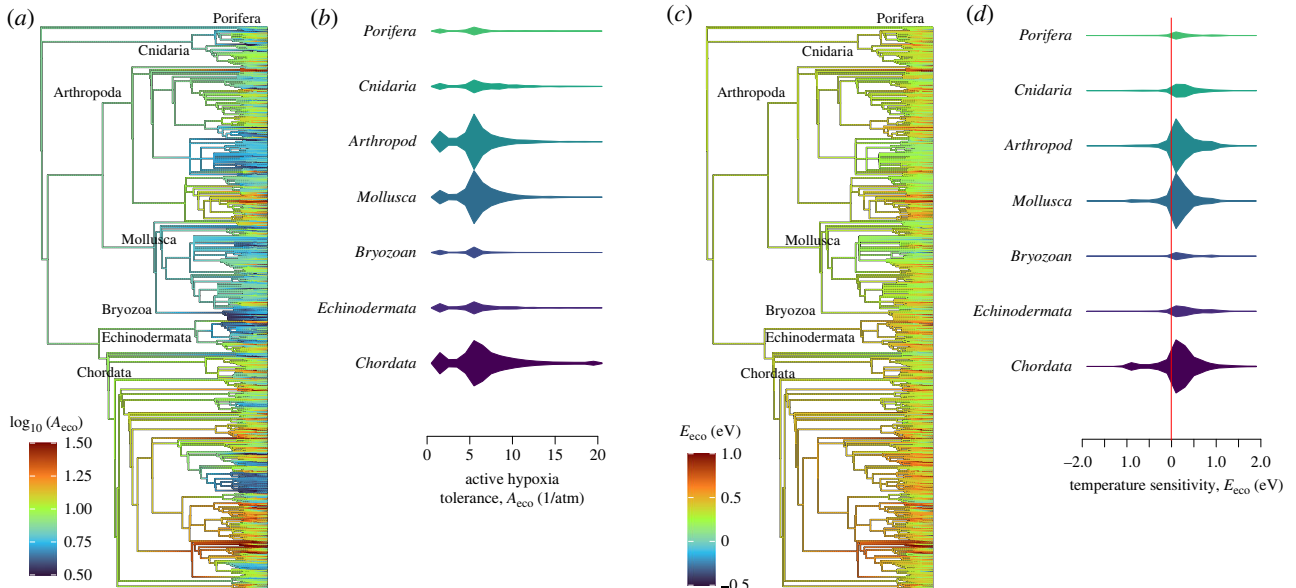


Figure 5. Phylogenetic variation of active temperature-dependent hypoxia traits. Closely related species on the phylogenetic tree can show similar (a) active hypoxia tolerances, A_{eco} , and temperature sensitivities (c), E_{eco} (colours). Phylogenetic relationships are constructed using the TreeofLife database [37]. In (a) and (c), tree branch lengths are set equal to the number of descendant tips minus one [38] and use a Grafen's rho parameter, which influences the ratio of branch lengths at tree roots versus tips, equal to 0.4, found to best explain observed variations in resting hypoxia tolerance in fishes [36]. Inter-taxa variations in traits decrease at higher ranks, such as phyla (labelled). (b,d) Intra-phyla distributions of E_{eco} and A_{eco} show convergence across phyla. Trees in (a,c) show a sample (1997 species with $F1 > 0.8$) of the full intra-phyla distributions plotted in kite diagrams (b,d), which are limited to phyla with trait estimates available for more than 500 species.

laboratory data can be paired to derive a new trait, the temperature dependence ($E_{\Phi_{\text{crit}}}$) of the ratio of active to resting metabolic rate (Φ_{crit}). The value of Φ_{crit} has previously been assumed to be independent of temperature [4,9], such that $E_{\Phi_{\text{crit}}} = 0$. Across species, the median E_{eco} (0.19 eV) falls below resting E_o (0.34 eV) (electronic supplementary material, figure S7), suggesting that the ratio of active to resting metabolic rates declines with warming (i.e. $E_{\Phi_{\text{crit}}} < 0$) since $E_{\text{eco}} = E_o + E_{\Phi_{\text{crit}}}$ (figure 1b,c). We estimated the distribution of $E_{\Phi_{\text{crit}}}$ at the species level and found that its distribution is indeed shifted below E_o and has a median value of -0.14 eV (figure 6a). Since resting metabolic rates increase with temperature, this implies that for the typical species, active metabolic costs are relatively constant.

We tested this prediction with laboratory measurements of MMR and RMR at multiple temperatures for six species using the definition for FAS ($\text{FAS} = \text{MMR}/\text{RMR}$), which measures the ratio of MMR to RMR, as a proxy for Φ_{crit} . This comparison is necessarily approximate because sustained activity in nature falls between MMR and RMR, and includes biological processes not resolved in short term MMR experiments, such as reproduction, feeding, growth and predation, in addition to uncertainty in laboratory measurements. Nevertheless, it serves as an initial check on the expected lower temperature-dependence of active hypoxia tolerance (E_{eco}) compared to the resting state (E_o). We find that experimental $E_{\Phi_{\text{crit}}}$ is consistently below 0 for all species (median $E_{\Phi_{\text{crit}}} = -0.25$ eV) (figure 6b), shifting the predicted estimates of E_{eco} for these species below their measured E_o . This finding implies that the ratio of active to resting metabolic rate declines with warming as predicted. Moreover, $E_{\Phi_{\text{crit}}}$ diagnosed from biogeographic data and estimated from laboratory FAS are correlated at the species level ($R^2 = 0.77$, $p = 0.02$), although the sample size is small. These findings are also in qualitative agreement with published inter-specific estimates of the temperature-dependence of MMR and resting metabolic rate in fishes [34]. Thus, experimental data on rates of active to resting

metabolic rate support the lower temperature-dependence of active compared to resting costs inferred here.

4. Discussion

Diagnosing species ecophysiological traits from biogeographic observations allows the active hypoxia tolerance and its temperature sensitivity to be estimated for tens of thousands of well-documented marine species in major animal phyla. Global spatial gradients in species traits appear to reflect adaptations needed to provide metabolically viable habitat under distinct oxythermal conditions, such as elevated active hypoxia tolerances that decline less with temperature in the warm, low O_2 tropics and highly positive E_{eco} that provides habitat in cold waters of the Southern Ocean.

Species range boundaries in T-p O_2 state-space are consistent with measured temperature-dependent hypoxia thresholds whose mean temperature dependence ($E_{\text{eco}} \approx 0.2 = E_d - E_s + E_{\Phi_{\text{crit}}}$) can be explained by the temperature sensitivity of organismal O_2 supply ($E_s \approx 0.4$), and metabolic demand ($E_d \approx 0.7$) [9,25], lowered by the ratio of active to resting metabolic rates ($E_{\Phi_{\text{crit}}} \approx -0.1$). In support of this mechanistic association, experimental data implies a lower temperature dependence for the cost of activity compared to maintenance, which imparts a decline in Φ_{crit} with warming. In addition, changes in diagnosed E_{eco} across the oceanic temperature range are consistent with direct laboratory measurements and predictions from an organismal model that includes the effect of changing O_2 supply mechanisms (figures 1d,e and 4). Together, these results support aerobic habitat limitation as a constraint on marine ectotherm species geographical distributions and their sensitivities to climate change.

The correspondence between $\Phi_{\text{max}}/\Phi_{\text{crit}}$ and maximum to sustained metabolic rates suggests that O_2 supply capacity

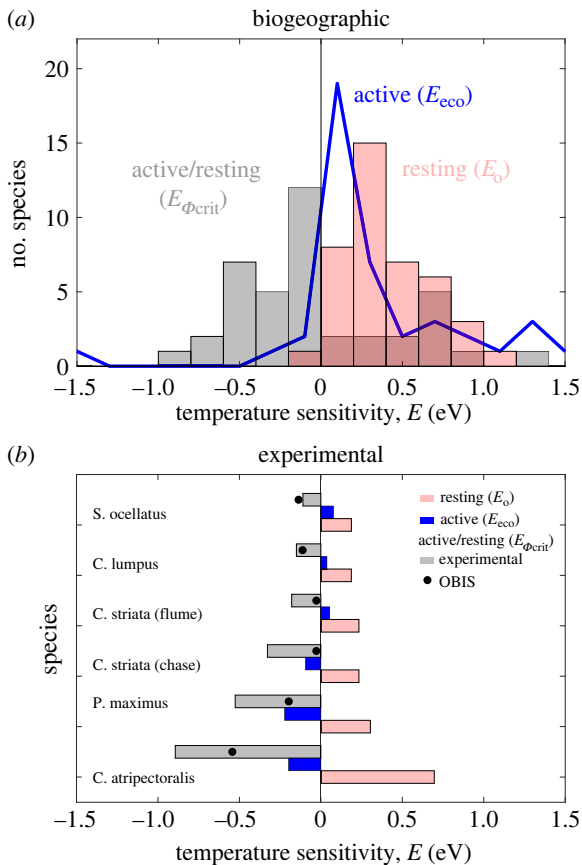


Figure 6. Temperature-sensitivity of active and resting hypoxia tolerance. (a) Active hypoxia tolerance varies less with temperature (E_{eco}) than in the resting state (E_0) because the ratio of active to resting energetic costs ($E_{\Phi_{\text{crit}}}$) declines with warming. $E_{\Phi_{\text{crit}}}$ distribution in (a) is diagnosed from the difference between OBIS (E_{eco}) and laboratory (E_0) at the species level. (b) Across species, experimental $E_{\Phi_{\text{crit}}}$ (grey) derived from ratios of maximum to resting metabolic rates at multiple temperatures (electronic supplementary material) also declines with warming, lowering predicted experimental E_{eco} compared measured E_0 . $E_{\Phi_{\text{crit}}}$ from experimental data (grey bars) and OBIS (points) are correlated at a species-level.

evolves to satisfy maximum metabolic demand [40]. Species occupying waters with O_2 supply above their maximum metabolic capacity for O_2 consumption might eventually be excluded via competition for limiting resources, including food, by other species with lower hypoxia tolerance that instead invest available energy elsewhere besides O_2 supply physiology (e.g. structures to increase gill surface area, ventilation or circulation rates). The large energetic costs of osmoregulation might also limit the benefits of investing in O_2 supply traits [41]. In addition, parasitic infection rates have been observed to increase with O_2 ventilation of the gills [42], one of the factors influencing hypoxia tolerance, suggesting species with lower A_{eco} (and thus Φ_{max}) might also have a lower infection risk.

Aerobic habitat constraints also appear to apply across diverse animal phyla, which have converged on a similar average and variance in species traits governing hypoxia tolerance and its temperature sensitivity despite widely varying morphology and physiology. This trait convergence points to a selective pressure to colonize the wide breadth of aerobic conditions found across the ocean. While we cannot rule out convergence in realized niches owing to species ecological interactions, as opposed to fundamental niches owing to ecophysiological traits, one would not expect to find

biogeographic niches consistent with temperature-dependent lower pO_2 curves unless some species were dependent on others, which were fundamentally constrained by the ecophysiological traits in the first place. Moreover, a similar convergence in inter-phyla trait distributions have been found for resting hypoxia tolerance and its temperature sensitivity from laboratory measurements, which include no effect from ecological interactions [9] and are a component of the active traits diagnosed here. The correspondence between diagnosed trait frequencies and global habitat availability suggests that traits providing larger habitats are selected for in a larger number of species. Open questions remain about the mechanistic factors shaping this selection, including how observed central tendencies, variability and trait coexistence is maintained across species, and how trait adaptation might have varied under different climate states in Earth history, with implications for past ecosystem sensitivities to climate change and taxonomic selectivity during marine extinctions [43].

Diagnosing species trait distributions provides a useful constraint on modern species habitat vulnerabilities to anthropogenic climate change. The spatial patterns of species traits diagnosed from biogeographic data support modelled trait gradients for the modern ocean and past time-intervals, which govern extinction patterns in future projections and palaeo simulations [21,23]. Diagnosed trait gradients thereby strengthen the hypothesis that extinction risk from climate warming increases with latitude, as species least hypoxia tolerant (lowest A_{eco}) and most temperature sensitive (highest E_{eco}) are found polewards, and have nowhere to seek refuge in a warmer, less oxygenated ocean. By contrast, species from lower latitudes that are more tolerant to hypoxia and with weaker temperature sensitivities might be able to track aerobic habitat niches to higher latitudes as these environments become more tropical, consistent with recently observed species poleward migrations under warming [44].

Diagnosed trait distributions also reveal the current limitations of direct experimental data compilations. Species found at the extremes of the ocean's oxythermal conditions are poorly sampled in laboratory data compared to their oceanic diversity, including for lowest and highest A_{eco} and E_{eco} . Trait estimates from species spanning Southern Ocean latitudes, where strong trait gradients are inferred and prior work indicates O_2 limitation [45], could provide a valuable test of these findings. The under-sampling of extreme traits in laboratory compilations is unlikely to impact global projections of species vulnerability to climate change [21], but could bias regional assessments. Polar oceans are projected to experience the strongest O_2 declines under anthropogenic climate change [46], while in the tropics, the frequency of metabolic storms, where hypoxic conditions co-occur with heat waves, is expected to become more severe [20]. Future efforts to understand climate impacts on aerobic habitat viability in the ocean should therefore prioritize species endemic to these biologically unique and climatically vulnerable ecosystems.

Ethics. This work did not require ethical approval from a human subject or animal welfare committee.

Data accessibility. Data, MATLAB and R code are available from the GitHub repository: https://github.com/jlpenn/MI_traits_obis [47].

Supplementary methods, figures, and tables are provided in the electronic supplementary material [48].

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. J.L.P.: conceptualization, data curation, formal analysis, investigation, methodology, validation, visualization, writing—original draft, writing—review and editing; C.D.: conceptualization, data curation, formal analysis, funding acquisition, investigation, methodology, project administration, resources, software, supervision, validation, visualization, writing—original draft, writing—review and editing.

Both authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

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References

- Koslow J, Goericke R, Lara-Lopez A, Watson W. 2011 Impact of declining intermediate-water oxygen on deepwater fishes in the California Current. *Mar. Ecol. Prog. Ser.* **436**, 207–218. (doi:10.3354/meps09270)
- Pörtner HO, Knust R. 2007 Climate change affects marine fishes through the oxygen limitation of thermal tolerance. *Science* **315**, 95–97. (doi:10.1126/science.1135471)
- Prince ED, Goodyear CP. 2006 Hypoxia-based habitat compression of tropical pelagic fishes. *Fish. Oceanogr.* **15**, 451–464. (doi:10.1111/j.1365-2419.2005.00393.x)
- Deutsch C, Ferrel A, Seibel B, Pörtner H-O, Huey RB. 2015 Climate change tightens a metabolic constraint on marine habitats. *Science* **348**, 1132–1135. (doi:10.1126/science.aaa1605)
- Rogers NJ, Urbina MA, Reardon EE, McKenzie DJ, Wilson RW. 2016 A new analysis of hypoxia tolerance in fishes using a database of critical oxygen level (P_{crit}). *Conserv. Physiol.* **4**, cow012. (doi:10.1093/conphys/cow012)
- Vaquero-Sunyer R, Duarte CM. 2008 Thresholds of hypoxia for marine biodiversity. *Proc. Natl Acad. Sci. USA* **105**, 15 452–15 457. (doi:10.1073/pnas.0803833105)
- Clarke A, Johnston NM. 1999 Scaling of metabolic rate with body mass and temperature in teleost fish. *J. Anim. Ecol.* **68**, 893–905. (doi:10.1046/j.1365-2656.1999.00337.x)
- Gillooly JF, Gomez JP, Mavrodiev EV, Rong Y, McLamore ES. 2016 Body mass scaling of passive oxygen diffusion in endotherms and ectotherms. *Proc. Natl Acad. Sci. USA* **113**, 5340–5345. (doi:10.1073/pnas.1519617113)
- Deutsch C, Penn JL, Seibel B. 2020 Metabolic trait diversity shapes marine biogeography. *Nature* **585**, 557–562. (doi:10.1038/s41586-020-2721-y)
- Cheung WWL, Sarmiento JL, Dunne J, Frölicher TL, Lam VWY, Deng Palomares ML, Watson R, Pauly D. 2013 Shrinking of fishes exacerbates impacts of global ocean changes on marine ecosystems. *Nat. Clim. Change* **3**, 254–258. (doi:10.1038/nclimate1691)
- Forster J, Hirst AG, Atkinson D. 2012 Warming-induced reductions in body size are greater in aquatic than terrestrial species. *Proc. Natl Acad. Sci. USA* **109**, 19 310–19 314. (doi:10.1073/pnas.1210460109)
- Horne CR, Hirst AG, Atkinson D. 2015 Temperature-size responses match latitudinal-size clines in arthropods, revealing critical differences between aquatic and terrestrial species. *Ecol. Lett.* **18**, 327–335. (doi:10.1111/ele.12413)
- Verberk WCEP, Atkinson D, Hoefnagel KN, Hirst AG, Horne CR, Siepel H. 2021 Shrinking body sizes in response to warming: explanations for the temperature–size rule with special emphasis on the role of oxygen. *Biol. Rev.* **96**, 247–268. (doi:10.1111/brv.12653)
- Deutsch C, Penn JL, Verberk WCEP, Inomura K, Endress M-GA, Payne JL. 2022 Impact of warming on aquatic body sizes explained by metabolic scaling from microbes to macrofauna. *Proc. Natl Acad. Sci. USA* **119**, 9. (doi:10.1073/pnas.2201345119)
- Chu JWF, Gale KSP. 2017 Ecophysiological limits to aerobic metabolism in hypoxia determine epibenthic distributions and energy sequestration in the northeast Pacific ocean: ecophysiological constraints in hypoxia. *Limnol. Oceanogr.* **62**, 59–74. (doi:10.1002/lno.10370)
- Duncan MI, James NC, Potts WM, Bates AE. 2020 Different drivers, common mechanism; the distribution of a reef fish is restricted by local-scale oxygen and temperature constraints on aerobic metabolism. *Conserv. Physiol.* **8**, coaa090. (doi:10.1093/conphys/coaa090)
- Franco AC, Kim H, Frenzel H, Deutsch C, Ianson D, Sumaila UR, Tortell PD. 2022 Impact of warming and deoxygenation on the habitat distribution of Pacific halibut in the northeast Pacific. *Fisheries Oceanogr.* **31**, 601–614. (doi:10.1111/fog.12610)
- Howard EM *et al.* 2020 Climate-driven aerobic habitat loss in the California Current System. *Sci. Adv.* **6**, eaay3188. (doi:10.1126/sciadv.aay3188)
- Clarke TM, Wabnitz CCC, Striegel S, Frölicher TL, Reygondeau G, Cheung WWL. 2021 Aerobic growth index (AGI): An index to understand the impacts of ocean warming and deoxygenation on global marine fisheries resources. *Prog. Oceanogr.* **195**, 102588. (doi:10.1016/j.pocean.2021.102588)
- Lucey NM, Deutsch CA, Carignan M-H, Vermandele F, Collins M, Johnson MD, Collin R, Calosi P. 2023 Climate warming erodes tropical reef habitat through frequency and intensity of episodic hypoxia. *PLoS Clim.* **2**, e0000095. (doi:10.1371/journal.pclm.0000095)
- Penn JL, Deutsch C, Payne JL, Sperling EA. 2018 Temperature-dependent hypoxia explains biogeography and severity of end-Permian marine mass extinction. *Science* **362**, aat1327. (doi:10.1126/science.aat1327)
- Breitburg D *et al.* 2018 Declining oxygen in the global ocean and coastal waters. *Science* **359**, eaam7240. (doi:10.1126/science.aam7240)
- Penn JL, Deutsch C. 2022 Avoiding ocean mass extinction from climate warming. *Science* **376**, 524–526. (doi:10.1126/science.abe9039)
- Appeltans W *et al.* 2012 The magnitude of global marine species diversity. *Curr. Biol.* **22**, 2189–2202. (doi:10.1016/j.cub.2012.09.036)
- Endress M-GA, Boag TH, Burford BP, Penn JL, Sperling EA, Deutsch CA. In press. Thermal optima in the hypoxia tolerance of marine ectotherms: physiological causes and biogeographic consequences. *PLoS Biol.*
- Slesinger E, Andres A, Young R, Seibel B, Saba V, Phelan B, Rosendale J, Wiczorek D, Saba G. 2019 The effect of ocean warming on black sea bass (*Centropristis striata*) aerobic scope and hypoxia tolerance. *PLoS ONE* **14**, e0218390. (doi:10.1371/journal.pone.0218390)
- Hammond KA, Diamond J. 1997 Maximal sustained energy budgets in humans and animals. *Nature* **386**, 457–462. (doi:10.1038/386457a0)
- Peterson CC, Nagy KA, Diamond J. 1990 Sustained metabolic scope. *Proc. Natl Acad. Sci. USA* **87**, 2324–2328. (doi:10.1073/pnas.87.6.2324)
- Nilsson GE, Östlund-Nilsson S. 2008 Does size matter for hypoxia tolerance in fish? *Biol. Rev.* **83**, 173–189. (doi:10.1111/j.1469-185X.2008.00038.x)
- Boag TH, Stockey RG, Elder LE, Hull PM, Sperling EA. 2018 Oxygen, temperature and the deep-marine stenothermal cradle of Ediacaran evolution. *Proc. R. Soc. B* **285**, 20181724. (doi:10.1098/rspb.2018.1724)
- Garcia H *et al.* 2018 *World ocean atlas 2018, volume 3: Dissolved oxygen, apparent oxygen utilization, and oxygen saturation*, (Technical ed. A. Mishonov), 38 pp. NOAA Atlas NESDIS 83.
- Locarnini RA *et al.* 2018 *World ocean atlas 2018, volume 1: Temperature* (Technical ed. A. Mishonov), 52 pp. NOAA Atlas NESDIS, 81.
- OBIS. 2022 Ocean biodiversity information system. Intergovernmental Oceanographic Commission of UNESCO (dataset). See www.obis.org.

34. Killen SS, Glazier DS, Rezende EL, Clark TD, Atkinson D, Willener AST, Halsey LG. 2016 Ecological influences and morphological correlates of resting and maximal metabolic rates across teleost fish species. *Am. Nat.* **187**, 592–606. (doi:10.1086/685893)
35. Bennett JM *et al.* 2021 The evolution of critical thermal limits of life on Earth. *Nat. Commun.* **12**, 1198. (doi:10.1038/s41467-021-21263-8)
36. Verberk WCEP, Sandker JF, van de Pol ILE, Urbina MA, Wilson RW, McKenzie DJ, Leiva FP. 2022 Body mass and cell size shape the tolerance of fishes to low oxygen in a temperature-dependent manner. *Glob. Change Biol.* **28**, 5695–5707. (doi:10.1111/gcb.16319)
37. OpenTree. *et al.* 2023 Open Tree of Life Synthetic Tree (version ott3.5) (dataset). (doi:10.5281/zenodo.3937741)
38. Grafen A. 1989 The phylogenetic regression. *Phil. Trans. R. Soc. Lond. B* **326**, 119–157. (doi:10.1098/rstb.1989.0106)
39. Chaudhary C, Richardson AJ, Schoeman DS, Costello MJ. 2021 Global warming is causing a more pronounced dip in marine species richness around the equator. *Proc. Natl Acad. Sci. USA* **118**, e2015094118. (doi:10.1073/pnas.2015094118)
40. Seibel BA, Deutsch C. 2020 Oxygen supply capacity in animals evolves to meet maximum demand at the current oxygen partial pressure regardless of size or temperature. *J. Exp. Biol.* **223**, jeb.210492. (doi:10.1242/jeb.210492)
41. Tseng Y-C, Hwang P-P. 2008 Some insights into energy metabolism for osmoregulation in fish. *Comp. Biochem. Physiol. C: Toxicol. Pharmacol.* **148**, 419–429. (doi:10.1016/j.cbpc.2008.04.009)
42. Mikheev VN, Pasternak AF, Valtonen E, Taskinen J. 2014 Increased ventilation by fish leads to a higher risk of parasitism. *Parasites Vectors* **7**, 281. (doi:10.1186/1756-3305-7-281)
43. Knoll AH, Bambach RK, Canfield DE, Grotzinger JP. 1996 Comparative earth history and Late Permian mass extinction. *Science* **273**, 452–457. (doi:10.1126/science.273.5274.452)
44. Gordó-Vilaseca C, Stephenson F, Coll M, Lavin C, Costello MJ. 2023 Three decades of increasing fish biodiversity across the northeast Atlantic and the Arctic Ocean. *Proc. Natl Acad. Sci. USA* **120**, e2120869120. (doi:10.1073/pnas.2120869120)
45. Beers JM, Sidell BD. 2011 Thermal tolerance of Antarctic notothenioid fishes correlates with level of circulating hemoglobin. *Physiol. Biochem. Zool.* **84**, 353–362. (doi:10.1086/660191)
46. Kwiatkowski L *et al.* 2020 Twenty-first century ocean warming, acidification, deoxygenation, and upper-ocean nutrient and primary production decline from CMIP6 model projections. *Biogeosciences* **17**, 3439–3470. (doi:10.5194/bg-17-3439-2020)
47. Penn JL, Deutsch C. 2023 Code for Geographical and taxonomic patterns in aerobic traits of marine ectotherms. GitHub repository. (https://github.com/jlpenn/ML_traits_obis)
48. Penn JL, Deutsch C. 2023 Geographical and taxonomic patterns in aerobic traits of marine ectotherms. Figshare. (doi:10.6084/m9.figshare.c.6978727)